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(RESEARCH ARTICLE)



Biological effectiveness of fungicides Rango, Tacora, Velficur and Opus for control of Karnal Bunt

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Abstract

Commercial fungicides Rango, Tacora, Velficur, and Opus were evaluated in the field, to determine their biological effectiveness to control karnal bunt ($Tilletia\ indica$) of wheat. A completely randomized design was used with four replications. Twenty heads of cultivar Tacupeto F2001 were inoculated during the boot stage with an allantoid sporidial suspension (10,000/mL). Commercial rates indicated in the containers of each product were followed. The first application was carried out ten days after inoculation (Zadoks stages 56-58), and the second one ten days later. Inoculated spikes were threshed by hand and the healthy and infected kernels were counted to determine the percentage of infection. Two hundred grains per treatment and per replication were weighed. The biological effectiveness of the products evaluated were Opus 91.6, Velficur 83.2, Tacora 80.2, and Rango 70.3%. The untreated inoculated check had a mean of 18.6% infection. There were no statistical differences for the products evaluated for level of infection after arcsin transformation (Tukey, p = 0.05) and the coefficient of variation was 16.2%. The average weight of 200 grains per treatment was 10.6, 10.5, 10.6, and 10.7 g, respectively, and 10.7 g for the untreated inoculated check. No phytotoxic effects of treatments applied to the wheat plant were observed.

Keywords: Karnal bunt; *Tilletia indica*; Biological effectiveness; Fungicides

1. Introduction

Karnal bunt (KB) of wheat caused by the fungus *Tilletia indica* Mitra has been reported in at least nine countries [1,2,3,4,5,6,7,8,9], and it is the most important disease of wheat seed and grain in northwest Mexico [10]. The causal agent affects partially some grains in a spike (Figure 1), and not all the spikes in a plant are infected [11]; occasionally, grains may be totally destroyed, and although the fungus may penetrate the embryo, it is not necessarily lethal [12,13]. Partially infected grains may produce healthy plants; some reports indicate that the percentage of seed germination decreases depending on the extent of infection [14,15,16]. Other reports indicate that severely affected seed lose viability or show abnormal germination [15]; on the other hand, Fuentes-Dávila *et al.* [17] found that seed with the greatest extent of infection, but with the embryo intact, produced the highest number of tillers. The effect of the disease on flour quality and the quarantine regulations, both, national and international, cause economic losses to farmers [18,19,20].

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Figure 1 Wheat grain infected with *Tilletia indica*, showing the characteristic lesion caused by the fungus

Breeding for genetic resistance to KB has been going on since the late 80's in northwest Mexico [21,22,23]; however, an integrated management program which would include seed density, rates of nitrogen for fertilization, sowing patterns, and chemical control, is of primary importance. Mitra [13,24]tried seed treatments with hot water and solar energy; since then, seed treatments with fungicides have been evaluated [25,26,27,28,29,30,31,32,33]. It has been found that some products inhibit teliospore germination, but do not control the disease since infection of the wheat plant is local and takes place during heading-flowering-anthesis; the life cycle of the fungus *Tilletia indica* is different than the other smuts of wheat [34]. According to Valenzuela-Rodríguez [35], incorporation of fungicides in soil drench does not reduce disease incidence. Since teliospores of *T. indica* are resistant to physical and chemical factors [36,37,38,39], but it causes floral infections during the dikaryotic stage [40] of its life cycle, the application of fungicides during heading-floweringanthesis of the wheat plant renders greater control of the disease and a more profitable economical margin [41]. Foliar have evaluated for control of fungicides been KB since [30,41,42,43,44,45,46,47,48,49,50,51,52,53]. Some of the products evaluated include: Approach Prima, Bavistin, Baycor, Bayfidan, Bayleton, Baytan, Bemistop, Benlate, Blitox, Ceresan, Consist, Dithane-M45, Duter, Folicur, Headline, Jewel, Kocide, Manzate, Maxtrobin Xtra, Nustar, Opus, Pointer, Priori Xtra, Sportak, Tilt, Topsin, Vangard, and Varon. The objective of this work was to evaluate several fungicides of the triazol group for control of karnal bunt in the field, under artificial inoculation.

2. Materials and methods

The experiment was carried out during the crop season 2020-2021 at the Norman E. Borlaug Experimental Station, located in block 910 of the Yaqui Valley at 27°22′04.64″ latitude north and 109°55′28.26″ longitude west, 37 masl, with climate warm [BW (h)] and extreme warm and dry [BS (h)], according to Köppen classification modified by Garcia [54]. Sowing date was December 17, 2020 with a seed density of 80 kg ha⁻¹. Treatments were established in a completely randomized experimental design (table 2) with four replications using bread wheat commercial cultivar Tacupeto F2001 [55]. The experimental plot consisted of 4 beds each with two rows 3 m long and 0.80 m between beds. The technical recommendation by INIFAP for the agronomic management was followed [56]. Inoculations were carried out during the boot stage by injection applying 1 mL per spike with an allantoid sporidial suspension (10,000/mL) in 20 spikes, in the central rows of each plot (Figure 2).

Inoculum was prepared as described by Fuentes-Bueno and Fuentes-Dávila [57]. Commercial rates indicated in the containers of each product were followed: Opus SC (BASF, epoxiconazol 12% a.i. in weight) as the regional check 1 L ha⁻¹ [58], Tacora 25 EW (Gowan, tebuconazole) 0.60 L ha⁻¹ [59], Velficur 25 EA (Velsimex, tebuconazole) 0.60 L ha⁻¹ [60], and Rango 250 EW (UPL, tebuconazole) 0.60 L ha⁻¹ [61] (Table 1). For application of fungicides, a manual Solo backpack sprayer (15 L) was used with a single nozzle, and the volume was based on 250 L of water/ha. To avoid the carry-over of the products applied, plastic barriers were used in each plot during the applications.

Table 1 Randomized complete distribution of treatments in the field for control of karnal bunt (*Tilletia indica*) by foliar applications, at the Norman E. Borlaug Experimental Station in the Yaqui Valley, Sonora, Mexico, during the crop season 2020-2021

20	19	18	17	16
R4	R4	R4	R4	R3
Opus	Untreated inoculated check	Rango	Tacora	Opus
11	12	13	14	15
R3	R2	R2	R4	R3
Tacora	Rango	Opus	Velficur	Rango
10	9	8	7	6
R3	R3	R2	R2	R2
Untreated inoculated check	Velficur	Untreated inoculated check	Tacora	Velficur
1	2	3	4	5
R1	R1	R1	R1	R1
Rango	Tacora	Velficur	Untreated inoculated check	Opus



Figure 2 Inoculation by injection with *Tilletia indica*, during the boot stage of the wheat plant

The first application was carried out ten days after inoculation (Zadoks stages 56-58) [62] and the second ten days later. Inoculated spikes were collected in paper bags and threshed by hand, and the percentage of infection was obtained by counting the number of infected and healthy grains from 20 inoculated spikes from each plot treated with the fungicides, and from 20 inoculated spikes from each plot of the untreated check. The biological effectiveness was obtained using Abbott's formula: effectiveness of treatments = average percentage of infection of the check – average percentage of infection of the treatment / average percentage of infection of the check x 100 [63]. The ANOVA was performed and mean comparison by Tukey's test (p = 0.05) to determine statistical differences among treatments, previous arcsin transformation $\sqrt{X} + 0.5$ [64]. The phytotoxicity was evaluated ten days after each application of the fungicides, according to the EWRS scale (Table 2) [65]. The weight of two hundred grains per treatment and per replication were recorded as well as the length, width and weight of 50 grains per treatment-replicate.

Table 2 Fungicides, formulation, concentration, and rates used to control karnal bunt by foliar applications, at the Norman E. Borlaug Experimental Station in the Yaqui Valley, Sonora, Mexico, during the crop season 2020-2021

Treatments	Formulation and concentrationy	Rate ^z CP ha ⁻¹
Tacora	250 EW 23.0% a.i.	0.60
Velficur	25.5 EA <u>></u> 25% a.i.	0.60
Rango	250 EW ≥ 25% a.i.	0.60
Opus	SC 12% a.i.	1.0
Untreated check		

yActive ingredient in weight. zLiters of commercial product

3. Results and discussion

Significant statistical differences were detected between treatments with products and the untreated check, with respect to the values of percentage of infection, and the coefficient of variation was 16.2% (Table 4). Mean comparison by Tukey's test indicated that all treatments with fungicide application were effective in reducing the percentage of infection, when compared with the untreated inoculated check, which showed the highest average percentage of infection (18.6%), with a range of 9.4 to 31.2 (Table 5).

Table 3 Values of the EWRS scale (1-9) to evaluate phytotoxicity in experimental plots, inoculated with karnal bunt and treated with Tacora, Velficur, Rango, and Opus, at the Norman E. Borlaug Experimental Station in the Yaqui Valley, Sonora, Mexico, during the crop season fall-winter 2020-2021

Value (Category)	Effect on the plant		
1	without effect		
2	very light symptoms		
3	light symptoms		
4	symptoms which are not reflected on yield		
5	Limit of acceptability medium damage		
6	elevated damage		
7	very elevated damage		
8	severe damage		
9	complete death		
Transformation of the EWRS punctual logarithmic scale to percentage			
Punctual value	value Phytotoxicity (%)		
1	0.0-1.0		
2	1.0-3.5		
3	3.5-7.0		
4	7.0-12.5		
5	12.5-20.0		
6	20.0-30.0		
7	30.0-50.0		
8	50.0-99.0		
9	99.0-100		

The real range of the mean percentage of infection obtained in spikes treated with the different products was 0.91 to 9.3% (Opus average 1.5, Velficur 3.1, Tacora 3.6, and Rango 5.5%). The biological effectiveness of the products evaluated were Opus 91.6, Velficur 83.2, Tacora 80.2, and Rango 70.3%.

Table 4 Analysis of variance of the percentage of infected grains with karnal bunt, in spikes treated with Opus, Tacora, Velficur, and Rango, and in spikes of an untreated check, at the Norman E. Borlaug Experimental Station in the Yaqui Valley, Sonora, Mexico, during the crop season fall-winter 2020-2021

Source of variation	DF	SS	MS	F value	F tab
Treatments	4	797.40	199.35	10.9	3.06
Error	15	272.09	18.13		
Total	19				
C.V.	16.2				

The overall average weight of 200 grains was 10.6 g with a range of 10.3 to 11.3; the average weight for each treatment was Rango 10.7, the untreated inoculated check 10.7, Opus 10.6, Tacora 10.6, and Velficur 10.5 g. Minimum differences were found among treatments and also with the untreated inoculated check, in grain length, width and weight; the overall average range of each trait were 0.72 to 0.74 cm, 0.36 to 0.38 cm, and 0.53 to 0.55 g, respectively.

Table 5 Mean separation by Tukey's test of the transformed percentages of infected grain with karnal bunt, in spikes treated with Opus, Tacora, Velficur, and Rango, at the Norman E. Borlaug Experimental Station in the Yaqui Valley, Sonora, Mexico, during the crop season fall-winter 2020-2021

Treatment	Infected grain (%)		Mean separation
	Real	Transformed	
Opus	1.5	7.09	A
Velficur	3.1	9.89	A
Tacora	3.6	9.99	A
Rango	5.5	13.47	A
Untreated inoculated check	18.6	25.06	В

Evaluation of fungicides applied during the heading-flowering-anthesis stage of the wheat plant, have demonstrated that products of the triazol group provide the best control of the disease, although, it does not control 100% [30,41,44,45,46,47,49,50,51,52,53], with the exception of the report by Sharma *et al.* [48] where they indicate that Folicur (tebuconazole) at 0.40 and 0.80%, and Contaf (hexaconazole) at 0.20% resulted in 100% control of karnal bunt under greenhouse conditions. Triazoles are the largest class of fungicides and their longevity is based on the fact that while being highly efficient broad spectrum products, resistance has occurred over time as a slow shift resulting in a decreased sensitivity to their mode of action [66]. The research carried out by Salazar-Huerta *et al.* [41] included experimentation in commercial fields using airplanes; the rate of 0.5 L of commercial product (Tilt - propiconazole) with two applications gave 99.2% control of the disease. This type of fungicides affect the biosynthesis of ergosterol, a primary component of the fungal cell plasma membrane [67,68]. The application of the different products did not cause any adverse effect on the growth and development of treated plants, according to the EWRS scale.

4. Conclusion

The biological effectiveness of Opus, Velficur, Tacora, and Rango for control of karnal bunt of wheat by foliar applications during heading-flowering-anthesis was 91.6, 83.2, 80.2, and 70.3%, respectively, being statistically similar.

The overall average weight of 200 grains was 10.6 g with a range of 10.3 to 11.3; the average weight for each treatment was Rango 10.7, the untreated inoculated check 10.7, Opus 10.6, Tacora 10.6, and Velficur 10.5 g. Minimum differences were found among treatments and also with the untreated inoculated check, in grain length, width and weight; the overall average range of each trait were 0.72 to 0.74 cm, 0.36 to 0.38 cm, and 0.53 to 0.55 g, respectively.

According to the EWRS scale, no phytotoxicity was detected on the wheat plants treated with the four fungicides.

Compliance with ethical standards

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Disclosure of conflict of interest

No conflict of interest to be disclosed.

References

- [1] Mitra M. 1931. A new bunt of wheat in India. Annals of Applied Biology 18:178-179.
- [2] Durán R. 1972. Further aspects of teliospore germination in North American smut fungi. II. Canadian Journal of Botany 50:2569-2573.
- [3] Munjal RL. 1975. Status of Karnal bunt (*Neovossia indica*) of wheat in Northern India during 1968-1969 and 1969-1970. Indian Journal of Mycology and Plant Pathology 5(2):185-187.
- [4] Singh DV, Agarwal R, Shrestha KJ, Thapa RB, and Dubin HJ. 1989. First report of *Tilletia indica* on wheat in Nepal. Plant Disease 73:273.
- [5] Da Luz WC, Mendes MAS, Ferreira MASV, and Urben AF. 1993. *Tilletia indica* on wheat in the south of the state of Rio Grande do Sul, Brazil and measures for eradication. Fitopatologia Brasileira 18:S329.
- [6] Ykema RE, Floyed JP, Palm ME, and Peterson JL. 1996. First report of Karnal bunt of wheat in United States. Plant Disease 80:1207. DOI: 10.1094/PD-80-1207B.
- [7] Torarbi M, Mardoukhi V, and Jalaiani N. 1996. First report on the occurrence of partial bunt on wheat in the southern parts of Iran. Seed and Plant 12:8-9.
- [8] Crous PW, Van Jaarsveld AB, Castlebury LA, Carris LM, Frederick RD, and Pretorius ZA. 2001. Karnal bunt of wheat newly reported from the African continent. Plant Disease 85:561.
- [9] CIMMYT (Centro Internacional de Mejoramiento de Maíz y Trigo). 2011. Training to beat karnal bunt in Afghanistan. http://blog.cimmyt.org/tag/karnal-bunt/. Accessed March 1, 2014.
- [10] Fuentes-Dávila G. 1997. Partial bunt of wheat: current situation and prospects. pp. 105-118. In: Primer Simposio Internacional del Trigo. Cd. Obregón, Sonora, México. 203 p.
- [11] Bedi SKS, Sikka MR, and Mundkur BB. 1949. Transmission of wheat bunt due to *Neovossia indica* (Mitra) Mundkur. Indian Phytopathology 2:20-26.
- [12] Chona BL, Munjal RL, and Adlakha KL. 1961. A method for screening wheat plants for resistance to *Neovossia indica*. Indian Phytopathology 14:99-101.
- [13] Mitra M. 1935. Stinking smut (bunt) of wheat with a special reference to *Tilletia indica* Mitra. Indian Journal of Agricultural Science 5:1-24.
- [14] Bansal R, Singh DV, and Joshi LM. 1984. Effect of Karnal bunt pathogen [*Neovossia indica* (Mitra) Mundkur] on weight and viability of wheat seed. Indian Journal of Agricultural Science 54:663-666.
- [15] Rai RC, and Singh AA. 1978. A note on the viability of wheat seeds infected with Karnal bunt. Seed Research 6:188-190.
- [16] Singh DV. 1980. A note on the effect of Karnal bunt infection on the vigour of wheat seed. Seed Research 8:81-82.

- [17] Fuentes-Dávila G, Figueroa-López P, Cortés-Jiménez JM, Félix-Valencia P, Camacho-Casas MA, Chávez-Villalba G, Félix-Fuentes JL, and Ortiz-Ávalos AA. 2013. Effect of the level of wheat seed infection with karnal bunt, on germination and tiller production. Annual Wheat Newsletter 59:45-48.
- [18] SARH (Secretariat of Agriculture and Water Resources). 1987. Internal Quarantine interior No. 16 against partial bunt of wheat. Secretaría de Agricultura y Recursos Hidráulicos. Diario Oficial, (jueves) 12 de Marzo de 1987, México.
- [19] Brennan JP, Warham EJ, Hernandez J, Byerlee D, and Coronel F. 1990. Economic Losses from Karnal bunt of wheat in Mexico. CIMMYT Economic Working Paper 90/02.
- [20] SAGARPA (Secretariat of Agriculture, Livestock, Rural Development, Fisheries and Food). 2002. Official Mexican Norm NOM-001-FITO-2001, establishing the campaign against wheat partial bunt. Diario Oficial viernes 8 de Febrero, 2002. México, D.F. pp. 1-18.
- [21] Metzger RJ. 1986. Screening for resistance to karnal bunt. pp. 25. In: Proceedings of the fifth biennial smut workers' workshop. April 28-30, 1986. Cd. Obregón, Sonora, México. p. 38
- [22] Warham EJ, Mujeeb-Kazi A, and Rosas V. 1986. Karnal bunt (*Tilletia indica*) resistance of *Aegilops* species and their practical utilization for *Triticum aestivum* improvement. Canadian Journal of Plant Pathology 8:65-70.
- [23] Rajaram S, Fuentes-Davila G, van Ginkel M, Getinet G, Camacho M, Montoya J, Amaya A, Peña J, He Zhong H, and Tinayou C. 1991. Breeding bread wheat resistance to Karnal bunt (*Tilletia indica*). pp. 14-15. In: Update on Karnal Bunt Research in Mexico. Wheat Special Report No. 7. CIMMYT, Mexico, D.F.
- [24] Mitra M. 1937. Studies on the stinking smut or bunt of wheat in India. Indian Journal of Agricultural Science 7:459-478.
- [25] Fuentes S, James WC, Torres E, Garcia C, Delgado S, and Elias-Calles A. 1983. Karnal bunt of wheat: risk appraisal and evaluation of the efficacy of seed dressings to control seedborne inoculum. CIMMYT Report. 49 pp.
- [26] Krishna A, and Singh RA. 1983. Effect of some organic compounds on teliospore germination and screening of fungicides against *Neovossia indica*. Indian Phytopathology 36:233-236.
- [27] Aujla SS, Sharma I, and Singh BB. 1986. Effect of various fungicides on teliospore germination of *Neovossia indica*. Journal of Research Punjab Agricultural University 23:442-443.
- [28] Salazar-Huerta FJ, Smilanick J, Prescott JM, Zillinsky F, and Metzger R. 1986a. Evaluation of 11 fungicides applied to foliage on three sowing dates for the control of partial bunt *Neovossia indica* (Mitra) Mundkur in wheat in the Yaqui Valley, Sonora, Cycle fall-winter 1985-86, CIANO-CAEVY, Cd. Obregón, Sonora, México.
- [29] Salazar-Huerta FJ, Prescott JM, Navarro-Soto AR, and Espinoza-Salazar T. 1986b. Chemical treatment of wheat seed to control partial bunt *Neovossia indica* (Mitra) Mundkur in the laboratory. Yaqui Valley, Sonora. Cycle fallwinter 1985-86. CAEVY-CIFAP, Cd. Obregón, Sonora, México.
- [30] Smilanick JL, Hoffmann JA, Cashion NL, and Prescott JM. 1987. Evaluation of seed and foliar fungicides for control of Karnal bunt of wheat. Plant Disease 71:94-96.
- [31] Figueroa-Lopez P, and Espinoza-Salazar T. 1988. Chemical treatment with fungicides of wheat seed to control partial bunt *Tilletia indica* Mitra in the laboratory. Technical report of the Yaqui Valley Experimental Agricultural Station, Sonora, México. CAEVY-CIANO-INIFAP. pp. 22.
- [32] Warham EJ, and Prescott JM. 1989. Effectiveness of chemical seed treatments in controlling Karnal bunt disease of wheat. Plant Disease 73:585-588.
- [33] Robles-Sosa SD, and Fuentes-Davila G. 1996. Fungicide seed treatments for Karnal bunt control. Xth Biennial Workshop on the Smut Fungi. June 9-12, 1996, University of Calgary, Calgary, Alberta, Canada. Page 51.
- [34] Wilcoxson RD, and Saari EE (eds.). 1996. Bunt and smut diseases of wheat: concepts and methods of disease management. CIMMYT, Mexico, D.F. 66 p.
- [35] Valenzuela-Rodriguez S. 1985. Evaluation of 4 fungicides for control of karnal bunt of wheat. Cycle fall-winter 1984-85. CIANO-CAEVY, Cd. Obregon, Sonora, México.
- [36] Krishna A, and Singh RA. 1982. Effect of physical factors and chemicals on the teliospore germination of *Neovossia indica*. Indian Phytopathology 35(3):448-455.
- [37] Zhang Z, Lange L, and Mathur SB. 1984. Teliospore survival and plant quarantine significance of *Tilletia indica* (causal agent of Karnal bunt) particularly in relation to China. European Plant Protection Bulletin 14:119-128.

- [38] Smilanick JL, Hoffmann JA, and Royer MH. 1985. Effect of temperature, pH, light, and desiccation on teliospore germination of *Tilletia indica*. Phytopathology 75:1428-1431.
- [39] Smilanick JL, Hoffmann JA, Secrest LR, and Wiese K. 1988. Evaluation of chemical and physical treatment to prevent germination of *Tilletia indica* teliospores. Plant Disease 72:46-51.
- [40] Fuentes-Davila G, and Duran R. 1986. *Tilletia indica*: cytology and teliospore formation *in vitro* and immature kernels. Canadian Journal of Botany 64(8):1712-1719. https://doi.org/10.1139/b86-229.
- [41] Salazar-Huerta FJ, Figueroa-Lopez P, Smilanick JL, and Fuentes-Davila G. 1997. Evaluation of foliar fungicides for control of Karnal bunt of wheat during 1986-1989 in northwestern Mexico. Revista Mexicana de Fitopatología 15:73-80.
- [42] Singh A, and Prasad R. 1980. Control of Karnal Bunt of wheat by a spray of fungicides. Indian Journal of Mycology and Plant Pathology 10:2. (Abstract).
- [43] Singh SL, and Singh PP. 1985. Effect of some fungicide applications against Karnal Bunt (*Neovossia indica*) of wheat. Indian Phytopathology 38:593. (Abstract).
- [44] Figueroa LP y Valdés AJC. 1991. Evaluation of systematic fungicides for the control of Partial bunt *Tilletia indica* (Mit.) in wheat in the Yaqui Valley. Proceedings of the XVIII Congreso Nacional de la Sociedad Mexicana de Fitopatología, del 24 al 26 de julio de 1991. Puebla, Puebla, México. (Abstract). p 209.
- [45] Figueroa-López P y Álvarez-Zamorano R. 2000. Opus (epoxiconazole): a new option to control partial bunt in wheat (*Tilletia indica* Mitra) in foliar application. pp. 31-34. In: G. Fuentes-Dávila (ed.). XIIth Biennial Workshop on the Smut Fungi. Sociedad Mexicana de Fitopatología, A.C. July 9-12, 2000, Puerto Vallarta, Jalisco, México. 65 p.
- [46] Goel LB, Singh DP, Sinha VC, Singh DV, Srivastava KD, Rashmi A, Aujla SS, Indu Sharma, Bagga PS, Singh RV, Singh AK, Singh SP. 2000. Evaluation of Tilt against Karnal bunt of wheat. Indian Phytopathology 53(3):301-302.
- [47] Singh R, Beniwal MS, Karwasra SS. 2000. Field evaluation of fungicides against Karnal bunt of wheat through foliar spray. Department of Plant Pathology, Chaudhary Charan Singh Haryana Agricultural University, Hisar 125 004, India. Wellesbourne, UK: Association of Applied Biologists. Tests of Agrochemicals and Cultivars 21:9-10.
- [48] Sharma AK, Singh DP, Kumar J, Sharma I, and Sharma BK. 2005. Efficacy of some new molecules against Karnal bunt (*Tilletia indica*) of wheats (*Triticum aestivum* and *T. durum*). Indian Journal of Agricultural Sciences 75(6):369-370.
- [49] Fuentes-Dávila G. 2007. Chemical control of karnal bunt by foliar applications. Phytopathology 97(7):S37. Supplement.
- [50] Fuentes-Dávila G, Tapia-Ramos E, Toledo-Martínez JA y Figueroa-López P. 2005. Evaluation of biological effectiveness of folicur 250 EW (Tebuconazole) for the control of partial bunt (*Tilletia indica*) of wheat (*Triticum aestivum*), in the Yaqui Valley, Sonora, Mexico, during the 2003-2004 crop cycle. Proceedings of the XIII Congreso Latinoamericano de Fitopatología, III Taller de la Asociación Argentina de Fitopatólogos. 19-22 de Abril, 2005. Villa Carlos Paz, Córdoba, Argentina. Resumen HC-29, página 271. 640 p.
- [51] Fuentes-Dávila G, Félix-Valencia P, Ayón-Ibarra CA, Figueroa-López P, Camacho-Casas MA, Félix-Fuentes JL, Chávez-Villalba G, and Rosas-Jáuregui IA. 2016. Biological effectiveness of several fungicides for control of karnal bunt (*Tilletia indica*) of wheat, in the field. Annual Wheat Newsletter 62:28-31.
- [52] Fuentes-Dávila G, Rosas-Jáuregui IA, Ayón-Ibarra CA, Álvarez-Amado KD, Félix-Valencia P, and Félix-Fuentes JL. 2018. Biological effectiveness of Opus, Folicur, Juwel, and Bemistop for control of Karnal bunt (*Tilletia indica*) of wheat in the field. Annual Wheat Newsletter 64:30-33.
- [53] Fuentes-Dávila G, Torres-Cruz MM, Félix-Valencia P, Rosas-Jáuregui IA, and Félix-Fuentes JL. 2022. Biological effectiveness of fungicides Priori Xtra, Folicur, Approach Prima, and Maxtrobin Xtra for control of karnal bunt. International Journal of Agriculture, Environment and Bioresearch 7(4):83-94. https://doi.org/10.35410/IJAEB.2022.5747.
- [54] García E. 2004. Modifications to the Köppen climate classification system. Institute of Geography of the National Autonomous University of Mexico. Book Series number 6. México, D.F. 90 p. Available at: http://www.publicaciones.igg.unam.mx/index.php/ig/catalog/view/83/82/251-1. Accessed on May 11, 2023.

- [55] Camacho-Casas MA, Singh RP, Figueroa-López P, Huerta-Espino J, Fuentes-Dávila G. y Ortiz-Monasterio-Rosas I. 2003. Tacupeto F2001: new variety of bread wheat for northwest Mexico. Technical Brochure Number 50, National Institute of Forestry, Agricultural and Livestock Research, Northwest Regional Research Center, Yaqui Valley Experimental Station. Ciudad Obregón, Sonora, México. 20 p.
- [56] Figueroa-López P, Fuentes-Dávila G, Cortés-Jiménez JM, Tamayo-Esquer LM, Félix-Valencia P, Ortiz-Enríquez E, Armenta-Cárdenas I, Valenzuela-Herrera V, Chávez-Villalba G. y Félix-Fuentes JL. 2011. Guide to producing wheat in southern Sonora. Brochure for producers No. 39. INIFAP-CIRNO, Norman E. Borlaug Experimental Station. Cd. Obregón, Sonora, México. 63 p.
- [57] Fuentes-Bueno I, and Fuentes-Dávila G. 2007. Reaction of wheat cultivars WL-711 (*Triticum aestivum*) and Altar C84 (*T. turgidum* subsp. *turgidum*) to inoculation with *Tilletia indica* cultures obtained from infected wheat cultivars Baviacora M92 (*T. aestivum*) and Altar C84 under natural conditions in the Yaqui valley, Sonora, Mexico. Annual Wheat Newsletter 53:48-52.
- [58] BASF. 2020. Data sheet. Opus agricultural fungicide. https://www.tacsa.mx/DEAQ/src/productos/155058.htm. Accessed on November 21, 2020.
- [59] Gowan México. 2020. Data sheet Tacora 25 EW. https://mx.gowanco.com/sites/default/files/mxgowancocom/attachments/product/resource/label/tacora_ft-031.pdf. Accessed on November 20, 2020.
- [60] Velsimex. 2020. Data sheet. Velficur 25 EW. http://www.velsimex.com/wp-content/uploads/2022/06/ fichatecnica-velficur-impetor.pdf. Accessed on November 22, 2020.
- [61] Agroquímicos de México. 2020. Agricultural technical portal. Rango 250 EW. https://www.buscador. portaltecnoagricola.com/vademecum/mex/producto/23370/RANGO%20250%20EW. Accessed on November 22, 2020.
- [62] Zadoks JC, Chang TT, and Konzak CF. 1974. A decimal code for the growth stages of cereals. Weed Research 14:415-421. https://doi.org/10.1111/j.1365-3180.1974.tb01084.x
- [63] Abbott WS. 1925. A method of computing the effectiveness of an insecticide. Journal of Economic Entomology 18:265-267. http://dx.doi.org/10.1093/jee/18.2.265a.
- [64] Steel RGD, and Torrie JH. 1980. Principles and procedures of statistics. A biometrical approach. Second edition. McGraw-Hill Book Company. New York, NY, USA. 633 p.
- [65] Champion GT. 2000. Bright and the field scale evaluations herbicides tolerant. G M Trials. AICC Newsletter, December 2000, 7.
- [66] Morton V, and Staub T. 2008. A short history of fungicides. APSnet Features. https://www.apsnet.org/edcenter/apsnetfeatures/Pages/Fungicides.aspx. doi: 10.1094/APSnet Feature-2008-0308.
- [67] Pérez-García A, Fernández-Ortuño D, De Vicente A, Torés JA y López-Ruiz FJ. 2005. Resistance to ergosterol biosynthesis inhibitors and strobilurins in cucurbit powdery mildew. https://www.phytoma.com/larevista/phytohemeroteca/173-noviembre2005/resistencia-a-inhibidores-de-la-biosntesis-de-ergosterol-y-a-estrobilurinas-en-odio-decucurbitceas. Accessed on April 15, 2022.
- [68] Ribas e Ribas AD, Spolti P, Medeiros Del Ponte E, Zawada Donato K, Schrekker H, and Meneghello Fuentefria A. 2016. Is the emergence of fungal resistance to medical triazoles related to their use in the agroecosystems? A mini review. Brazilian Journal of Microbiology 47:793-799.