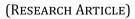


# GSC Biological and Pharmaceutical Sciences

eISSN: 2581-3250 CODEN (USA): GBPSC2 Cross Ref DOI: 10.30574/gscbps Journal homepage: https://gsconlinepress.com/journals/gscbps/





퇹 Check for updates

Lipid profile of the ethanol - methanol (1:1) extracts of *Anacadium occidentale* and *Jatropha tanjorensis* administration in Wistar rats

Saviour God's wealth Usin  $^{1,\,*}$ , Yusuff Dimeji Iybayilola<br/>  $^2$ , Unwana Ema Okon $^3$  and Oluwatoy<br/>in Omolara Daramola  $^3$ 

<sup>1</sup> Department of Medical Biochemistry, Faculty of Basic Medical Sciences, Cross River University of Technology (CRUTECH), Okuku Campus, Yala Local Government Area, Cross River State, Nigeria.

 <sup>2</sup> Department of Physiology, Faculty of Basic Medical Sciences, College of Medicine, University of Lagos, Lagos State, Nigeria.
<sup>3</sup> Department of Science Laboratory Technology, School of Engineering and Sciences, D. S. Adegbenro ICT Polytechnic, Eruku-Itori, Ewekoro Local Government Area, Ogun State, Nigeria.

GSC Biological and Pharmaceutical Sciences, 2022, 18(03), 001-010

Publication history: Received on 12 January 2022; revised on 12 February 2022; accepted on 14 February 2022

Article DOI: https://doi.org/10.30574/gscbps.2022.18.3.0030

## Abstract

Hyperlipidaemia is characterized by an increase in one or more of the plasma lipids. This study evaluated the lipid profile of albino Wistar rats administered ethanol-methanol extracts of *Anacadium occidentale* and *Jatropha tanjorensis* leaves. Twenty-five (25) male Wistar rats weighing between 180 - 220 g were divided into five (5) groups of five (5) rats each and treated thus: Group 1 (normal control received normal saline), group 2 and 3 administered low dose (400 mg/kg<sup>-1</sup> b.wt.) and high dose (800 mg/kg<sup>-1</sup> b.wt.) of *Anacadium occidentale* extract respectively, group 4 and 5 administered low dose (400 mg/kg<sup>-1</sup> b.wt.) and high dose (800 mg/kg<sup>-1</sup> b.wt.) of *Jatropha tanjorensis* extract respectively. At the end of the experiment, the rats were sacrificed to obtain the sera for the evaluation of serum lipid profile. The result revealed a significant (*P*<0.05) decrease in TC in all the groups except group IV compared with control. There was a significant (*P*<0.05) increase in TG and VLDL-c levels in all groups compared with control. HDL-c level significantly (*P*<0.05) increased in group III only compared with control. LDL-c level significant (*P*≥0.05) decreased in group III and V only compared with control. The LDL-c/HDL-c ratio showed no significant (*P*≥0.05) difference between groups II, V and control, however group III significantly (*P*<0.05) decreased and group IV significantly (*P*<0.05) increased compared with the rest groups. The study suggests that the plant extracts possess lipid lowering potentials and may be employed in the treatment of metabolic disorders such as obesity and cardiovascular diseases.

Keywords: Hyperlipidaemia; Lipid profile; Metabolic disorders; Anacadium occidentale; Jatropha tanjorensis

## 1. Introduction

Cardiovascular disease (CVD) and related disorders remains the dominant cause of death both in men and women globally [1], and while it is recognized as a multifactorial disease with many risk factors, atherosclerosis is responsible for the major pathology contributing to end stage heart disease [2]. Hyperlipidaemia is considered one of the major risk factors causing cardiovascular diseases (CVDs). CVDs accounts for one third of total deaths around the world [3, 4]. Hyperlipidaemia is a medical condition characterized by an increase in one or more of the plasma lipids, including triglycerides, cholesterol, phospholipids and or plasma lipoproteins including very low-density lipoprotein and low-density lipoprotein along with reduced high-density lipoprotein levels [5]. This elevation of plasma lipids is among the leading risk factors associated with cardiovascular diseases. In the meantime, statins and fibrates remain the major anti-

\* Corresponding author: Saviour God'swealth Usin; Email: savioladausin@gmail.com

Department of Medical Biochemistry, Faculty of Basic Medical Sciences, Cross River University of Technology (CRUTECH), Okuku Campus, Yala Local Government Area, Cross River State, Nigeria.

Copyright © 2022 Author(s) retain the copyright of this article. This article is published under the terms of the Creative Commons Attribution Liscense 4.0.

hyperlipidaemic agents for the treatment of elevated plasma cholesterol and triglycerides respectively, with severe side effects on the muscles and the liver [5]. It has been ranked as one of the greatest risk factors contributing to the prevalence and severity of coronary heart diseases. Coronary heart disease, stroke, atherosclerosis and hyperlipidaemia are the primary cause of death [6]. Hyperlipidaemia associated lipid disorders such as hypercholesterolemia and hypertriglyceridemia are considered to cause atherosclerotic cardiovascular disease [7]. The main aim of treatment in patients with hyperlipidaemia is to reduce the risk of developing ischemic heart disease or the occurrence of further cardiovascular disease or cerebrovascular disease [8]. Ineffectiveness and inability to afford synthetic drugs is a major constraint in management of hyperlipidaemia in developing countries such as Nigeria, so the search for naturally occurring locally available anti-hyperlipidaemic agent still continues.

Plants appear to be the major source of drugs for the majority of the world's population [9], with substances derived from higher plants constituting about a quarter of all prescribed medicines [10]. Several herbal medicines have advanced to clinical use in modern times [11]. It has been estimated that 25% of the modern medicines are made from plants first used traditionally. The reasons for this are complicated, probably from the ability of the plant to produce structurally diverse molecules, these molecules are made from renewable resource of raw by eco-friendly process [12]. Among several factors contributing towards the potential use of phytomedicine are safety, lack of adverse reactions and side effects which have been mostly found to particularly influence the use of such medicines in developed countries [13]. In rural areas, there are additional cultural factors that encourage the use of herbal preparations, people believe that where an area give rise to a particular disease it will also support plants that can be used to cure it, also hundreds of primary health care centres which are intended to serve rural areas which lack staff, diagnostic facilities and adequate supplies of medicines [14]. Although there is a growing popularity of herbal medicines as safe, scientists still advocate proper physiological and toxicological tests in order to ensure safety in the use of traditional medicines [15, 16].

*Jatropha tanjorensis* belongs to the family 'Euphorbiaceae' and is widely grown in southern Nigeria [17]. Its leaf is commonly consumed as vegetable in many parts of southern Nigeria. It is commonly called 'hospital too far' (Pidgin English), 'lapalapa' or 'Iyana-Ipaja' (Yoruba), 'Ugu-Oyibo' (Igbo) [18]. It is called 'Catholic vegetable or Reverend father's vegetable [19], possibly because it is grown in the premises of the catholic churches as ornamentals. *Jatropha tanjorensis* has been used locally as a source of leafy vegetable and as medicinal plant for a number of years. A study by Olayiwola *et al.*, showed that *Jatropha tanjorensis* is popular as a natural remedy against diabetes in southern Nigeria [20]. *Jatropha tanjorensis* has also been showed to exhibit antibacterial activity [18]. It is also used ethnomedically in the treatment of hypertension [19]. The leaf extract also has antioxidant property and is effective in the treatment of malaria in southern Nigeria [21]. Extracts from the plant leaves have also been used in Nigeria to control sickle cell anaemia [22]. *Jatropha tanjorensis* has received a lot of attention due to its potential health benefit, availability and affordability [23, 24]. Its primary use is for fencing, and as medicine [25]. Phytochemical screening of *Jatropha tanjorensis* leaf revealed that it contains bioactive principles such as alkaloids, flavonoids, tannins, cardiac glycoside, anthraquinones and saponins [23]. The pharmacological studies revealed that the plant showed some wide range of biological activities, which include antihypertensive, antioxidant, antimicrobial, antimalarial, hypoglycaemic, hypolipidaemic and haematological activities [25].

Anacardium occidentale L. (cashew) belongs to the family 'Anacardiaceae', which is native to Brazil [26]. This family consists of 400-600 species. In Nigeria, It is commonly called 'Kashew' (Hausa), 'Kaju' (Yoruba), 'Kausu' (Igbo), 'Shase' (Tiv) and 'Kashiwu' (Nupe) [27, 28, 29]. Anacadium occidentale has been cultured essentially, and whole fruit is used for medicinal and food purposes, e.g., apple and kernel. Anacadium occidentale gained its importance during World War II due to the utilization of its significant by-product, the Anacadium occidentale nut shell liquid [30]. Anacadium occidentale kernels have shown low-density lipoprotein cholesterol levels and coronary risk diseases [31]. Anacadium occidentale part contains proteins and fats. The proteins include lysine, cysteine, arginine tyrosine, valine, and many vitamins like vitamin C, E, and D [32]. The key component of Anacadium occidentale is anacardic acid. It is used as an antimicrobial, killing bacteria, fungi, worms and protozoa [33]. Anacadium occidentale gum has been used widely for many health-related issues. These are less in saturated fatty acids and more in unsaturated fatty acids. Its health benefits have been used to decrease the risk of cardiovascular diseases, oxidative stress, inflammation, high cholesterol, and diabetes [30]. Anacadium occidentale nuts are used for the treatment of obesity, diabetes, heart disease, urinary disorders, digestive disorders, and many other clinical applications like bone relaxation, cold and flow, etc. It also has importance in cancer and protects from aging [34]. The young and tender leaves of A. occidentale are a popular herb consumed raw as ulam and sometimes blanched to reduce their stringent taste. In traditional medicine, leaves are used for treating dysentery [35], malaria [36], vaginal douche [37], diarrhoea and piles, and an infusion of bark and leaves are applied to relief toothache and sore gums [38]. Other uses of leaves include remedy for rheumatism and hypertension [39, 40]. However, there have been several research on the uses of Anacadium occidentale and Jatropha tanjorensis in the treatment or management of certain diseases, but there is little or no research on the determination

of the  $LD_{50}$  of ethanol- methanol (1:1) extracts of these plants as well as their effects on serum lipid profile of Wistar rats, hence, the need to conduct this research.

# 2. Material and methods

#### 2.1. Chemicals/Reagents

Commercially available kit for chemical analyses like serum cholesterol, HDL-c, LDL-c and triglyceride were purchased from Biosystems S.A. (Barcelona, Spain). All other chemicals of analytical grade were obtained from Merck (Darmstadt, Germany).

## 2.2. Plant Materials

Freshly collected leaves of *Anacadium occidentale* and *Jatropha tanjorensis* were obtained from local garden at Bebi, Obanliku Local Government Area of Cross River State, Nigeria. The plants specimen were identified and authenticated in the Department of Plant Science, Cross River University of Technology (CRUTECH), Calabar, Cross River State, and the voucher numbers; CRUTECH/PSB/0045 for *Anacadium occidentale* and CRUTECH/PSB/0046 for *Jatropha tanjorensis* were deposited in the herbarium. The leaves were thoroughly washed, then air-dried at room temperature.

#### 2.3. Extraction of Plant Samples

The leaves were sorted to eliminate any dead matter and other unwanted particles. The leaves were air-dried for 21 days and then grinded in a domestic mixer grinder and coarse powder was prepared. 100g of each of the plant samples was extracted with methanol (500mls) and ethanol (500mls) in a Soxhlet extractor for 72 hours at 60°C respectively. The extract was evaporated to dryness at 40°C. The obtained extracts was in chocolate colour with aromatic odour [41].

#### 2.4. Experimental Animals

Twenty five (25) male Wistar rats weighing between 180 to 220g were used for the lipid profile study while eighteen (18) Wistar rats were used for the acute toxicity study. The animals were maintained under laboratory conditions of humidity, temperature (23 to 25°C) and 12 hours light-dark cycle in the Animal House of Department of Medical Biochemistry, Cross River University of Technology, Okuku Campus and allowed free access to standard grower's mash (Hybrid Feeds Ltd., Kaduna) and water *ad libitum*. The animals were acclimatized for two weeks.

#### 2.5. Determination of Lethal Dose (LD<sub>50</sub>) (Acute Toxicity Study)

The determination of median lethal dose ( $LD_{50}$ ) of the ethanol and methanol extract was carried out by procedure described by [42]. Eighteen (18) albino Wistar rats weighing 180 to 220 g were used. The test involved two stages. In phase I, the rats were grouped into three (3) groups of three (3) rats each. They were administered orally 10, 100 and 1000 mg/kg<sup>-1</sup> b.wt., of the extracts respectively. In the phase II, the animals were divided into three (3) groups of three (3) rats each also and administered the graded doses of 1600, 2900 and 5000 mg/kg<sup>-1</sup> b.wt of extracts and then observed for 24 hours for behaviour as well as mortality. It was calculated to be given 10% of the extracts to low doses and 20% of the extracts to high doses.

Then, the LD<sub>50</sub> was derived based on the formula:

$$LD_{50} = \sqrt{D_0 \times D_{100}}$$

Where,  $D_0$  = Highest dose that produced mortality  $D_{100}$  = Lowest dose that produced mortality

## 2.6. Experimental Design

Twenty five (25) male Wistar rats weighing 180 to 220g were used for the study while Eighteen (18) Wistar rats were used for the acute toxicity (LD<sub>50</sub>) study. After acclimatization, the animals were divide randomly into six (6) groups of three (3) rats each for phase I and II acute toxicity study. Animals for lipid profile study were divided into five (5) groups, each group containing five animals (n = 5). Group I: Normal Control; treated with normal saline; Group II: Experimental rats administered low dose (400 mg/kg<sup>-1</sup> b.wt.) of *Anacadium occidentale* extract; Group IV: Experimental rats administered high dose (800 mg/kg<sup>-1</sup> b.wt.) of *Anacadium occidentale* extract; Group IV: Experimental rats administered high dose (400 mg/kg<sup>-1</sup> b.wt.) of *Jatropha tanjorensis* extract; and Group V: Experimental rats administered high dose (800 mg/kg<sup>-1</sup> b.wt.) of *Jatropha tanjorensis* extract.

## 2.7. Animal Sacrifice and Serum Collection

At the end of the administration, the Wistar rats were weighted using weighing balance, euthanized under chloroform. The abdominal region was opened long the linear Alba, dissected using surgical blade to expose the organs. Blood sample was collected through cardiac puncture using a sterile needle. A syringe was used to collect the blood and transferred into a properly labeled plain sample bottles. It was centrifuged at 3000rpm for 10 minutes. A sterile Pasteur pipette was used to transfer the serum from the clotted blood into a serum container.

## 2.8. Lipid Profile Assays

#### 2.8.1. Determination of Total Cholesterol (TC)

Cholesterol (TC) was estimated based on the method of [43].

## 2.8.2. Determination of Triglycerides (TG)

Triglycerides (TG) was determined according to method of [44].

#### 2.8.3. Determination of High Density Lipoprotein Cholesterol (HDL-c)

High density lipoprotein cholesterol (HDL-c) concentration was estimated according to the method of [45], using Randox kit.

# 2.8.4. Determination of Low Density Lipoprotein Cholesterol (LDL-c) and Very Low Density Lipoprotein Cholesterol (VLDL-c)

Very low density lipoproteins cholesterol (VLDL-c) and low density lipoprotein cholesterol (LDL-c) were estimated or derived from TC, TG and HDL-c according to Friedewald's formula for lipids derivation [46].

#### 2.9. Statistical Analysis

Data were recorded as mean and standard error of the Mean. Statistical difference between the means was determined by one-way ANOVA using SPSS 16.0. Any significant difference between means was assessed by and P<0.05 was accepted as the significant level.

## 3. Results

The results in table 1 revealed the effect of administration of ethanol-methanol (1:1) extracts of *Anacadium occidentale* and *Jatropha tanjorensis* on serum lipid profile of Wistar rats. Following the administration of the extracts, the extracts produce a significant (P<0.05) decrease in total cholesterol (TC) of all the groups except group IV compared with the normal control (NC). Group III and IV administered high dose of *Anacadium occidentale* and low dose of *Jatropha tanjorensis* respectively produced a significant (P>0.05) decrease in TC compared with each other.

The extract produced a significant (P<0.05) increase in total triglycerides (TG) in all the groups compared with control. However, there was a significant (P<0.05) decrease in TG of the groups administered *Jatropha tanjorensis* (IV and V) compared with those administered *Anacadium occidentale* (II and III). High density lipoprotein cholesterol (HDL-c) level was significantly (P<0.05) increased in group III only compared with control. Low density lipoprotein cholesterol (LDL-c) level was significantly decreased in groups that were administered with high dose of *Anacadium occidentale* and *Jatropha tanjorensis* respectively compared with control.

Also, the extract produced a significant (P<0.05) increase in very low density lipoprotein cholesterol (VLDL-c) in all the groups compared with control. However, there was a significant (P<0.05) decrease in VLDL-c of the groups administered *Jatropha tanjorensis* (IV and V) compared with those administered *Anacadium occidentale* (II and III). The LDL-c/HDL-c ratio showed no significant (P>0.05) difference between groups II, V and the control, however group III was significantly decreased compared with the rest of the groups, conversely, that of group IV was significantly increased compared with the rest of the groups.

The acute toxicity  $(LD_{50})$  of the ethanol-methanol extract of *Anacadium occidentale* and *Jatropha tanjorensis* leaves showed no death or adverse reaction in the Wistar rats administered with various doses of the extract. However, the Wistar rats administered 5000 mg/kg<sup>-1</sup> of *Jatropha tanjorensis* showed death or adverse reaction (Table 2).

GROUPS	ТС	TG	HDL-c	LDL-c	VLDL-c	LDL-c /HDL-c
	(mg/dL)	(mg/dL)	(mg/dL)	(mg/dL)	(mg/dL)	
NC	190.36±0.80ª	123.69±1.34ª	$46.4 \pm 1.00^{a}$	168.88±1.48ª	24.92±0.27ª	$3.65 \pm 0.10^{a}$
LAO	178.40±1.43 <sup>b</sup>	224.00±2.28 <sup>b</sup>	46.6±1.32ª	177.20±2.19 <sup>b</sup>	45.20±0.28 <sup>b</sup>	$3.84 \pm 0.14^{a}$
HAO	157.60±2.59°	217.00±1.39°	53.0±1.39 <sup>b</sup>	152.20±1.93°	43.40±0.28 <sup>b</sup>	$2.88 \pm 0.04^{b}$
LJT	187.70±2.00 <sup>a</sup>	150.00±3.19 <sup>d</sup>	41.2±0.33 <sup>c</sup>	177.72±1.60 <sup>b</sup>	30.76±1.43 <sup>c</sup>	4.32±0.05 <sup>c</sup>
HJT	160.88±2.11 <sup>c</sup>	182.20±0.72 <sup>e</sup>	41.4±0.22 <sup>c</sup>	155.92±2.04 <sup>c</sup>	$36.44 \pm 0.14^{d}$	3.77±0.05ª

**Table 1** Result showing lipid profile of Wistar rats administered with Ethanol - Methanol (1:1) Extracts of Anacadiumoccidentale and Jatropha tanjorensis

Values are expressed as Mean ± SD. Identical superscript (i.e. a) means there is no significant difference between the comparing group P>0.05. Non – identical superscripts (i.e. a, b, c, d, e) means there is significance between the comparing groups at P<0.05. Legend: NC: Normal control; LAO: Low dose of *Anacadium occidentale*; HAO: High dose of *Anacadium occidentale*; LJT: Low dose of *Jatropha tanjorensis* and HJT: High dose of *Jatropha tanjorensis*.

Table 2: Phase I and II of the median lethal dose of Anacadium occidentale and Jatropha tanjorensis

(mg/kg <sup>-1</sup> b.wt)	Anacadium occidentale extracts	Jatropha tanjorensis extracts							
		Phase I							
10	0/3	0/3							
100	0/3	0/3							
1000	0/3	0/3							
·									
1600	0/3	0/3							
2900	0/3	0/3							
5000	0/3	3/3							
	100 1000 1600 2900	100     0/3       1000     0/3       1600     0/3       2900     0/3							

Values are Mean ± standard derivation (n = 3)

# 4. Discussion

Lipids are group of naturally occurring molecules that include fats, waxes, sterols, fat-soluble vitamins, monoglycerides, diglycerides, triglycerides, phospholipids and others. The main biological functions of lipids include storing energy, signaling and acting as structural component of cell membranes [47]. Lipids may be broadly defined as hydrophobic or amphiphilic small molecules [48]. Although humans and other mammals use various biosynthetic pathways to both break down and synthesize lipids, some essential lipids cannot be made this way and must be obtained from the diet [48]. In order to ensure that the body lipid concentration is normal, lipid profile test is done.

In recent years, many people have been unaware of the benefits of knowing one's profile. Lipid profile is a panel of blood tests that serves as an initial screening tool for abnormalities in lipids, such as cholesterol and triglycerides. There are two common concerns people have about lipids in their diet. One is their high caloric level which may result in undesirable weight gain. The other is their association with high cholesterol level which is a risk factor for cardiovascular diseases [49]. The effect of ethanol-methanol extracts of *Anacadium occidentale* and *Jatropha tanjorensis* on total cholesterol, triglycerides, high density lipoprotein cholesterol, low density lipoprotein cholesterol, and very low-density lipoprotein cholesterol of Wistar rats were investigated. The non-toxicity of the ethanol-methanol extracts of *Anacadium occidentale* leaves was observed up to 5000 mg/kg<sup>-1</sup> b.wt (highest dose), suggesting the safety of the extracts for human and animal consumption and complements. However, non-toxicity was observed in ethanol-methanol extracts of *Jatropha tanjorensis* leaves up to 2900 mg/kg<sup>-1</sup>b.wt. But Wistar rats administered 5000 mg/kg<sup>-1</sup> b.wt of *Jatropha tanjorensis* died, indicating that *Jatropha tanjorensis* is highly toxic than *Anacadium occidentale*.

Cholesterol is the principal sterol synthesized by all animals and occurs mainly in the cell membrane due to its amphipathic nature [50]. Its synthesizes begins with the mevalonate or HMG-CoA reductase pathway, the target of statin drugs, which encompasses the first 18 steps, then followed by 19 additional steps to convert the resulting lanosterol into cholesterol via either of two pathways, the Bloch Pathway, or the Kandutsch-Russell Pathway [51, 52]. It is reportedly a major cause of cardiovascular derangements such as atherosclerosis, myocardial infarction and coronary heart diseases [1]. In this study, the plant extracts produced a decreased in serum cholesterol which might be due to a reduce absorption from the intestine by binding with bile acid within the intestine and increasing bile acid secretion [53], or due to the presence of saponins, a phytochemical which forms insoluble complexes with cholesterol or their bile salt precursor, thus making them unavailable for absorption [54]. Therefore, it implies that the plant extracts possess anticholesterolaemic activities.

Triglyceride is an ester derived from glycerol and three fatty acids and the most common type of lipid in the body. Triglycerides are the main constituents of body fat in humans and other vertebrates, as well as vegetable fat [50]. They are also present in the blood to enable the bidirectional transference of adipose fat and blood glucose from the liver, and are a major component of human skin oils [55]. It is not cholesterol but it is measured because when it is high and high density lipoprotein cholesterol (HDL-c) is low, it may results in atherosclerosis and coronary heart diseases [1, 56]. Hypertriglyceridemia is a high level of triglyceride in the blood and could results in cardiovascular disease [57]. In this study, it was observed in all the test groups that the plant extracts elevates triglyceride levels.

Low density lipoprotein cholesterol (LDL-c) transports cholesterol form the liver to the exact site where it is going to be utilized. If there is excess of LDL-cholesterol, it may initiate the process of atherosclerosis [54]. It transports about 60-70 % of total cholesterol. Therefore, an increase in TC level consequently increase LDL-c [58]. The plant extracts administered at high doses appeared to have a decreased in serum LDL-c level, hence a non-predisposition to atherosclerosis and other cardiovascular related diseases. Atherosclerosis narrows the area where blood flows through the vessels. This reduces the supplied with the blood, and it is perfect place for clot formation. If there is too much of LDL-cholesterol, it can lead to many other illness such as angina, coronary heart diseases, heart attacks, stroke and hypercholesterolemia [1, 59].

High density lipoprotein cholesterol (HDL-c) is an anti-atherogenic lipoprotein which transports cholesterol from peripheral tissues back to the liver where it is broken down to bile acids [60, 61, 62], as revealed in the group that was administered with high dose of *Anacadium occidentale* only. The inhibition of HMG-CoA reductase (a microsomal enzyme which catalyze the rate of limiting step in cholesterol synthesis pathway), reduces LDL-c and concurrently increase HDL-c [63]. Increased level of HDL-cholesterol observed is associated with healthy heart thereby reduce risk for cardiovascular diseases development and related complications such as stroke, myocardial infarction and death [64, 65]. Also, this could possibly be due to increasing activity of lecithin-cholesterol acyl transferase (LCAT), an enzyme responsible for incorporating free cholesterol into HDL-c [66], thereby promoting reverse cholesterol transport and competitively inhibiting the uptake of LDL-c by endothelial cells and preventing the generation of oxidized LDL-c [67]. Previous studies revealed that one out of three deaths would be due to cardiovascular disease and the prevailing factors remain elevated levels of serum total cholesterol (TC), low density lipoprotein cholesterol (LDL-c), triglyceride (TG) and decreased level of high density lipoprotein cholesterol (HDL-c) [68, 69]. These Prevailing factors predisposing to cardiovascular disease was not observed in the study. The effect was dose dependent with respect to *Anacadium occidentale*.

Moreover, the LDL-c/HDL-c ratio is often used as an index for cardiovascular disorders [70, 71, 72], and in this study the LDL-c/HDL-c ratio in groups that were administered low dose of *Anacadium occidentale* and high dose of *Jatropha tanjorensis* showed non-significant difference compared to the control. However, the group that was administered high dose of *Anacadium occidentale* revealed a decreased compared to the control, the reverse was the case in the group administered with low dose of *Jatropha tanjorensis*. This suggests the anti-atherogenic potential of the *Anacadium occidentale* and *Jatropha tanjorensis* extract, however the effect is dose-dependent and revealed more effective when high dose of *Anacadium occidentale* is administered.

# 5. Conclusion

The study suggests that the plant extracts exhibit lipid lowering effects which could be employed in the treatment of metabolic disorders such as atherosclerosis and cardiovascular diseases by the inhibition of biosynthesis, absorption and secretion of lipids, which may be possibly due to the presence of secondary metabolites in the plants used. However, further research is needed to investigate the anti-hyperlipidaemic components in *Anacadium occidentale* and *Jatropha tanjorensis* and their mechanism of actions.

## **Compliance with ethical standards**

#### Acknowledgments

We are grateful to the Department of Medical Biochemistry, Cross River University of Technology, Okuku Campus for the infrastructural facilities and access to scientific instrumentation. We acknowledge Unique Analytical and Diagnostic Laboratories Services Limited, Phase IV, Kubwa, Abuja for assisting with the lipid profile analysis.

#### Disclosure of conflict of interest

The authors declare that there are no conflict of interests.

#### Statement of ethical approval

The research study was carried out according to the guidelines approved by CRUTECH Institutional Research Ethical Committee (IREC) following the principle laid down in the Declaration of Helsinki (1964), as revised in 2013 and National Institute of Health (NIH) Principles of Laboratory Animal Care. No human participants were involved in the study.

#### References

- [1] Ighodaro OM, Omole JO. Effects of Nigerian Piliostigma thonningii species leaf extract on lipid profile in Wistar rats. International Scholarly Research Notices (Pharmacology). 2012; 387942.
- [2] Lefkowitz RJ, Willerson JT. Prospects for cardiovascular research. Journal of the American Medical Association. 2001; 285(5): 581–587.
- [3] Ginghina C, Bejan I, Ceck CD. Modern risk stratification in coronary heart disease. Journal of Medical Life. 2011; 4(4): 377-386.
- [4] Jorgensen T, Capewell S, Prescott E, Allender S, Sans S, Zdrojewski T. Population-level changes to promote cardiovascular health. European Journal of Preventive Cardiology. 2013; 20(3): 409-421.
- [5] Shattat GF. A review article on hyperlipidemia: Types, treatments and new drug targets. Biomedical and Pharmacology Journal. 2014; 7(2): 7-11.
- [6] Mishra PR, Panda PK, Apanna KC, Panigrahi S. Evaluation of acute hypolipidemic activity of different plant extracts in Triton WR-1339 induced hyperlipidemia in albino rats. Pharmacologyonline. 2011; 3: 925-934.
- [7] Brouwers MC, VanGreevenbroek MM, Stehouwer CD, de Graaf J, Stalenhoef AF. The genetics of familial combined hyperlipidaemia. Nature Reviews in Endocrinology. 2012; 8(6): 352-362.
- [8] Davey SG, Pekkanen J. Should there be a moratorium on the use of cholesterol lowering drugs? British Medical Journal. 1992; 304(6824): 431–440.
- [9] Oluyemi KA, Omotuyi IO, Jimoh OR. Erythropoietic and anti-obesity effects of Garcinia cambogia (bitter kola) in Wistar rats. Biotechnology and Applied Biochemistry. 2007; 46(1): 69–72.
- [10] Kumar S, Kumar R, Khan A. Medicinal Plant Resources: Manifestation and Prospects of Life sustaining Healthcare System. Continental Journal of Biological Sciences. 2011; 4(1): 19–29.
- [11] Mahmood ZA, Sualeh M, Mahmood SB. Herbal treatment for cardiovascular disease the evidence based therapy. Pakistan Journal of Pharmaceutical Sciences. 2010; 23(1): 119–124.
- [12] Kamboj VP. Herbal medicine. Current Science. 2000; 78(1): 35-39.
- [13] Renckens CNM, Dorlo TPC. Please, let not Western quackery replace traditional medicine in Africa. Tropical Medicine and International Health. 2013; 18(2): 242-244.
- [14] Pal SK, Shukla Y. Herbal medicine: Current status and the future. Asian Pacific Journal of Cancer Prevention. 2003; 4(4): 281-288.
- [15] Ozolua RI, Eriyamremu GE, Okene EO, Ochei U. Hypoglyceamic effects of viscous preparation of lrvingia gabonensis (Dikanut) seeds in streptozotocin induced Wistar rats. Journal of Herbs, Species and Medicinal Plants. 2006; 12: 1-9.

- [16] Oyewole IO, Magaji ZJ, Awoyinka OA. Biochemical and toxicological studies of aqueous extract of Tithonia diversifolia leaves in Wistar albino rats. Journal of Medicinal Plants Research. 2007; 1(2): 30-33.
- [17] Sofowora A. Medicinal Plants and Traditional Medicine in Africa. 2nd Edition. Spectrum Books Ltd., Ibadan. Nigeria. 1993; 289.
- [18] Iwalewa EO, Adewunmi CO, Omisore NO, Adebanji OA, Azike CK. Pro-and antioxidant effects and cytoprotective potentials of nine edible vegetables in Southwest Nigeria. Journal of Medicinal Foods. 2005; 8(4): 539-544.
- [19] Omoregie ES, Osagie AU. Antioxidant properties of methanolic extract of some Nigerian plants on nutritionallystressed rats. Nigerian Journal of Basic and Applied Science. 2012; 20(1): 17-20.
- [20] Olayiwola G, Iwalewa EO, Omobuwajo OR, Adebajo AC, Adeniyi AA, Verspohl EJ. The antidiabetic potential of Jatropha tanjorensis leaves. Nigerian Journal of Natural Products and Medicine. 2004; 8: 55-58.
- [21] Oluwole IO, Oluwaseun TO, Bukola VA. Assessment of renal and hepatic functions in rats administered methanolic leaf extract of J. tanjorensis. Annals of Biological Research. 2012; 3(2): 837-41.
- [22] Iloudu EM, Enwa FO. Commonly used medicinal plants in the management of sickle cell anaemia and diabetes mellitus by the local people of Edo State, Nigeria. International Journal of Pharmaceutical, Biological and Chemical Sciences. 2013; 2(2): 14-19.
- [23] Omoregie ES, Osagie AU. Phytochemical screening and antianaemic effect of Jatropha tanjorensis leaf in protein malnourished rats. Plants Archives. 2007; 7: 509-516.
- [24] Omobuwajo OR, Alade GO, Akanmu MA, Obutor EM, Osasan SA. Microscopic and toxicity studies on the leaves of J. tanjorensis. African Journal of Pharmacy and Pharmacology. 2011; 5(1): 12-17.
- [25] Oboh FOJ, Masodje HI. Nutritional and antimicrobial properties of Jatropha tanjorensis leaves. American-Eurasian Journal of Scientific Research. 2009; 4(1): 7-10.
- [26] Lim TK. Anacardium occidentale. In: Edible Medicinal and Non-medicinal Plants, Vol. I, Fruits. Dordrecht, Heidelberg, London and New York: Springer Science and Business Media BV. 2012; 45-68.
- [27] Saganuwan SA. A photo album of some medicinal plants of the Nigerian middle belt. Journal of Herbs, Spices and Medicinal Plants. 2010; 16(3): 219-292.
- [28] Mann A, Gbate M, Nda Umar A. Medicinal and economic plants of Nupeland. Jube Evans Books and Publication, Bida, Nigeria. 2003; 3-276.
- [29] Offiah NV, Makama S, Elisha IL. Ethnobotanical survey of medicinal plants used in the treatment of animal diarrhoea in Plateau State, Nigeria. BMC Veterinary Research. 2011; 7: 36.
- [30] Cardoso BR, Duarte GBS, Reis BZ, Cozzolino SMF. Brazil nuts: Nutritional composition, health benefits and safety aspects. Food Research International. 2017; 100: 9-18.
- [31] Gómez-Caravaca AM, Verardo V, Caboni MF. Chromatographic techniques for the determination of alkyl-phenols, tocopherols and other minor polar compounds in raw and roasted cold pressed cashew nut oils. Journal of Chromatography A. 2010; 1217: 7411–7417.
- [32] Bes-Rastrollo M, Sabaté J, Gomez-Gracia E, Alonso A, Martinez JA. Nut consumption and weight gain in a Mediterranean cohort: the SUN study. Obesity. 2007; 15(1): 107-116.
- [33] Tan YP, Chan EWC. Antioxidant, antityrosinase and antibacterial properties of fresh and processed leaves of Anacardium occidentale and Piper betle. Food Biosciences. 2014; 6: 17-23.
- [34] Davis CD. Low dietary copper increases fecal free radical production, fecal water alkaline phosphatase activity and cytotoxicity in healthy men. Journal of Nutrition. 2003; 133(2): 522-527.
- [35] Santos FO, Da Costa JGM, Rodrigues FF, Rodrigues OG, De Medeiros RS. Antibacterial evaluation of Anacardium occidentale (Linn) (Anacardiaceae) in semiarid Brazil. African Journal of Biotechnology. 2013; 12: 4836-4840.
- [36] Sokeng SK, Kamtchouing P, Watcho P, Jatsa HB, Moundipa PF, Lontsi D. Hypoglycemic effect of Anacardium occidentale L. methanol extract and fractions on streptozotocin-induced diabetic rats. Global Journal of Pharmacology. 2007; 1: 1-5.
- [37] Trevisan MTS, Pfundstein B, Haubner R, Wurtele G, Spiegelhalder B, Bartsch H, Owen RW. Characterization of alkyl phenols in cashew (Anacardium occidentale) products and assay of their antioxidant capacity. Food Chemistry and Toxicology. 2006; 44: 188-197.

- [38] Akinpelu DA. Antimicrobial activity of Anacardium occidentale bark. Fitoterapia. 2001; 72: 286-287.
- [39] Andarwulan N, Kurniasih D, Apriady RA, Rahmat H, Roto AV, Bolling BW. Polyphenols, carotenoids, and ascorbic acid in underutilized medicinal vegetables. Journal of Functional Foods. 2012; 4: 339-347.
- [40] Nugroho AE, Malik A, Pramono S. Total phenolic and flavonoid contents and in vitro anti-hypertension activity of purified extract of Indonesian cashew leaves (Anacardium occidentale L.). International Food Research Journal. 2013; 20: 299-305.
- [41] Olagunju JA, Adeneye AA, Fagbohunk BS, Bisuga NA, Ketiku AO, Benebo AS, Olufowobi OM, Adeoye AG, Alimi MA, Adeleke AG. Nephroprotective activities of the aqueous seed extract of Carica papaya Linn. In carbon tetrachloride induced renal injured Wistar rats: a dose and time-dependent study. Biology and Medicine. 2009; 1(1): 11-19.
- [42] Lorke D. A new approach to practical acute toxicity testing. Archives of Toxicology. 1983; 54: 275-287.
- [43] Roeschlau P, Bernt E, Gruber WA. Enzymatic termination of total cholesterol. Clinical Biochemistry. 1974; 12(5): 226–226.
- [44] Tietz NW. Fundamental of Clinical Chemistry. W. B. Saunders Company, Philadelphia. 1986; 723.
- [45] Albers JJ, Warnick GR, Chenng, MC. Quantitation of high density lipoproteins. Lipids. 1978; 13(12): 926-932.
- [46] Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clinical Chemistry. 1972; 18(6): 499–502.
- [47] Akoh CC. Handbook of Functional Lipids. CRC Press, Athens, USA: the University of Georgia. 1994; 544.
- [48] Sidhu D, Naugler C. Fasting time and lipid levels in a community-based population: A cross-sectional study/fasting time and lipid levels. Archives of Internal Medicine. 2012; 172(22): 1–4.
- [49] Khovidhunkit W, Kim M, Memon R, Shigenaga J, Moser A. Effects of infection and inflammation on lipid and lipoprotein metabolism: Mechanisms and consequences to the host. Journal of Lipid Research. 2004; 45: 1169-1196.
- [50] Lehninger AL, Nelson DL, Cox MM. Lehninger Principles of Biochemistry (3rd edition). Worth Publishers, New York, USA; 2000.
- [51] Rhodes CM, Stryer L, Tasker R. Biochemistry (4th ed.). San Francisco: W.H. Freeman. 1995; 280.
- [52] Singh P, Saxena R, Srinivas G, Pande G, Chattopadhyay A. Cholesterol biosynthesis and homeostasis in regulation of the cell cycle. PLOS ONE. 2013; 8(3): e58833.
- [53] Dasofunjo K, Nwodo OFC, Johnson JT, Ukpanukpong RU, Ugwu MN, Ayo VI. Phytochemical screening and effect of ethanolic leaf extract of Piliostigma thonningii on serum lipid profile of male albino rats. International Journal of Plant Production. 2013; 3(2): 5-9.
- [54] Beckmann N, Cannet C, Babin AL, Blé FX, Zurbruegg S, Kneuer R, Dousset V. In vivo visualization of macrophage infiltration and activity in inflammation using magnetic resonance imaging. Wiley interdisciplinary reviews. Nanomedicine and Nanobiotechnology. 2009; 1(3): 272–298.
- [55] Lampe MA, Burlingame AL, Whitney J, Williams ML, Brown BE, Roitman E, Elias M. Human stratum corneum lipids: characterization and regional variations. Journal of Lipid Research. 1983; 24(2): 120–130.
- [56] Eisenhaver LA, Nicholes LW, Spencer RT, Bergan FW. Clinical Pharmacology and Nursing Management. Lippincott, Philadelphia, USA. 1998.
- [57] Wang L, Sun H, Pan B, Zhu J, Huang G, Huang X, Tian J. Inhibition of histone acetylation by curcumin reduces alcohol-induced expression of heart development – related transcription factors in cardiac progenitor cells. Biochemical and Biophysical Research Communications. 2012; 424: 593-596.
- [58] Sheneni VD, Odiba VA, Omede A, Idih FM. Anti-hyperlipidemic effect of Vitex doniana in poloxamer induced hyperlipidemia. MedCrave Online Journal of Biology and Medicine. 2018; 3(4): 168–173.
- [59] Ramarathnan N, Ochi H, Takeuchi M. Antioxidant defense system in vegetable extracts. In natural antioxidants: Chemistry, health effects, and application. Shahidi, F, Ed, AOCS Press: Champaign, IL. 2007; 76-87.
- [60] Shirwaikar A, Rajendran K, Kumar CD, Bodla R. Antidiabetic activity of aqueous leaf extract of Annona squamosa in streptozotocin-nicotinamide type 2 diabetic rats. Journal of Ethnopharmacology. 2004; 91: 171-175.

- [61] Lewis GF, Rader DJ. New insights into the regulation of HDL metabolism and reverse cholesterol transport. Circulation Research. 2005; 96(12): 1221–1232.
- [62] Kim HY, Jeong DM, Jung HJ, Jung, YJ, Yokozawa T, Choi JS. Hypolipidemic effects of Sophora flavescens and its constituents in poloxamer 407-induced hyperlipidemic and cholesterol-fed rats. Biological and Pharmaceutical Bulletin. 2008; 31(1): 73–78.
- [63] Diepen JA, Vroegrijk IOCM, Berbée JFP, Shoelson SE, Romijn JA, Havekes LM. Aspirin reduces hypertriglyceridemia by lowering VLDL-triglyceride production in mice fed a high-fat diet. American Journal of Physiology-Endocrinology and Metabolism. 2011; 301(2011): E1099-E1107.
- [64] Gordon DJ, Probstfield JL, Garrison RJ. High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. Circulation. 1989; 79(1): 8–15.
- [65] Singh V, Sharma R, Kumar A, Deedwania P. Low high-density lipoprotein cholesterol: Current status and future strategies for management. Vascular Health and Risk Management. 2010; 6(1): 979–996.
- [66] Geetha G, Kalavalarasariel GP, Sankar V. Anti-diabetic effect of Achyranthes rubrofusca leaf extracts on alloxan induced diabetic rats. Pakistan Journal of Pharmaceutical Sciences. 2011; 24(2): 193–199.
- [67] Yokozawa T, Cho EJ, Sasaki S, Satoh A, Okamoto T, Sei Y. The protective role of Chinese prescription Kangenkaryu extract on diet-induced hypercholesterolemia in rats. Biological and Pharmaceutical Bulletin. 2006; 29(4): 760–765.
- [68] Magnus P, Beaglehole R. The real contribution of the major risk factors to the coronary epidemics: time to end the 'only 50%' claim. Archives of Internal Medicine. 2001; 161: 2657–2660.
- [69] Kumar A, Dhaliya SA, Surya AS, Dawn VT, Carla B, Sunil C. A review of hyperlipidemia and medicinal plants. International Journal of Applied Pharmaceutical Science. 2013; 2(4): 219-237.
- [70] Panagiotakos DB, Pitsavos C, Skoumas J. Importance of LDL/HDL cholesterol ratio as a predictor for coronary heart disease events in patients with heterozygous familial hypercholesterolaemia: a 15-year follow-up (1987– 2002). Current Medical Research and Opinion. 2003; 19(2): 89–94.
- [71] Nicholls SJ, Tuzcu EM, Sipahi I, Grasso AW, Schoenhagen P, Hu T, Wolski K, Crowe T, Desai MY, Hazen SL, Kapadia SR, Nissen SE. Statins, high-density lipoprotein cholesterol, and regression of coronary atherosclerosis. Journal of American Medical Association. 2007; 297(5): 499–508.
- [72] Kastelein JJ, van der Steeg WA, Holme I, Gaffney M, Cater NB, Barter P, Deedwania P, Olsson AG, Boekholdt SM, Demicco DA, Szarek M, LaRosa JC, Pedersen TR, Grundy SM, TNT Study Group, IDEAL Study Group. Lipids, apolipoproteins, and their ratios in relation to cardiovascular events with statin treatment. Circulation. 2008; 117(23): 3002–3009.