

GSC Advanced Research and Reviews

eISSN: 2582-4597 CODEN (USA): GARRC2 Cross Ref DOI: 10.30574/gscarr Journal homepage: https://gsconlinepress.com/journals/gscarr/

(RESEARCH ARTICLE)

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Gas chromatography-Mass spectrometry (GC-MS) analysis of bioactive components present in grape citrus peel in Nigeria

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GSC Advanced Research and Reviews, 2021, 08(01), 166-174

Publication history: Received on 13 June 2021; revised on 19 July 2021; accepted on 21 July 2021

Article DOI: https://doi.org/10.30574/gscarr.2021.8.1.0152

Abstract

Grapefruit (*Citrus paradisi*) is an important cultivar of the Citrus genus which contains a number of nutrients beneficial to human health. Grapefruit peels are usually thrown away in this part of Nigeria. The research work investigated the bioactive components present in an indigenous citrus peel, grape (*Citrus paradisi*). Grape fruits were purchased from the fruits garden market in D-Line, Port-Harcourt metropolis and washed with ionized water and allowed to shade dry. The peel of the fruits were separated and subjected to cold extraction using 95% ethanol. The extracts obtained were further extracted in dichloromethane and subjected to GC/MS analysis for characterization of various bioactive components. The gas chromatographic model: 789A (GC) analysis was performed on an agilent technologies interfaced with mass selective detector model: 5975(MSD). The results revealed 25 bioactive components in grape peel with n-Hexadecanoic acid showing the highest concentration of 20.36% and retention time of 18.522min. Nootkatone was the lowest component in the grape peel with concentration 0.74% and retention time of 16.459min. Results shows that grapefruit (*Citrus paradisi*) has considerable potential as a source of natural bioactive components with different retention times. These fruits residues which otherwise regarded as waste hold promising potentials for medicinal therapy and value added food supplements.

Keywords: Citrus peels; Bioactive; Gas chromatography; Mass spectrometry

1. Introduction

Citrus is a genus of flowering trees and shrubs in the rue family, Rutaceae which include fruits such as orange, lime, lemon, grapefruits, tangerine appear as a well-known promising source of multiple beneficial nutrients for human beings. Citrus fruits are one of the largest fruit crops in the world. These citrus fruits have a well-inscribed nutritional value, along with high levels of elemental bioactive compounds such as essential oils (EOs), flavonoids, limonoids and vitamins. Essential oils are a good source of several bioactive compounds, which possess antioxidative and antimicrobial properties. Essential oils are concentrated liquids of complex mixtures of volatile compounds and can be extracted from several plant organs. Citrus (Citrus L. from Rutaceae) is one of the most popular world fruit crops, contains active phytochemicals that can protect health. In addition to this, it provides an ample supply of vitamin C, folic acid, potassium and pectin. The contribution of citrus species in deterrence of life threatening diseases have been assessed [1] [2] [3] [4] and it has been reported that citrus fruits and citrus fruits extracts and citrus flavonoids exhibits a wide range of promising biological properties due to their phenolic profile and antioxidant properties [5] [6] [7] Global production of citrus fruits has significantly increased during the past few years and has reached 82 million tons in the years 2009-2010 of which oranges- commercially most important citrus fruits has accounts for about 50 million tons and 34% of which was used for juice production, yielding about 44% peel by-product [8] Therefore, a large amount of peels is produced every year. Citrus peel, the primary waste, is a good source of molasses, pectin and limonene and is usually

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dried, mixed with pulps and sold as cattle feed [9]. Uses of essential oils have received increased attention as natural additives for the shelf-life extension of food products, due to risk in using synthetic preservatives. The objective of this research is to investigate the bioactive components, their percentages, retention time, molecular weights and structures of the dichloromethane extract of grape citrus peel.

2. Material and methods

2.1. Preparation Ethanol Extract

A large quantity of grape fruits was procured from markets within Port Harcourt metropolis, Rivers State, Nigeria. The citrus fruit was washed with ionized water, dried up and peeled, after which the peels were blended using homemade grinder/blender into powder form. The peels were then weighed using a Radwag-Wagi electronic top loading balance (AS 220/C/2) to ensure uniformity in weight. The peels were then pureed using MarlexElectroline (Excella) blender. Two hundred grams (200g) of the milled sample was weighed and soaked in 100ml of 95% ethanol for 48 hours after which they were sieved using a muslin cloth and filtered with Whatman filter paper size 1. The filtrate was concentrated using rotary evaporator at 45°C the weight of the concentrates were taken and the percentage yield calculated and kept at 4°C until usage.

2.2. Determination of Essential Oils Content

The milled sample was extracted in dichloromethane after soaking for 5 days. Ten grams of the sample was weighed into a well dried stopper bottle and 20mls of the organic solvent was added. The mixtures were vigorously agitated and were left to stand for 5 days. The crude extract was collected by fitting into a quartz beaker, the process were repeatedly carried out for two consecutive times. The combined aliquot collected was concentrated on a steam bath to about 5ml and purified by passing through a pasture pipette packed with silica gel and anhydrous sodium sulphate on a membrane and air dried to about 2ml for gas chromatographic analysis.

The extract of the sample was subjected to GC/MS analysis, this group of powerful instruments interfaced helped to characterize the various compositions. The gas chromatographic Model: 7890A (GC) analysis was performed on an Agilent Technologies interfaced with Mass Selective Detector model: 5975C (MSD). The electron ionization was at a 70v with an iron source temperature at250°C highly pure helium gas (99.9% purity) was used as carrier gas, while HP-5 (30mm X 0.25mm X 0.320 μ m) was used as the stationary phase. The oven temperature was at at 60°C held for 0.5 minute and ramped to at 140°C at the rate of 4°C/minutes holding for a minute, then ramped to 280 degrees while holding for 5 minutes at the rate of at 8°C /minutes. 1 μ /l was auto injected.

3. Results

The results of the essential oils and bioactive components of the dichloromethane extract of grape citrus peels analyzed are shown in Figure 1 and Table 1 respectively. The chromatogram shows the various peaks while the tables consist of the bioactive components, their retention time, percentages, molecular weights and structures.

3.1. GC/MS and phytochemical analysis of dichloromethane extract of grape peel

Figure 1 shows the chromatogram of GC/MS of bioactive compounds of dichloromethane extract grape peel with highest peak observed at a retention time of 18.522min followed by retention time of 20.630min, 21.755min and 27.606min with the lowest peak observed at a retention time of 4.220min. Table 1 shows the presence of 25 active bioactive components in the dichloromethane extract of grape peels. N-Hexadecanoic acid was the highest in concentration (20.360% and retention time of 18.522min). This was followed by 9,12,-Octadecadienoic acid(Z,Z)-(11.710% and retention time of 20.573min), Isoauraptene(9.28% and retention time of 21.758min) and 2H-1-Benzpyrane-2-one, 7-[(3,7,-dimethyl-2,6,-octadienyl)oxy]-,(E)- (7.990% and retention time of 27.606min) respectively. Nootkatoone concentration was the lowest value of 0.74% at a retention time of 16.459min.



Figure 1 GC/MS Chromatogram of Grape Peel

Table 1 Bioactive Components of Grape Peel (Citrus paradisi)

S/N	Compound	Retention Time (min)	Percentage of the total	Molecular formula	Molecular weight	Structure
1	D-Limonene	4.220	3.64	C10H16	136.24	104 90 155 155 157 157 157 157 157 157
2	1,2-Cyclohexanediol, 1- methyl-4- (methylethenyl)-	9.307	3.40	C10H18O2	170.2487	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

3	Caryophyllene	10.697	1.28	C15H24	204.36	100- 41 69 79 105 103 50- 39 55 67 77 81 105 147 161 10 20 30 40 50 60 70 80 90 100 10 120 147 161 175 180 204 10 20 30 40 50 60 70 80 90 100 10 120 130 140 150 150 150 206 210 (maintb) Caryophyline
4	(E)betaFamesene	12.047	1.04	C15H24	204.357	$\begin{array}{c} & & & & & \\ & & & & \\ & & & & & \\ & & & & & \\ & & &$
5	1-Isopropyl-4,7- dimethyl-1,2,3,5,6 ,8a- hexahydronaphthalene	12.316	0.94	C15H24	204.351	$\begin{array}{c} & & & & & & & & & & & & & & & & & & &$
6	Tetradecanoic acid	15.721	0.82	C14H28O2	228.3709	100- 50- 29 43 55 69 83 97 15 129 04 129 04 155 143 19 19 19 19 19 20 20 20 20 20 20 20 20 20 20
7.	Nootkatone	16.459	0.74	C15H22O	218.34	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

8	Hexadecanoic acid, methyl ester	17.878	0.81	C17H34O2	270.4507	100- 50- 51- 52- 52- 15- 23- 15- 15- 15- 15- 15- 15- 15- 15
9	n-Hexadecanoic acid	18.522	20.36	$C_{16}H_{32}O_2$	256.4241	100 50 27 28 27 28 29 27 28 29 27 29 20 20 20 20 20 20 20 20 20 20
10.	Benzeneethanol, 4- hydroxy-	18.857	1.82	C8H10O2	138.1638	107 50- 107 107 107 107 10 10 10 10 10 10 10 10 10 10
11.	Tritetracontane	19.600	4.53	C43H88	605.177	10 57 54 57 54 57 57 57 57 57 57 57 57 57 57
12.	cis-13-Octadecenoic acid, methyl ester	20.035	0.81	C ₁₉ H ₃₆ O ₂	296.4879	100 55 64 15 59 15 59 15 59 15 59 15 59 15 59 15 59 15 59 15 59 15 59 15 59 15 15 15 15 15 15 15 15 15 15
13.	9,12-Octadecadienoic acid (Z,Z)-	20.573	11.71	C18H32O2	280.4455	100- 50- 50- 50- 50- 50- 50- 50-

14.	9,12-Octadecadienoic acid (Z,Z)-	20.630	7.98	C18H32O2	280.4455	100- 50- 50- 50- 50- 50- 50- 50-
15.	Octadecanoic acid	20.831	3.59	C ₁₈ H ₃₆ O ₂	284.4772	100- 50- 50- 50- 50- 50- 50- 50-
16.	Isoauraptene	21.758	9.28	C19H22O3	298.38	100- 50- 43- 45- 45- 45- 45- 45- 45- 45- 45
17.	Butyl 9,12,15- octadecatrienoate	21.987	1.81	C22H38O2	334.5433	100 100 100 100 100 100 100 100
18.	Auraptenol	22.250	1.20	C19H22O3	298.38	$100 - \frac{100}{50} - \frac{39}{51} + \frac{51}{50} + \frac{77}{7} + \frac{89}{10} + \frac{103}{110} + \frac{118}{10} + \frac{147}{15} + \frac{151}{10} + \frac{175}{10} + \frac{219}{200 210 220 230 240 250 280 270} + \frac{131}{100 100 100 100 100 100 100 100 100 100$
19.	1-Hexadecanethiol	22.507	1.95	C ₁₆ H ₃₄ S	258.506	100- 100-

20.	Heptadecane	23.560	2.99	C17H36	240.4677	102- 102- 102- 105
21.	8-(2,3-Dihydroxy-3- methylbutyl)-7- methoxy-2H-chromen- 2-one	24.155	4.29	C15H20O6	296.319	100- 50- 28 31 43 51 ⁵⁰ 55 71 77 91 103 115 122 147 180 1 20 30 40 50 60 70 80 90 100 110 120 130 140 150 160 170 180 190 200 210 220 230 240 250 260 270 280 290 (martic) 82,320/hydroy-3rethyloch/7-rethyro2H-choren-2-ore
22.	Bis(2-ethylhexyl) phthalate	25.986	1.74	C24H38O4	390.56	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
23.	2-Hexenal, 2-methyl-	26.444	2.49	C7H12O	112.170	100 - 41 - 55 - 0 - 7 - 7 - 83 - 97 - 112 - 11
24.	2H-1-Benzopyran-2- one, 7-[(3,7-dimethyl- 2,6-octadienyl)oxy]-, (E)-	27.606	7.99	C19H22O3	298.376	100- 50- 51- 52- 53- 54- 53- 54- 53- 54- 53- 54- 53- 54- 55- 54- 55- 55- 55- 55- 55
25.	Hexacosane	27.829	2.79	C26H54	366.718	57 59 50 50 50 50 50 50 50 50 50 50

4. Discussion

Every part of citrus family, starting from the leaves, the root, the back of the tree, the fruits juice etc which the peels are not exclusive has their functions. The fruits peel; grape (*Citrus paradise*), which was investigated for its essential oils. Grape peels revealed a number of 25 bioactive compounds with n-Hexadecanoic acid having the highest concentration of 20.36% and a retention time of 18.522min. The results agrees with at [10] which states that citrus peels contain significant amounts of biologically active polyphenols, specifically phenolic acids and flavonoids, which have exhibited anti-viral. important antioxidants, anti-inflammatory, anti-proliferative, anti-allergic, anti-carcinogenic. neuroprotective and antimicrobial properties. The presence of D-Limonene in grape peel agrees with [11] which state that Limonene is the most abundant compound of monoterpene hydrocabons for all of the examined juices, and [12] which states that experimental studies have demonstrated its analgesic, antibacterial, antiviral, hepatoprotective, antiatherogenic, radical scavenging activity, uricosuric, hypothensive agent, carminative and antilarvicidal. N-Hexadecanoic acid with the highest percentage in grape has its therapeutic function such as: inhibitory activity and anti-inflammatory agent [13]. N-hexadecanoic acid shows inhibitory activity against mycobacterium tuberculosis [14]. 9.12. Octadecadienoic acid(Z,Z)- acid which is also one of the bioactive component in grape peel has its own benefits: 9.12.-Octadecadienoic acid(Z,Z)-, acid has antiarthritic and anti-inflammatory property [15]. [16]. stated that 9,12,-Octadecadienoic acid(Z,Z)- has functions such as Hypocholesterolemic, Anticoronary, Nematicide, Hepatoprotective, Hypocholesterolemic etc. These bioactive components and others have a lot of medicinal and synthetic values.

5. Conclusion

Recent research concerning essential oils of citrus peels has added to our knowledge. Due to the low cost and easy availability of fruit residues which are commonly discarded as waste in our immediate environments should be regarded as potential medicinal and synthetic resources, capable of offering significant low-cost, medicinal and dietary supplements. In addition, an established use of bioactive components from grape citrus peel would also help alleviate environmental pollution problems caused due to poor disposal of such residues. This will go a long way to enhance the health stability and reduce environmental pollution.

Compliance with ethical standards

We confirm that we complied to all ethical standard that concerns with publishing an article.

Acknowledgments

I acknowledge everybody who, in one way or another contributed to making this works a success.

Disclosure of conflict of interest

The authors declared that there is no conflict of interest regarding the publication of this paper.

References

- [1] Proteggente A, Saija RA, De Pasquale A, Rice-Evans CA. The compositional characterisation and antioxidant activity of fresh juices from Sicilian sweet orange (Citrus sinensis L. Osbeck) varieties. Free Radic. Res. 2003; 37: 681-687.
- [2] Gorinstein S, Cvikrova M, Machackova I, Haruenkit R, Park YS, Jung ST, Yamamoto K, Ayala ALM, Katrich E, Trakhtenberg S. Characterization of antioxidant compounds in Jaffa sweeties and white grapefruits. Food Chemistry. 2004; 84: 503-510
- [3] Anagnostopoulou P, Kefalas VP, Papageorgiou AN, Assimopoulou DB. Radical scavenging activity and fractions of sweet orange flavedo (Citrus sinensis). Food Chemistry. 2006; 94: 19-25.
- [4] Guimarães R, Barros L, Barreira JCM, Sousa MJ, Carvalho AM, Ferreira ICFR. Targeting excessive free radicals with peels and juices of citrus fruits: grapefruit, lemon, lime and orange. Food Chemistry Toxicology. 2009; 48(1): 99 106.
- [5] Middleton Jr. E, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. Pharmacol. Rev. 1994; 52: 673-651.

- [6] Montanari A, Chen J, Widmer W. Citrus flavonoids: a review of past biological activity against disease. In: Manthey, J.A., Buslig, B.S. (Eds.), Flavonoids in the Living System. Plenum Press, New York. 1998; 103 113.
- [7] Samman S, Lyons Wall PM, Cook NC. Flavonoids and coronary heart disease: dietary perspectives. In: Rice-Evans CA, Packer L. (Eds.), Flavonoids in Health and Disease. Marcel Dekker, New York. 1998; 469–482.
- [8] Li JM, Che CT, Lau CB, Leung PS, Cheng CHK. Inhibition of intestinal and renal Na+-glucose cotransporter by naringenin. Int. J. Biochem. Cell Biol. 2006; 38: 985–995.
- [9] Bucco A, Cuvelier ME, Richard H, Berset C. Antioxidant activity and phenolic composition of citrus peel and seed extracts. Journal of agricultural and food chemistry. 1998; 46(6): 2123 2129.
- [10] Oboh G, Ademosun A. Characterisation of the antioxidant properties of phenolic extracts from citrus peels. Journal of Food Science and Technology. 2012; 49(6): 729-736.
- [11] Saidani M, Brahim M. Biochemical characterization of blood orange, sweet orange, lemon, bergamot and bitter orange. 2003; 62(8): 1283-1289.
- [12] Kanaze FI, Termentzi A, Gabrieli C, Niopas I, Georgarakis M, Kokkalou E. The phytochemical analysis and antioxidant activity assessment of orange peel Citrus sinensis (L) Osbeck cultivated in Greece-Crete indicates a new commercial source of hesperidin. Biomed Chromato gr. 2009; 23(3): 239-249.
- [13] Aparna V, Dileep KV, Mandal PK, Karthe P, Sadasivan C, Haridas M. Anti-inflammatory property of n-hexadecanoic acid: Structural evidence and kinetic assessment. Chemical biology & drug design. 2012; 80(3): 434-439
- [14] Suresh M, Panneerselvam A, Dhanasekaran D, Thajuddin N. Anti-mycobacterial effect of leaf extract of Centellaasiatica (Mackinlayaceae). Research Journal of Pharmacy and Technology. 2010; 3(3): 872-876.
- [15] Lalitharani S, Mohan VR, Regini GS, Kalidass C. GC-MS analysis of ethanolic extract of Pothosscandens leaf. J. Herb. Medical Toxicology. 2009; (3): 159-160.
- [16] Sudha T, Chidambarampillai S, Mohan VR. GC-MS analysis of bioactive components of aerial parts of Fluggealeucopyrus Willd. (Euphorbiaceae). Journal of Applied Pharmaceutical Science. 2013; 3(5): 126.