Phytochemical screening, total hydroxycinnamic acids, total phenolic and antioxidant activity of *Lavandula angustifolia* Mill

Dumitru Mihaela Gabriela *

*University of Craiova, Faculty of Science, Department of Chemistry, Calea București 107I, Romania.*

Publication history: Received on 10 December 2019; revised on 16 December 2019; accepted on 24 December 2019

Article DOI: https://doi.org/10.30574/gscbps.2020.10.2.0235

**Abstract**

The aim of this study was the screening of phytochemicals and determination of total hydroxycinnamic acids, total polyphenols and antioxidant activity of hydroalcoholic extract of *Lavandula angustifolia* Mill. cultivated in Craiova - Romania. Phytochemical screening showed that all of samples are positively. Quantitative determination of hydroxycinnamic acids was calculated as chlorogenic acid equivalent in % of dried plant raw material. The total polyphenol content was measured using the Folin Ciocalteu reagent colorimetric method. Antioxidant activity was determined using 2, 2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenger methods. It has established that in sample of *Lavandula angustifolia* the total content of hydroxycinnamic acid derivates is 2.65%, total polyphenols 1.4 mg GAE g⁻¹ DW. The hydroalcoholic extract displayed DPPH free radical scavenging activity with 85.7%.

**Keywords:** *Lavandula angustifolia* Mill; Phytochemical screening; Hydroxycinnamic acids; Total phenolic; DPPH

**1. Introduction**

The use of traditional medicine is widespread and plants are indeed the first source for preparing remedies in this form of alternative medicine. Plants, especially medicinal herbs, have been used for the prevention and/or treatment of several diseases since very old times [1]. Plant extracts, such as flavonoids and phenolics, have raised public interest in their potential to act as antioxidants. In a biological system, an antioxidant can be defined as “any substance that when present at low concentrations compared to that of an oxidizable substrate would significantly delay or prevent oxidation of that substrate” [1]. Natural antioxidants are widely distributed in medicinal plants. These natural antioxidants, especially polyphenols, exhibit a wide range of biological effects, including anti-inflammatory, anti-aging, anti-atherosclerosis and anticancer. The effective extraction and proper assessment of antioxidants from medicinal plants are crucial to explore the potential antioxidant sources and promote the application in pharmaceuticals.

Polyphenols of plant origin comprise a wide variety of molecules that have a polyphenol structure (i.e. several hydroxyl groups on aromatic rings), but also molecules with one phenol ring, such as phenolic acids and phenolic alcohols. Polyphenols are divided into several classes according to the number of phenol rings that they contain and to the structural elements that bind these rings to one another [2].

The main groups of polyphenols are, Fig. 1[3].

* Corresponding author
E-mail address: dumngs@yahoo.com

Copyright © 2020 Author(s) retain the copyright of this article. This article is published under the terms of the Creative Commons Attribution License 4.0
The class of compounds known as the hydroxycinnamic acids are found both covalently attached to the plant cell wall poly-saccharides and as soluble forms in the cytoplasm. The freeform occurs very rarely in plants. Hydroxycinnamic acids and their derivates all stem from cinnamic acid, and are present as four basic molecules: p-coumaric, caffeic, ferulic and sinapic acids, Fig. 2. The diversity of hydroxycinnamic acids and their resultant nutraceutical properties derives from the nature of the bonds and that of the molecule(s) involved. In addition, the presence of a double bond in the lateral chain of the hydroxycinnamic acids leads to the possible existence of two isomeric forms: cis (Z) and trans (E) [4].

The present study was performed in order to assess the activity antioxidant potential of phenolics found in the aerial parts of *Lavandula angustifolia*. Lavender belongs to the mint family Lamiaceae. While the plant is believed to have originated in Asia, it is considered native in areas ranging from the Canary Islands, to Europe, Africa, the Middle East, and parts of India. Today lavender is cultivated and grows wild in temperate zones worldwide. There are 39 species of lavender, along with numerous hybrids and cultivars. In Europe, have been used routinely as remedies: *Lavandula angustifolia* (English lavender), the closely related *Lavandula latifolia* (spike lavender), and *Lavandula stoechas* (French lavender). Lavender is a perennial shrub that from early June to late July produces flowers that are borne on spikes. The flowers range in color from white, to mauve, to purple. The mature bud, or flower spike of the lavender plant is the primary source of the chemicals [5].

Phenolic compounds is a large group of secondary plant metabolites. They can be utilized for preventing oxidative damage by scavenging free radicals, quenching singlet oxygen and reducing reactive species, chelating catalytic metal...
ions. This article focuses on identification of the chemical constituents and antioxidant properties of the hydroalcoholic extract of *Lavandula angustifolia* Mill.

### 2. Material and methods

#### 2.1. Plant material

Were used freshly picked flowers of *Lavandula angustifolia* Mill. cultivated in Craiova (Romania) in 2017. Samples have been dried in the shade after the harvest and the dry flowers were separated from their stems.

#### 2.2. Preparation of alcoholic extracts for screening

Dry flowers separated from their stems were extracted for analysis of with 100 mL of 80% aqueous methanol for 24 h in water bath at 50°C. The obtained extracts were filtered through Whatman No. 1 filter paper and then centrifuged at 4000 rpm for 5 minutes. The extracts were kept at 4°C and used for analyses.

#### 2.3. Phytochemical screening

Phytochemical screening was implemented by using the standard procedures [6].

**2.3.1. Test for tannins**

Two gram of methanolic extract was placed in a test tube. Then, 5% of ferric chloride drops were added. A bluish black or greenish coloration was observed. It was an indication of the presence of pyrogallol tannins or catechol, respectively [7, 8].

**2.3.2. Test for flavonoids (Shinoda test)**

Two gram of methanolic extract was placed in a tube. A few fragments of magnesium were added, followed by adding 0.5 mL of hydrochloric acid. The reddish color was an indication of flavonoids presence [9].

**2.3.3. Test for saponins**

Approximately 1 g of the methanolic extract was boiled with mL deionized filtered and wait for 2 min. The content was shaken vigorously. The persistent froth appearance that lasted for 15 min was an indication of saponins presence [10].

**2.3.4. Test for terpenoids (Salkowski test)**

Approximately 2 mL of chloroform was mixed with 0.5 g of the extract. Then, 3 mL of conc. H$_2$SO$_4$ was added carefully to form a layer. The red color appearance is an indication of terpenoids presence [11].

**2.3.5. Test for carbohydrates (Molisch’s test)**

Two milliliter of the methanolic extract solution was mixed with 0.2 mL of alcoholic solution of α-naphthol (10%) in a test tube and followed with an addition of 2 mL of conc. sulphuric acid by the test tube side. At the interphase of the two layers, a bluish violet zone is formed that indicates the presence of carbohydrates or/and glycosides [12].

**2.3.6. Test for cardiac glycosides (Keller-Kiliani test)**

In a test tube 2 mL of glacial acetic acid containing 1-2 drops of 2% solution of FeCl$_3$ was mixed with the methanolic extract and poured into another test tube containing 2 mL of concentrated H$_2$SO$_4$. Brown ring was formed. It was taken as presence of cardiac glycosides [9].

**2.3.7. Test for alkaloids**

One gram of methanolic extract was mixed with 2 mL of dil. HCl (1%) in a test tube. Then gently heated, followed by adding 2-3 drops of Mayer’s reagent. The formation of cream or white precipitate was an indication for the presence of alkaloids [9, 10].
2.3.8. Test for coumarins

In a test tube, 1 g of methanolic extract was placed and covered with filter paper moistened with dilute NaOH, then heated on water bath for a few minutes. The filter paper was examined under UV light. Fluorescence was detected by the UV test (365 nm), yellow fluorescence regarded as positive for the presence of coumarins [13].

2.3.9. Test for steroids (Liebermann-Burchard test)

In a test tube, 1 mL of acetic acid anhydride was added to 1 mL of methanolic extract, the solution was cooled well in ice followed by the addition of conc. sulphuric acid carefully. Appearance of color development from violet to blue or bluish-green was an indication for the presence of steroids [12].

2.3.10. Test for protein (Millon’s test)

About 1 g of methanolic extract was mixed with 2 mL of Millon’s reagent (mercuric nitrate in nitric acid containing traces of nitrous acid), white precipitate formed, which transformed to red upon gentle heating. It indicates presence of amino acids and protein [14].

2.4. Determination of hydroxycinnamic acids

Quantitative determination of hydroxycinnamic acids was calculated as chlorogenic acid equivalent in % of dried plant raw material. Maximum absorption of chlorogenic acid reference solution occurred at 327 nm, so the measurements were carried out at this wavelength by modified method [15]. About 1 g (accurate weight) flowers of *Lavandula angustifolia* was placed in 100 mL flask and 70 mL of 20% alcohol was added. The flask was joined to condenser and heated on a water bath for 1 hour. Extraction was carried out twice. Extracts were cooled, filtered through a paper filter, quantitatively transferred to a 200 mL volumetric flask. The volume of filtrate in a volumetric flask was leaded to the mark by 20% alcohol and mixed. 3 mL of solution A were placed in 50 mL volumetric flask and was leaded to the mark by 20% alcohol. Absorbance of the solution was measured on Varian Cary-50 spectrophotometer at a wavelength of 327 nm against 20% ethanol blank. Content of hydroxycinnamic acid (X, %) was calculated by a formula:

\[
X = \frac{A \cdot 200 \cdot 50 \cdot 100}{E_{1\%1cm} \cdot m \cdot 3 \cdot (100 - W)}
\]

where, A - absorption of the studied solution; \(E_{1\%1cm}\), 1 cm - specific absorption rate of chlorogenic acid, which is equal to 531; m - weight of the plant material, g; W - the loss in weight at drying, %.

2.5. Determination of total polyphenol (TP)

The total polyphenol content was measured using the Folin Ciocalteu reagent colorimetric method. To 800 µL of deionised water, 50 µL of Folin-Ciocalteu reagent and a volume of sample ranging from 10 to 50 µL were added and accurately mixed. After 1 min, 100 mL of 20% sodium carbonate solution was added and mixed. Deionised water was then added up to a volume of 1 mL. The solution was carefully mixed and total phenol content was spectrophotometrically estimated at 765 nm after 2 h incubation. The results were expressed as mg Gallic acid equivalents (GAE) g\(^{-1}\) of *Lavandula angustifolia* extracts using a standard curve, Fig. 3, generated with 10 µg, 20 µg, 40 µg 60 µg, 80 µg and 100 µg Gallic acid per mL [16, 17].
2.6. DPPH radical scavenging activity

The DPPH radical scavenging activity (SA) was determined spectrophotometrically using the DPPH method. Briefly, 1 mL of extract solution in distilled water or 1 mL of distilled water (blank) was mixed with 2 mL of DPPH solution (2 mg of DPPH was dissolved in 50 mL of methanol). The range of the investigated extract concentrations was 0.002 - 0.5 mg mL⁻¹. The mixture was shaken vigorously and left at room temperature for 30 min, then the absorbance was read at 517 nm using a spectrophotometer Varian Cary-50. The capability to scavenge the DPPH radicals (DPPH radical scavenging activity) was calculated using the following equation:

\[
SA(\%) = 100 \times \frac{(A_{\text{blank}} - A_{\text{sample}})}{A_{\text{blank}}}
\]

where: \(A_{\text{blank}}\) is the absorbance of the blank and \(A_{\text{sample}}\) is the absorbance of the sample.

3. Results and discussion

Phytochemicals are the core of phytomedicines; their therapeutic efficiency directly correlates with the presence of various phytochemicals. Table 1 shows the phytochemical screening of hydroalcoholic extract. Preliminary phytochemical screening experiments are commonly performed to promote a guidance of substantial phytochemicals that may be involved in the antioxidant activity of plant extracts [18].

Table 1 Phytochemical screening of hydroalcoholic extract of *Lavandula angustifolia*

<table>
<thead>
<tr>
<th>Phytoconstituents</th>
<th>Extract</th>
<th>Phytoconstituents</th>
<th>Extract</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tannins</td>
<td>+</td>
<td>Cardiac glucosides</td>
<td>+</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+</td>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>Saponins</td>
<td>+</td>
<td>Coumarines</td>
<td>+</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>+</td>
<td>Steroids</td>
<td>+</td>
</tr>
<tr>
<td>Carbohydrates</td>
<td>+</td>
<td>Proteins</td>
<td>+</td>
</tr>
</tbody>
</table>

As the result of the carried out investigations, the spectrophotometric method for determination of the totality of hydroxycinnamic acid derivates, expressed as chlorogenic acid. It has established that in sample of *Lavandula*
The total content of hydroxycinnamic acid derivatives is 2.65%. The results obtained in this study showed a significant level of phenolic compounds in the ethanolic extract of flowering aerial parts.

The phenolic content was expressed as Gallic acid equivalent and was deduced from the pre-established calibration curve (Fig. 1) and calculated using the following equation:

\[ Y = 0.0134x + 0.0171; \quad R^2 = 0.9981 \]

where, \( Y \) = absorbance, \( x \) = corresponding concentration (\( \mu g \cdot mL^{-1} \)) and \( R^2 \) = correlation coefficient.

The amount of total polyphenols found in the ethanolic extract of *Lavandula angustifolia* is of 1.4 mg GAE g\(^{-1}\) DW.

Polyphenols are strong antioxidants that complement and add to the functions of antioxidant vitamins and enzymes as a defense against oxidative stress caused by excess reactive oxygen species (ROS) [19]. Phenolic antioxidants were proved to interact with free radicals according to the four mechanisms [20, 21]:

- **direct hydrogen atom transfer (HAT)**
  \[ ArOH + X{\cdot} \rightarrow ArO{\cdot} + XH \]

- **electron transfer-proton transfer (ET-PT)**
  \[ ArOH + X{\cdot} \rightarrow ArOH{\cdot} + X^- \rightarrow ArO{\cdot} + XH \]

- **proton-coupled electron transfer (PCET)**
  \[ ArOH + X{\cdot} \rightarrow [ArOH{\cdot} \ldots X{\cdot}] \rightarrow [ArO{\cdot} \ldots X^-] \rightarrow ArO{\cdot} + XH \]

- **sequential proton loss electron transfer (SPLET)**
  \[ ArOH \rightarrow H^+ \rightarrow Ar^- + X^- \rightarrow ArO{\cdot} + H^+ + XH \]

Hydroxycinnamic acids are important phytochemicals possessing significant antioxidant properties. Several investigators have studied in vitro antioxidant activity of hydroxycinnamic acids in detail. Most of the reports indicated that the presence of an unsaturated bond on the side chain of hydroxycinnamic acids is vital to their activity. The structural features that were reported to be of importance to the antioxidant activity were categorized as follows: modifications of the aromatic ring, which include alterations in the number and position of hydroxy groups and insertion of electron donating or withdrawing moieties as well as modifications of the carboxylic function that include esterification and amidation process. Most of the investigations concluded that the presence of ortho-dihydroxy phenyl group (catechol moiety) is of significant importance to the antioxidant activity, while, the presence of three hydroxy groups does not necessarily improve the activity [22].

The total phenolic compounds and hydroxycinnamic acids, may play important role in determined antioxidant activity of extract obtained from *Lavandula angustifolia*.

The antioxidant activity of the *Lavandula angustifolia* extract was measured on the basis of the scavenging activity of the stable 1,1-diphenyl 2-picrylhydrazyl (DPPH) free radical. DPPH scavenging activity of *Lavandula angustifolia* was evaluated 85.7%.

The results are supported by findings from literature [23 - 26].

### 4. Conclusion

The phytochemical screening of hydroalcoholic extract of *Lavandula angustifolia* shown the presence of pharmacologically active substances such as tannins, flavonoids, saponins, terpenoids, carbohydrates, cardiac glucosides, alkaloids, coumarines, steroids and proteins. Positive high correlation between phenolic acids contents and the ability to reduce DPPH indicate that the phenolic compounds are main co-authors of the antioxidant activity of *Lavandula angustifolia*. Based on the results obtained in the present study, it is concluded that the hydroalcoholic extract...
of *Lavandula angustifolia* possess substantial amounts of phenolic compounds. Thus, hydroalcoholic extract can be considered a good source of natural antioxidants and might be used alone or as an adjunct to other drugs in medicine.

**Compliance with ethical standards**

**Acknowledgments**

The author are grateful to everyone who contributed in achieving this feat.

**Disclosure of conflict of interest**

There is no conflict of interest with publishing the present data of the study.

**References**


---

**How to cite this article**