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Overview of implantable and injectable biomaterials in immunotherapy

Prerana Prakash *, Pushpa Agrawal and AH Manjunatha Reddy

Department of Biotechnology RV College of Engineering, Bangalore-560-059, Karnataka, India.

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Abstract

Immunotherapy has shown promising applications in cancer treatment as it boosts the systemic immune response. Existing immunotherapy strategies have certain drawbacks which can be addressed by engineered biomaterials. In this review, we focused on advanced immunotherapy methods involving implantable and injectable biomaterials for the treatment of cancer. Engineered biomaterials as carriers for immunomodulatory agents aid in the local drug delivery, thus reducing the frequency of off-target side effects. Also, biomaterial-based cancer vaccines have the potential to target specific tissues by finely altering the physical properties of the drug to achieve desired drug release kinetics.

Keywords: Immunotherapy; Implantable biomaterial; Injectable biomaterial; Hydrogels; Colony stimulating factors (CSFs)

1. Introduction

The earliest definition of biomaterials was developed in 1972, during the Consensus Conference of the European Society for Biomaterials (ESB), during which biomaterial was defined as 'a nonviable material used in a medical device, intended to interact with biological systems; As per the current definition, it is a 'material intended to interface with biological systems to evaluate, treat, augment or replace any tissue, organ or function of the body [1,4]. This change in definition is suggestive of how the field of biomaterials has changed. Biomaterials have moved from merely interacting with the body to prompting biological processes toward the goal of tissue regeneration [1,4]

Tissue engineering is an interdisciplinary field that includes material science, genetics, clinical medicine, mechanical engineering [2]. It is the application of engineering and life sciences towards repair, restore and improve the function of tissue or a whole organ.

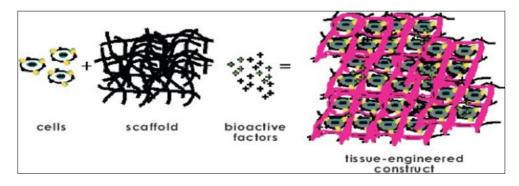


Figure 1 Main components of the tissue-engineered construct [1]

* Corresponding author: Prerana Prakash

Department of Biotechnology RV College of Engineering, Bangalore-560-059, Karnataka, India.

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This utilizes Scaffold for structural support and environment for cell proliferation, adhesion, and Extracellular matrix deposition until new tissue is completely restored [3]. Furthermore, the scaffolds are combined with growth factors, cells, and other signaling molecules that represent the key elements of tissue engineering (fig 1) [4]. After integration into host tissue, scaffold material should not elicit an immune response to avoid inflammatory reactions that might reduce the healing or result in rejection [5].

2. Biomaterials in immunotherapy

Recently, synthetic and natural biomaterials have become important elements in tissue engineering and regenerative medicine [4]. Several types of biomaterials have been currently used with key properties such as biocompatibility, biodegradability, bioactivity, based on applications these materials must be carefully designed and synthesized to fulfil their role. The use of biomaterials can elicit adverse reactions by the immune system, recognizing it as foreign origin, which interferes in the drug delivery system. Thus, it is noteworthy in understanding the interactions involved between biomaterials and the immune system [6].

In the case of implants, the foreign body response is a two-step process consisting of an inflammatory reaction followed by wound-healing [7] which triggers the accumulation of a thick layer of collagen on the implant. This deteriorates the efficacy of implanted material [7, 8]

The potential applications of biomaterials involving synthetic nanoparticles and microparticles, lipid carriers, injectable scaffolds, and hydrogels are directed to the field of immunotherapy. These materials can be designed not only to release immunomodulatory factors but also to navigate the host immune response [9-11].

The immune system plays a vital role in the prevention of tumor development and cancer [13-15]. Cancer immunotherapy invokes the activity of the intrinsic immune system to fight cancer cells [16-20]. Immunotherapy can be defined as the treatment that uses certain substances to suppress or stimulate the immune system to fight against cancer, infection, or other diseases. This recent development in the treatment of cancer has led to the eradication of systemic tumor, in contrast to traditional chemotherapies which uses cytotoxic drugs to act against cancer cells [12]. As yet, there are three main types of cancer immunotherapy which include, cancer vaccines [21-23], immune checkpoint blockades [24-26], chimeric antigen receptor-modified T cells [27]. These immunotherapeutics have shown excellent success in treating some tumor types but still, some challenges persist.

Engineered biomaterials that are injectable, implantable, and transdermal devices (fig 2) have paved the way in the development of localized delivery systems for immune therapeutics [28-35]. Advanced biomaterials are designed and developed to optimize the pharmacokinetics of the drug, controlled drug release, and increased drug retention within the target site while reducing immunotoxicity.

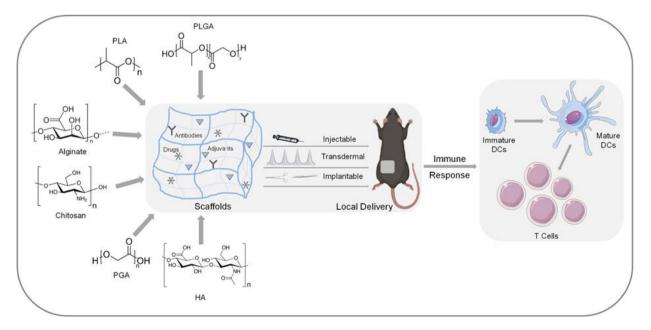


Figure 2 Representation of engineered biomaterials in cancer immunotherapeutic [36].

3. Implantable biomaterials

Implantable biomaterials are implanted subcutaneously or at the site of excised tissue via surgery. Before implantation, biomaterials are preloaded with various chemical substances, biologic factors, or bioactive agents. These bioactive agents are released in a controlled fashion from a porous scaffold matrix, and immune cells are recruited at the site [37, 38]. Engineered biomaterials such as polyglycolide (PGA), chitosan, hyaluronic acid (HA), polylactide (PLA), alginate, and poly (lactic-co-glycolic acid) (PLGA), are utilized to create porous scaffolds that encapsulate therapeutic substances in injectable, implantable, or transdermal delivery systems [fig 2]. Further, immune responses such as the proliferation of T-cells and maturation of dendritic cells are activated by localized drug delivery [36].

Antigen-presenting cells (APCs) such as dendritic cells play a major role in initiating humoral and cellular immune responses by presenting tumor-associated antigens in different ways. This mechanism has been studied in mouse cancer models, wherein dendritic cells capture antigens from dead or live tumor cells and present the antigens to T cells in tumor-draining lymph nodes. Further, this resulted in the production of tumor-specific cytotoxic T lymphocytes (CTLs) [39].

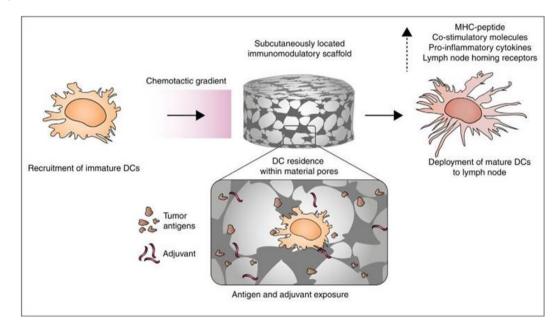


Figure 3 Implantable porous scaffold designed to release chemoattractants in order to recruit immature dendritic cells [39]

Several types of biodegradable polymers such as PLG are used in implantable drug-delivery scaffolds. PLG is extensively used in cancer immunotherapy due to its ease of surface modification, customizable biodegradation rate, high biocompatibility, and FDA approval for clinical use. A novel method was devised to prepare uniform PLG scaffolds of molecular mass 120kD in 85:15 ratio of copolymer D,L-lactide, and glycolide using a gas-foaming process [40]. PLG scaffolds are used to encapsulate hematopoietic cytokine, granulocyte-macrophage colony-stimulating factor (GM-CSF) that induces expansion of DCs, macrophages, and neutrophils. It also inhibits the proliferation of cancer cells via immune-independent effects and regulates T-cell proliferation and activation. GM-CSF attracts DCs that engulf apoptotic tumor cells. Release of GM-CSF from the scaffold induces DC maturation. The mature DCs then migrate to draining lymph nodes and present tumor antigen to T cells. This results in T-cell activation and expansion, enhancing the antitumor immune response (fig 3) [39].

On contrary to traditional cancer vaccines, an implantable GM-CSF delivery system can release GM-CSF continuously for 2 weeks, resulting in a longer immune response [41]. In a melanoma mouse model, the survival rate increased significantly after implanting PLG scaffold encapsulated with tumor-associated antigens, and GM-CSF.

Implantable delivery systems with modified biomaterial scaffolds have several advantages due to their ability to control the release of immune cell recruitment factors, continuous release of bioactive agents for a sustained period, and various other functions. One of the drawbacks of the implantable delivery system is the requirement of surgery, this can become a potential source of infection [36].

4. Injectable biomaterial

Implantable biomaterials have certain limitations that have led to the development of injectable immunotherapeutic biomaterials. Injectable biomaterials include cryogels, hydrogels, and other *in-situ* systems derived from synthetic or natural sources [12]. These materials enable controlled and localized release of the drug (fig 4). Injectable material can be modified for easy move and flow across the membrane, thus conforming to any accessible site [42]. An injectable therapeutic system avoids any tissue damage and complications associated with an inflammatory wound response. It is a less invasive procedure compared to surgical implantation.

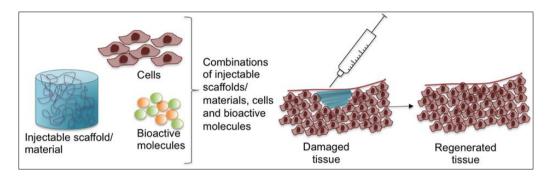


Figure 4 Injectable biomaterial in tissue regeneration [46]

In the recent research carried out by Bencherif et al., an alginate-based cryogel biomaterial was studied as an injectable cancer vaccine using CpG as a danger signal, GM-CSF for cell recruitment, and irradiated melanoma cells as antigen source. RGD peptides were incorporated into the cryogel for better adhesion of irradiated melanoma cells. Thus, resulted in sustained release of the immunotherapeutic factors [43,44].

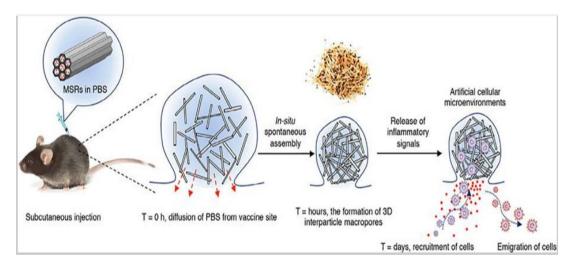


Figure 5 illustration of MSR assembly and cell recruitment in vivo [48]

Injectable hydrogels are currently applied for designing cancer vaccines. An injectable smart hydrogel was designed similar to extracellular matrix that self assembles into the microporous network, and by using simple hypodermic needles it can be injected subcutaneously. Scaffold for encapsulating GM-CSF and ovalbumin has been constructed with a temperature-responsive copolymer of PCLA and PEG combined with HA. A single injection of this loaded hydrogel into B16/OVA mice enhanced the recruitment of DCs and other immune cells to the tumor site and led to substantial inhibition of tumor growth [45].

Injectable hydrogels have shown potential tumor growth inhibition and prevention of recurrence in combination therapy in vivo. Fibrin hydrogel possesses excellent biodegradability and biocompatibility. It was loaded with chemotherapeutics cyclophosphamide (CTX; a small molecule drug) and anti-PD-L1 (an immune checkpoint-blocking monoclonal antibody). The drug release kinetics of CTX and anti-PD-L1 vary widely due to their large size difference. CTX being the smaller molecule is released first from the fibrin gel, thus creating an immunogenic tumor microenvironment. This generates a synergistic antitumor effect due to staggered delivery of the immune checking-

blocking antibody and small molecule from the fibrin gel reservoir. Further, it was evaluated in mouse models of ID8 ovarian cancer and TNBC 4T1 breast cancer, resulting in increased inhibition rate of tumor recurrence after surgery [47].

Kim et al. in 2014 developed a strategy for injectable cancer vaccine scaffold based on mesoporous silica rods (MSRs) [48]. It is a well-designed compromise between implantable scaffolds and injectable biomaterial that self-assembles into a separate pocket-like depot when injected subcutaneously (fig 5). When the scaffold is pre-loaded with CpG, GM-CSF, and other molecules, it creates a depot of immunotherapeutic agents that is capable to recruit and influence local immune cells [48].

A recent development in injectable biomaterials involved the application of hydrogel spray onto the surface of the tumor site after the surgery, to inhibit the tumor regrowth by increasing the pH of the tumor environment. This spray consists of FDA-approved fibrin gel to encapsulate CaCO₃ nanoparticles loaded with anti-CD47 antibody. CaCO3 nanoparticles serve as a reservoir for releasing immunomodulatory antibody as well as a proton scavenger that reduces the pH of tumor environment. When it is sprayed onto the tumor site post-surgery, the CaCO3 nanoparticles dissolve and release the encapsulated anti-CD47 antibodies which activates the macrophages to engulf cancer cells and inhibit tumor recurrence at the site [36, 49].

5. Conclusion

Current developments in immunotherapy and biomaterials have paved the way for a promising approach towards overcoming clinical limitations in the field of cancer immunotherapy. These advancements were depicted in this review showcasing the exciting potential of engineered biomaterials in drug delivery systems, whether it is in the form of implantable or injectable. Over the past few decades, Immunotherapy strategies have shown promising clinical results. Yet they possess certain limitations such as systemic side effects, poor delivery kinetics, and narrow patient response profiles. Engineered biomaterials overcome these problems, allowing tunable drug release kinetics up to several days or months depending on the design of biomaterials, improved efficacy, and reduced side effects.

Compliance with ethical standards

Acknowledgments

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Disclosure of conflict of interest

There are no conflicts of interest to disclose.

References

- [1] Fergal J. O'Brien. Biomaterials & scaffolds for tissue engineering. Materials Today. 2011; 14(3): 88-95.
- [2] Langer R, Vacanti JP. Tissue engineering. Science. 1993; 260(5110): 920-926.
- [3] Salgado AJ, Coutinho OP, Rui L, Reis RL. Bone tissue engineering: State of the art and future trends. Macromolecular Bioscience. 2004; 4: 743-765.
- [4] A Dolcimascolo, G Calabrese, S. Conoci, R Parenti. Innovative Biomaterials for Tissue Engineering. Biomaterialsupported Tissue Reconstruction or Regeneration. 2019.
- [5] Hutmacher DW. Scaffolds in tissue engineering bone and cartilage. Biomaterials, 2000; 175-189.
- [6] Kowalski P, Bhattacharya C, Afewerki S, Langer R. Smart Biomaterials: Recent Advances and Future Directions. ACS Biomaterials Science & Engineering. 2018; 4(11): 3809-3817.
- [7] Anderson J. M, Rodriguez A, Chang D. T. Foreign Body Reaction to Biomaterials. Semin. Immunol. 2008; 20 (2): 86–100.
- [8] Wick G, Grundtman C, Mayerl C, Wimpissinger T.F, Feichtinger J, Zelger B, Sgonc R. Wolfram D. The Immunology of Fibrosis. 2013; 31.

- [9] Singh A, Peppas NA, Hydrogels and scaffolds for immunomodulation. Adv Mater. Oct 2014; 26(38): 6530-41.
- [10] Hotaling NA, Tang L, Irvine DJ, Babensee JE; Biomaterial Strategies for Immunomodulation. Annu Rev Biomed Eng. 2015; 17: 317-49.
- [11] Mehta NK, Moynihan KD, Irvine DJ. Engineering New Approaches to Cancer Vaccines. Cancer Immunol Res. Aug 2015; 3(8): 836-43.
- [12] Leach, D. G., Young, S., & Hartgerink, J. D. Advances in immunotherapy delivery from implantable and injectable biomaterials. Acta biomaterialia. 2019; 88: 15–31.
- [13] Carr EJ, Dooley J, Garcia-Perez JE. The cellular composition of the human immune system is shaped by age and cohabitation. Nat Immunol. 2016; 17(4): 461-468.
- [14] Jones JD, Vance RE, Dangl JL. Intracellular innate immune surveillance devices in plants and animals. Science. 2016; 354(6316).
- [15] Irvine DJ. Materializing the future of vaccines and immunotherapy. Nat Rev Mater. 2016; 1(1): 1-2.
- [16] Palucka AK, Coussens LM. The basis of oncoimmunology. Cell. 2016; 164(6): 1233-1247
- [17] Papaioannou NE, Beniata OV, Vitsos P, et al. Harnessing the immune system to improve cancer therapy. Ann Transl Med. 2016; 4(14): 241.
- [18] Ribas A. releasing the brakes on cancer immunotherapy. N Engl J Med. 2015; 373(16): 1490- 1492.
- [19] Cousin-Frankel J; cancer immunotherapy; Science. 2013; 342(6165): 1432-1433.
- [20] [20] Klevorn LE, Teague RM. Adapting cancer immunotherapy models for the real world. Trends Immunol. 2016; 37(6): 354- 363.
- [21] Fesnak AD, June CH, Levine BL. Engineered T cells: the promise and challenges of cancer immunotherapy. Nat Rev Cancer. 2016; 16(9): 566-581.
- [22] Goldberg MS. Immunoengineering: how nanotechnology can enhance cancer immunotherapy. Cell. 2015; 161(2): 201-204.
- [23] Ali OA, Verbeke C, Johnson C, et al. Identification of immune factors regulating antitumor immunity using polymeric vaccines with multiple adjuvants. Cancer Res. 2014; 74(6): 1670-1681.
- [24] Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. Science. 2018; 359(6382): 1350- 1355.
- [25] Moynihan KD, Opel CF, Szeto GL, et al. Eradication of large established tumors in mice by combination immunotherapy that engages innate and adaptive immune responses. Nat Med. 2016; 22(12): 1402- 1410.
- [26] Ishihara J, Fukunaga K, Ishihara A, et al. Matrix-binding checkpoint immunotherapies enhance antitumor efficacy and reduce adverse events. Sci Transl Med. 2017; 9(415): eaan 0401.
- [27] Curran KJ, Silverman LB, Kobos R, et al. Chimeric antigen receptor T cells for cancer immunotherapy. J Clin Oncol. 2015; 33(15): 1703- 1706.
- [28] Wang C, Ye Y, Hochu GM, et al. Enhanced cancer immunotherapy by microneedle patch-assisted delivery of anti-PD1 antibody. Nano Lett. 2016; 16(4): 2334- 2340.
- [29] Tsao CT, Kievit FM, Ravanpay A, et al. Thermoreversible poly (ethylene glycol)-g-chitosan hydrogel as a therapeutic T lymphocyte depot for localized glioblastoma immunotherapy. Biomacromolecules. 2014; 15(7): 2656-2662.
- [30] Kelly SH, Shores LS, Votaw NL, Collier JH. Biomaterial strategies for generating therapeutic immune responses. Adv Drug Deliv Rev. 2017; 114: 3- 18.
- [31] Vishwakarma A, Bhise NS, Evangelista MB, et al. engineering immunomodulatory biomaterials to tune the inflammatory response. Trends Biotechnol. 2016; 34(6): 470- 482.
- [32] Hu X, Wu T, Bao Y, Zhang ZJ. Nanotechnology based therapeutic modality to boost anti-tumor immunity and collapse tumor defense. J Control Release. 2017; 256: 26- 45.
- [33] Shi J, Kantoff PW, Wooster R, Farokhzad OC. Cancer nanomedicine: progress, challenges and opportunities. Nat Rev Cancer. 2017; 17(1): 20- 37.

- [34] Wang C, Wen D, Gu Z. Cellular bioparticulates with therapeutics for cancer immunotherapy. Bioconjugate Chem. 2017; 29(3): 702-708.
- [35] Iang W, Von Roemeling CA, Chen Y, et al. Designing nanomedicine for immuno-oncology. Nat Biomed Eng. 2017; 1(2): 1-11.
- [36] Cai L, Xu J, Yang Z, Tong R, Dong Z, Wang C, & Leong K. W. Engineered biomaterials for cancer immunotherapy. MedComm. 2020.
- [37] Leach DG, Young S, Hartgerink JD. Advances in immunotherapy delivery from implantable and injectable biomaterials. Acta Biomater. 2019; 88: 15- 31.
- [38] Koshy ST, Mooney DJ. Biomaterials for enhancing anti-cancer immunity. Curr Opin Biotechnol. 2016; 40: 1-8.
- [39] Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. Nat Rev Cancer. 2012; 12(4): 265-277.
- [40] Ali OA, Huebsch N, Cao L, et al. Infection-mimicking materials to program dendritic cells in situ. Nat Mater. 2009; 8(2): 151-158.
- [41] Zhang C, Zhang J, Shi G, et al. A light responsive nanoparticle-based delivery system using pheophorbide a graft polyethylenimine for dendritic cell-based cancer immunotherapy. Mol. Pharmaceutics. 2017; 14(5): 1760-1770.
- [42] Myron S, Teck Chuan L, Injectable biomaterials: a perspective on the next wave of injectable therapeutics, Biomed. Mater. 2016; 1(1): 014110.
- [43] Bencherif SA, Sands RW, Bhatta D, Arany P, Verbeke CS, Edwards DA, Mooney DJ. Injectable preformed scaffolds with shape-memory properties. 2012; 109(48): 19590–19595.
- [44] Bencherif SA, Sands RW, Ali OA, Li WA, Lewin SA, Braschler TM, Shih T-YS, Verbeke CS, Bhatta D, Dranoff G, Mooney DJ. Injectable cryogel-based whole cell cancer vaccines. Nat. Commun. 2015; 6: 7556
- [45] Duong HTT, Thambi T, Yin Y, et al. Degradation-regulated architecture of injectable smart hydrogels enhances humoral immune response and potentiates antitumor activity in human lung carcinoma. Biomaterials. 2020; 230: 119599.
- [46] Ercan H, Durkut S, Koc-Demir A, Elçin AE, Elçin YM. Clinical Applications of Injectable Biomaterials. Adv Exp Med Biol. 2018; 1077: 163-182.
- [47] Zhang L, Zhou J, Hu L, et al. In situ formed fibrin scaffold with cyclophosphamide to synergize with immune checkpoint blockade for inhibition of cancer recurrence after surgery. Adv Funct Mater. 2020; 30(7): 1906922.
- [48] Kim J, Li WA, Choi Y, Lewin SA, Verbeke CS, Dranoff G, Mooney DJ, Injectable, spontaneously assembling, inorganic scaffolds modulate immune cells in vivo and increase vaccine efficacy, Nat. Biotechnol. 2014; 33(64).
- [49] Chen Q, Wang C, Zhang X, et al. In situ sprayed bioresponsive immunotherapeutic gel for post-surgical cancer treatment. Nat Nanotechnol. 2019; 14(1): 89-97.