

Available online at GSC Online Press Directory

GSC Biological and Pharmaceutical Sciences

e-ISSN: 2581-3250, CODEN (USA): GBPSC2



Journal homepage: <u>https://www.gsconlinepress.com/journals/gscbps</u>

(RESEARCH ARTICLE)



Comparative secondary metabolite compositions and anti-microbial properties of nhexane and ethyl-acetate fractions of *Nelsonia campestris* 

Mohammed Hadiza, Muhammad Hadiza Lami and Makun Anthony Hussaini

Department of Biochemistry, Federal University of Technology Minna, Nigeria.

Publication history: Received on 21 August 2019; revised on 28 September 2019; accepted on 04 October 2019

Article DOI: https://doi.org/10.30574/gscbps.2019.9.1.0180

# Abstract

Nelsonia campestris is a medicinal plant commonly used to treat measles caused by morbillivirus. This study assessed the phytochemical composition, in vitro antimicrobial activity of n-hexane and ethyl-acetate fractions of *Nelsonia campestris*. Phytochemical composition was determined using standard procedures. Antimicrobial test against six microorganisms, including K. pneumonia, S. aureus, B. subtilis, E. coli, P. aeruginosa and S. typhi was investigated using agar diffusion method while minimum inhibitory concentrations were determined by broth micro dilution method. Results revealed the presence of alkaloids, phenols, tannis, flavonoids, saponins and steroids in both n-hexane and ethyl-acetate fraction. Glycoside was present in n-hexane fraction but absent in ethyl-acetate fraction while volatile oil and anthraquinone were completely absent. Quantitatively, n-hexane fraction had higher phenol (57.12±0.01 mg/100g), tannins (19.76±0.04 mg/100g), and saponins (104.78±0.05 mg/100g) while ethyl-acetate had higher flavonoid (27.85±0.50 mg/100g) and alkaloids (16.62±0.01 mg/100g). The ethyl-acetate fraction was highly active against all test organisms with inhibition diameters in the range of 7.50±0.56 mm (*P. aeruginosa*) to 21.50±0.32 mm (E. coli) The MIC ranges between 2.5-20 mg/mL while the MBC ranged between 5.0-30 mg/ml for the ethyl-acetate fraction. The n-hexane fraction was active only against *P. aeruginosa* and *B. subtilis* with inhibition diameters in the range of 12.50±0.10 mm-17.55±1.00 mm. This study has shown that the n-hexane and ethyl-acetate fractions of Nelsonia campestris contain some useful potential bioactive principles that are inhibitory to some pathogenic organisms. The ethyl-acetate fraction was more active than n-hexane fraction and thus could be considered as a natural source of antimicrobials for therapeutic purposes.

Keywords: Nelsonia campestris; Secondary metabolite; Antimicrobial; N-hexane and ethyl-acetate fractions

# 1. Introduction

Microbial infections remain a threat to millions of lives globally as they are ranked the second leading cause of death across the world and the third leading cause of death of economically developed countries [1]. Currently there is an increased problem of antibiotic resistance due to microbial resistance [2]. The uses of antibiotics are widespread in clinical medicine, agriculture, and veterinary promote the development of antibiotic resistances among infectious microbial strains and eventually reflects a very serious problem in the treatment of pathogenic microbes [3]. This has led to the search of new antimicrobial agents mainly among plant extracts with the goal to discover new chemical structures which overcome the above disadvantages [4]

Plants are free gifts of nature available to man for various pharmacological uses. The World Health Organization (WHO) recognizes traditional medicine, particularly plant medicine, as an important alternative healthcare delivery system for most of the world's population [5]. They are source of active chemical compounds such as alkaloids tannins, flavonoids, sugars phenols, coumarins, terpenoids, saponins, etc useful to both Man and animals. In Africa,

\* Corresponding author

Copyright © 2019 Author(s) retain the copyright of this article. This article is published under the terms of the Creative Commons Attribution Liscense 4.0

E-mail address: cutedeeja@gmail.com

traditional medicine, especially plant medicine, provide many citizens with affordable healthcare services [6-8]. Since prehistoric times man has used plants for various purposes and will continue to do so as long as life continues on this planet [9].Traditional medicine is either the mainstay of health care delivery or serves as a complement to it across the world [10]. The medicinal value of plants lies in some chemical substances that produce a definite physiological action on the human body. Many plant leaves have antimicrobial principles such as tannins, essential oils tannins, flavanoids and other aromatic compounds [11].

*Nelsonia campestris* (Lam.) Spreng of the family Acanthaceae, is a small trailing delicate herb with long underground root and aerial spreading stem with plenty of leaves. The traditional healers use the root, fruit, and leaves of the plant for different disease conditions [12]. This plant, known as "Isa bakaggi," by the traditional practitioners of Nupe speaking people of Niger state, Nigeria, is used to treat measles caused by morbillivirus of the Paramyxovirus family, with concurrent development of other opportunistic infections such as pneumonia, conjunctivitis, diarrhea, and infection of the otitis media [13]. It is also used traditionally for the treatment of skin infections such as chicken pox, fevers, constipation and gastric ulcer, in addition to measles and lots of others. Studies has documented the antibacterial and safety profile of the crude extract of this plant [12-14]. However, scientific literature on antimicrobial properties of solvent fractions of this plant is currently unavailable, the present study therefore evaluated the secondary metabolites compositions and *in vitro* antimicrobial profile of n-hexane and ethyl-acetate fractions of *Nelsonia campestris* leaf.

# 2. Material and methods

## 2.1. Plant sample

Fresh samples of *Nelsonia campestris* was obtained from the premises of Federal University of Technology, Minna, Bosso Campus, Niger State Nigeria. A taxonomic authentication of the plant was carried out by Botanist at the Department of Biological science, Federal University of Technology, Minna. Nigeria.

### 2.2. Sources of microorganisms

The microbial isolates used were collected from the Laboratory of Microbiology Department Federal University of Technology, Minna, Nigeria. The microorganisms include: *Staphylococcus aureus, Bacillus subtilis, Pseudomonas aeruginosa, Klebsiella pneumonie, Escherichia coli*, and *Salmonella typhii*. These organisms were obtained and identified by standard microbiological methods. The organisms were maintained on agar slants at 4oC and subcultured for 24 hours before use.

### 2.3. Sample preparation and extraction

The collected fresh samples of *Nelsonia campestris* were washed with clean-water, dried at room temperature and finally grounded using a grinder mill and dried at room temperature. Extraction of plant materials was performed by weighing 50 g of the powdered plants and extracted by soxhlet extraction using 200 ml each of methanol [13]. The resulting extract was concentrated in a water bath and then fractionated with n-hexane and ethyl-acetate to give n-hexane and ethyl-acetate fractions respectively.

### 2.4. Qualitative and quantitative screening for secondary metabolites

The plant extract was analyzed for the presence of some secondary metabolite including alkaloids, terpenes, tannins, saponins, phenols, steroids, phlobatannins and flavonoids using standard procedures [15-18]. Quantitative analysis was conducted for flavonoid, alkaloids, total phenol, tannin and saponins using standard procedures [19].

### 2.5. Antimicrobial activity

The antimicrobial activities of the fractions were evaluated at concentrations of 20, 30, and 40 mg/ml using agar well diffusion method as described by Yusuf *et al.* [20]. The antibacterial activity was expressed as the mean zone of inhibition diameters (mm) produced by the plant fractions. The minimum inhibitory concentration (MIC) and Determination of minimum bactericidal concentration (MBC) were evaluated as reported by Tsado et al. [21]. MIC was determined as the lowest concentration of the fractions permitting no visible growth (no turbidity) of the organism. The MBC was determined by sub-culturing the test dilution on fresh solid medium and further incubating at 37°C for 18-24 hours. The lowest concentration of MIC tubes with no visible bacterial growth on solid medium was regarded as MBC.

## 2.6. Statistical analysis

Values were analyzed using statistical package for social science (SPSS) version 21 and presented as means  $\pm$  SE of the mean. Comparisons between different groups were carried out by one-way analysis of variance (ANOVA) followed by Duncan's Multiple Range Test (DMRT). The level of significance was set at *P* < 0.05.

# 3. Results

## 3.1. Phytochemical composition

Phytochemical screening of n-hexane and ethyl-acetate fractions of *Nelsonia campestris* revealed the presence of alkaloids, phenols, tannis, flavonoids, saponins and steroids in both n-hexane and ethyl-acetate fraction. Glycoside was present in n-hexane fraction but absent in ethyl-acetate fraction while volatile oil and anthraquinone were completely absent (Table 1). Quantitatively, n-hexane fraction had higher phenol (57.12±0.01 mg/100g), tannins (19.76±0.04 mg/100g), and saponins (104.78±0.05 mg/100g) while ethyl-acetate had higher flavonoid (27.85±0.50 mg/100g) and alkaloids (16.62±0.01 mg/100g (Table 2).

Table 1 Qualitative phytochemical composition of n-hexane and ethyl-acetate fractions of Nelsonia campestris extract

Phytochemicals	n-hexane	Ethyl-acetate
Total phenol	+	+
Total flavonoids	+	+
Tannins	+	+
Alkaloids	+	+
Saponins	+	+
Glycoside	-	+
Volatile oil	-	-
Anthraquinonne	-	-

Key: + means present and – means absent

Table 2Quantitative phytochemical composition of n-hexane and ethyl-acetate fractions of Nelsonia campestrisextract

Phytochemicals	n. Hexane (mg/100g)	Ethyl-acetate (mg/100g)
Total phenol	57.12±0.01	54.59±0.01
Total flavonoids	11.06±0.01	27.85±0.50
Tannins	19.76±0.04	12.58±0.02
Alkaloids	12.84±0.00	16.62±0.01
Saponins	104.78±0.05	86.78 ±0.03

### 3.2. Antimicrobial activities

The Zones of inhibition of n-hexane and ethyl-acetate fractions of *Nelsonia campestris* against *K. pneumoniae, S. typhi, S. aureus, E. coli, P. aeruginosa* and *B. subtilis* are presented in table 3. The zones of inhibition demonstrated by the extracts increased with increasing concentration. The ethyl-acetate fraction was highly active against all test organisms with inhibition diameters in the range of  $7.50\pm0.56$  mm (*P. aeruginosa*) to  $21.50\pm0.32$  mm (*E. coli*) The MIC ranges between 2.5-20 mg/mL while the MBC ranged between 5.0-30 mg/ml for the ethyl-acetate fraction. The n-hexane fraction was active only against *P. aeruginosa* and *B. subtilis* with inhibition diameters in the range of  $12.50\pm0.10$  mm- $17.55\pm1.00$ mm. However standard antibiotics demonstrated highest activity against all organisms tested with inhibition zone in the range  $28.06\pm0.34$  mm- $35.07\pm0.03$  mm.

Organism	Ethyl-acetate fraction (zone of inhibition MM)		n-hexane fraction (zone of inhibition MM)			Ciprofloxacin (mg/ml)	
	20 mg/ml	30 mg/ml	40 mg/ml	20 mg/ml	30 mg/ml	40 mg/ml	40
P. aeruginosa	7.50±0.56	11.5±0.56	12.5±0.32	12.50±0.10	13.50±0.23	17.55±1.00	28.06±0.34
B. subtilis	12.50±0.50	14.00±0.40	17.5±0.10	13.00±0.50	12.00±0.55	15.00±0.15	33.00±0.00
K. pneumonia	$10.00 \pm 0.04$	14.0±0.55	16.00±0.10	-	-	-	35.07±0.03
S. aureus	13.00±0.42	16.00±0.07	17.50±0.50	-	-	-	27.50±0.50
E. coli	16.50±0.50	16.00±0.15	21.50±0.32	-	-		28.50±0.50
S. typhi	10.50±0.10	11.50±0.55	13.00±0.50	-	-	-	30.50±0.0

**Table 3** Antimicrobial activities of n-hexane and ethyl-acetate fractions of *Nelsonia campestris* showing zones ofinhibition of the organisms

**Table 4** Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentrations (MBC) of ethyl-acetate

 fractions of *Nelsonia campestris*

Test Organisms	MIC (mg/ml)	MBC (mg/ml)	
P. aeruginosa	20	30	
B. subtilis	20	20	
K. pneumonia	2.5	5.0	
S. aureus	10	20	
E. coli	5	`10	
S. typhi	10	20	

# 4. Discussion

Plants are admittedly, a valuable reservoir of bioactive compounds of substantial medical importance [22]. Qualitative phytochemical analysis clearly demonstrated the presence of a number of important active constituents and revealed that both n-hexane and ethyl-acetate fractions of *N. campestris* have similar phytochemical constitution of alkaloids, phenols, tannis, flavonoids, saponins and steroids. The presence of these phytochemicals has been previously reported for crude aqueous extract of *Nelsonia campestris* [13]. Only the glycoside was present in n-hexane fraction but absent in ethyl-acetate fraction. This agrees with the previous study which reported that not all phytochemicals are present in medicinal plants and those present varies with their solvents [23]. Indeed, all kinds of metabolite classes detected into the samples (saponins, flavonoids, tannins, alklaoids and phenols) are well known to have significant inhibitory action against bacteria and fungi [24]. This fact can justify why all fractions showed antimicrobial activity.

Several authors have established that compounds, such as phenols, flavonoids, alkaloids and tannins, have antimicrobial activity [25-27]. These properties arise from polyphenol and tannin capacity to inhibit enzymes and alter membrane characteristics. Enzyme inhibition by phenols results from compound oxidation, possibly through sulfhydryl group reaction with or through nonspecific protein interactions. Flavonoids can cause cell membrane disruption, and form complexes with soluble and extracellular proteins as well as with membrane walls [25-26]. On the other hand, quantitative assays revealed that the extracts have different quantity of the secondary metabolite which can be correlated to their antimicrobial activity.

The antimicrobial activities of n-hexane and ethyl-acetate fractions of *Nelsonia campestris* showed that the fractions had varying degree of antimicrobial activity against the test organisms. On the basis of the results the fractions

demonstrated to be more active on Gram-positive than on Gram-negative bacteria. This was not surprising since previous studies have reported that generally plant extracts are usually more active against Gram-positive bacteria than Gram-negative bacteria, and the susceptibility may be due to structural differences in the cell wall of these classes of bacteria. Cells of Gram-negative bacteria are surrounded by an additional outer membrane, which provide them with a hydrophilic surface that functions as a permeability barrier for many substances including natural compounds [28-29]. It was also observed that increase in the concentration of all the extracts yielded their corresponding increase in the zones of inhibition. This linear relationship between the concentrations of extracts and zones of inhibition could be that the extracts were able to diffuse into the inoculated nutrient agar [30]. The ability of the n-hexane and ethyl-acetate fractions of *Nelsonia campestris* to inhibit the growth *Klebsiella pneumoniae, Pseudomonas aeruginosa, Salmonella typhi, Staphylococcus aureus* and *E.coli* explains why it is used in folk medicine for the treatment of gastrointestinal and respiratory complications associated with measles.

The extracts were found to be more active with greater zone of inhibition at concentration of 40 mg/ml, this was in accordance with the results of the previous studies, who reported that the higher concentration of antibacterial substance showed appreciable growth inhibition of bacteria [13, 30-31]. Ethyl-acetate fraction was highly active against all test organisms with inhibition diameters in the range of  $7.50\pm0.56$  mm (*P. aeruginosa*) to  $21.50\pm0.32$ mm (*E. coli*) while the n-hexane fraction was active only against *P. aeruginosa* and *B. subtilis* with inhibition diameters in the range of  $12.50\pm0.10$  mm- $17.55\pm1.00$ mm. This result is not surprising since total flavonoids and alkaloids which were the major antimicrobial metabolites are found in higher concentration in the ethyl-acetate fraction than the n-hexane fraction. However, despite the higher zone of inhibition demonstrated by the ethyl-acetate fraction of *Nelsonia campestris*, the zone of inhibitions was lower on all test organism compare to zone of inhibitions demonstrated by standard antibiotics drug (ciprofloxacin) used in this study.

Minimum inhibitory concentration (MIC) is the lowest concentration of an extract that inhibit the visible growth of the test organism after 24hrs incubation [32-33]. In the present study MIC was measured based on turbidity or visible growth show by the organism. The MICs of ethyl-acetate fractions of *Nelsonia campestris* MIC ranges between 2.5-20 mg/ml. This MIC is lower than 80-90 mg/ml previously reported for crude aqueous extract of *Nelsonia campestris* [13], thus indicating higher activity of the fractionated extract compared to the crude extract.

# 5. Conclusion

This study has shown that the n-hexane and ethyl-acetate fractions of *Nelsonia campestris* contain some useful potential bioactive principles that are inhibitory to some pathogenic organisms. The ethyl-acetate fraction was more active than n-hexane fraction and thus could be considered as a natural source of antimicrobials for therapeutic purpose.

# **Compliance with ethical standards**

### Acknowledgments

The authors are grateful to the academic and non-academic staff of Biochemistry Department Federal University of technology Minna. We express our gratitude to Mallam Hamidu of Microbiology Department Federal University of technology Minna for assistance during the antimicrobial analysis.

# Disclosure of conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# References

- [1] De Zoysa MHN, Rathnayake H, Hewawasam RP and Wijayaratne WMD. (2019). Determination of *in vitro* antimicrobial activity of five Sri Lankan medicinal plants against selected human pathogenic bacteria.
- [2] Kitonde CK, Dossaji SF, Lukhoba CW and Jumba MM. (2013). Antimicrobial activity and phytochemical study of vernonia glabra(steetz) oliv. and hiern.in Kenya. African Journal of Traditional, Complementary and Alternative Medicines, 10, 149-157.

- [3] Bereksi MS, Hassaïne H, Bekhechi C and Abdelouahid DE. (2018). Evaluation of Antibacterial Activity of Some Medicinal Plants Extracts Commonly Used in Algerian Traditional Medicine against Some Pathogenic Bacteria. Pharmacog J, 10(3), 507-12.
- [4] Pandian MR, Banu GS and Kumar G. (2006). A study of antimicrobial activity of Alanguim salviifolium .Indian Journal of pharmacology, 38, 203-204.
- [5] WHO (2010). Traditional Medicine and Modern Health Care: Progress report by the Director General Document A 44(10), 22.
- [6] Lawal B, Shittu OK, Oibiokpa FI, Berinyuy EB and Muhammed H. (2016). African natural products with potential antioxidants and hepatoprotectives properties, a review Clinical Phytoscience, 2(23), 1-66.
- [7] Bashir L, Shittu OK, Sani S, Busari MB and Adeniyi KA.(2015). African Natural Products with Potential Antitrypanosoma Properties, A Review International Journal of Biochemistry Research & Review, 7(2), 45-79.
- [8] Lawal B, Shittu OK, Kabiru AY, Jigam AA, Umar MB, Berinyuy EB and Alozieuwa BU. (2015). Potential antimalarials from African natural products, A review J Intercult Ethnopharmacology, 4(4), 318-343.
- [9] Okigbo RN and Igwe DI. (2007). The antimicrobial effects of Piper guineense 'uziza'and Phyllantus amarus 'ebebenizo'on Candida albicans and Streptococcus faecalis. Acta Microbiologica et Immunologica Hungarica, 54 (4), 353-366.
- [10] Okoro IO, Auguster O and Edith OA.(2010). Antioxidant and antimicrobial activities of polyphenols from ethnomedicinal plants of Nigeria. African Journal of Biotechnology, 9, 2989-2993.
- [11] WHO. (2008). Traditional medicine; growing needs and potential.WHO Policy perspectives on medicines. Geneva 1-6.
- [12] Nabèrè O, Adama H, Samson S, Kiessoum K, Patrice Z, Roland MN, Moussa C, Martin K etc.. (2013). Antibacterial and Phytochemical studies of three Acanthaceaespecies used in Burkina Faso traditional medicine. J App Pharm Sci, 3 (05), 049-055.
- [13] Muhammad HL, Makun HA, Kabiru AY, Mann A, Busari MB, Abdullah AS and Fatima A. (2015). *In vitro* Antibacterial Activity and *In vivo* Acute Toxicological Studies of Nelsonia campestris aqueous Leaf Exrtact. International Journal of Biochemistry Research and Review, 7(1), 27-35.
- [14] Olaniyan JM, Muhammad HM, Makun HA, Busari MMB and Abdullah A.S. (2015). Acute and sub-acute toxicity studies of aqueous and methanol extracts of Nelsonia campestris in rats. Journal of Acute Disease, 5(1), 62–70.
- [15] Tsado AN, Lawal B, Mohammed SS, Famous IO, Yahaya AM, Shu'aibu M and Caleb T. (2015). Phytochemical Composition and Antimalarial Activity of Methanol Leaf Extract of Crateva adansonii in P.berghei Infected Mice. British Biotechnology Journal, 6(4), 165-173.
- [16] Harborne JB. (1993). Phytochemical Methods; A guild to modern Techniques to plant anaylsis Freeman and Company. NY, 78-80.
- [17] Trease GE and Evans WC. (1989). Pharmacognosy. 11th ed. London: Brailliar Tiridel CanMacmillan Publishers, 60–75.
- [18] Sofowora AE. (1993). Recent trends in research into African medicinal plants. J Ethnopharmacol. 389, 209–14.
- [19] AOAC. (Association of official analytical chemist) (2000). Official Method analytical chemist, Washinton, D.C.
- [20] Yusuf AA, Lawal B, Yusuf MA, Omonije YO, Adejoke AA, Raji FH and Wenawo DL.(2018). Free Radical Scavenging Antimicrobial Activities and Effect of Sub-Acute Exposure to Nigerian Xylopia Aethiopica Seed Extract On Liver and Kidney Functional Indices of Albino Rat Iranian journal of toxicology ,12 (3), 51-58.
- [21] Tsado NA, Lawal B Kontagora GN, Muhammad BM, Yahaya MA, Gboke JA Muhammad UA and Hassan MK. (2016). Antioxidants and Antimicrobial- Activities of Methanol Leaf Extract of Senna occidentalis Journal of Advances in Medical and Pharmaceutical Sciences, 8(2), 1-7.
- [22] Harvey AL, Edrada-Ebel R and Quinn RJ. (2015). The re-emergence of natural products for drug discovery in the genomics era. Nat. Rev. Drug Discov. 14, 111–129.
- [23] Lawal B, Ossai PC, Shittu OK and Abubakar AN. (2014). Evaluation of phytochemicals, proximate, minerals and anti-nutritional compositions of yam peel, maize chaff and bean coat. International Journal of Applied Biological Research, 6(2), 01-17.

- [24] Tiwari P, Kumar B, Kaur M, Kaur G and Kaur H. (2011). Phytochemical screening and extraction: A review. Int. Pharm. Sci., 1, 98–106.
- [25] Cowan MM. (1999). Plant products as antimicrobial agents. Clin. Microbiol. Rev., 12, 564–582.
- [26] Pandey AK and Kumar S. (2013). Perspective on plant products as antimicrobials agents: A review. Pharmacologia, 4, 469–480.
- [27] Hayek SA, Gyawali R and Ibrahim SA. (2013). "Antimicrobial natural products" in Microbial Pathogens and Strategies for Combating them: Science, Technology and Education, ed A. Méndez-Vilas (Badajoz: FORMATEX), 910–921.
- [28] Hemaiswarya S, Kruthiventi AK and Doble M. (2008). Synergism between natural products and antibiotics against infectious diseases. Phytomedicine, 15, 639–652.
- [29] Briers Y and Lavigne R. (2015). Breaking barriers: expansion of the use of endolysins as novel antibacterials against Gram-negative bacteria. Future Microbiol, 10, 377–390.
- [30] Tsado AN, Lawal B, Ossai PC, Jagaba A, Gwadabe NK, Jiya AG, Umar AM and Oladunjoye JO. (2016). Antioxidants and Antimicrobial Activities of Methanol Extract of Newbouldia laevis and Crateva adansonii Journal of Pharmacy and Allied Health Sciences 6, 14-19.
- [31] Lawal B, Shittu OK, Oibiokpa IF, Mohammed H, Umar SI and Haruna GM. (2016). Antimicrobial evaluation, acute and sub- acute toxicity studies of Allium sativum. Journal of Acute Disease, 5(4), 296–301.
- [32] Ibrahim AM, Lawal B, Abubakar AN, Tsado NA, Kontagora GN, Gboke GA and Berinyuy EB. (2017). Antimicrobial and Free Radical Scavenging Potentials of N-Hexane and Ethyl-acetate Fractions of Phyllanthus Fraternus Nigerian Journal of Basic and Applied Science, 25(2), 06-11.
- [33] Yusuf AA, Lawal B, Abubakar AN, Berinyuy EB, Omonije YO, Umar SI, Shebe MN and Alhaji YM. (2018). *In-vitro* antioxidants antimicrobial and toxicological evaluation of Nigerian *Zingiber officinale*. Clinical Phytoscience, 4(12), 1-8.

#### How to cite this article

Mohammed H, Muhammad HL and Makun AH. (2019). Comparative secondary metabolite compositions and antimicrobial properties of n-hexane and ethyl-acetate fractions of *Nelsonia campestris*. GSC Biological and Pharmaceutical Sciences, 9(1), 46-52.