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(RESEARCH ARTICLE)



Antimicrobial susceptibility pattern of urinary isolates from outpatients suspected for urinary tract infection

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Abstract

Urinary tract infection (UTI) is one of the most common bacterial infections in humans and a major cause of morbidity. The pathogens traditionally associated with UTI and their antibiotic sensitivity patterns are changing from time to time and across different environment. Knowledge of the antibiotic resistance patterns of uropathogens in specific geographical locations is an important factor for choosing an appropriate empirical antimicrobial treatment. This study therefore evaluates the causative organisms present in urine specimen and their antibiotic susceptibility profile among patients suspected for UTI attending the general outpatient department clinic of the University of Port Harcourt Teaching Hospital, Nigeria. One hundred and fifty (150) mid-stream urine samples were collected from patients suspected of having a UTI and subjected to macroscopic assessment, isolation, and characterization as well as resistance- susceptibility test of isolates using standard conventional techniques. Results showed that UTI was frequently encountered in females 39 (26%) than males 21 (14%) while 60 % of the samples yielded no growth after 48 hours incubation. The most common pathogens isolated were *Staphylococcus aureus* (47.19%), followed by *Escherichia coli* (20.22%), *Klebsiella pneumoniae* (15.73%), *Candida albicans* (10.11%), *Pseudomonas aeruginosa* (5.62%) and *Proteus mirabilis* (1.12%). *Staphylococcus aureus* isolates were highly susceptible to Amoxicillin/Clavulanate (88.10%) with lower susceptibility to Ofloxacin (52%), Cloxacillin (50%). This information will directly affect selection of empirical therapy for UTI and emphasizes the need for choosing an appropriate antimicrobial treatment in specific geographical locations.

Keywords: Urinary tract infection; Pathogens; Susceptibility; Resistance; Antimicrobial; Etiology

1. Introduction

Urinary tract infections (UTIs) consists of microbial invasion and multiplication in any of the structures of the urinary system with attendant signs of inflammation. Terms such as significant bacteriuria, asymptomatic bacteriuria, cystitis, urethral syndrome, acute or chronic pyelonephritis have also been used to describe this infection [1]. Infection may occur at any part of the genitourinary tract, including urethra, bladder, ureter, renal pelvis, or renal parenchyma [2]. This tract collects and stores urine and provides the urinary system of tubes necessary to release it from the body.

UTI can be asymptomatic or symptomatic, characterized by a wide spectrum of symptoms ranging from mild irritative voiding to bacteremia, sepsis or even death. Other major symptoms may include: strong urge to urinate frequently, even immediately after the bladder is emptied (Urgency), painful burning sensation when urinating (Dysuria), discomfort, pressure, or bloating in the lower abdomen, pain in the pelvic area or back, cloudy or bloody urine (Haematuria), which may have a strong smell, urination during the night (Nocturia), frequent urination (Polyuria) [3].

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UTI represents one of the most common diseases encountered in medical practice today with an estimated 150 million UTIs per annum worldwide [4]. Although UTIs occur in both men and women, studies suggest that the overall prevalence of UTI is higher in women due to their anatomy [5, 6]. Some factors that may contribute to urinary tract infections are, structure of the female urinary tract, sexual intercourse, irregular urination, birth control methods, inadequate personal hygiene, history of previous UTIs, insufficient water intake, catheters or tubes placed in the bladder [6, 7]. While up to 90 % of the patients with UTIs complain of urinary tract symptoms, one third or more of the patients with these symptoms do not have bacteriuria [8]. The most common symptoms for which most patients seek treatment are dysuria and frequency. Dysuria and frequency together raise the probability of UTI to more than 90 %, effectively ruling in the diagnosis by history alone [9]. The etiology of UTI is related to the great diversity of microbial invaders, such as: bacteria, viruses and fungi. According to Ramesh *et al.* [10] bacteria are the major causative organism and are responsible for more than 95 % of UTI cases. Several studies on the prevalence of bacterial isolates from suspected urinary tract infection patients have shown that the commonest isolates were species of *Escherichia*, *Staphylococcus*, *Klebsiella*, *Pseudomonas* and *Proteus* [11-16]. In acute uncomplicated UTI acquired in the community, *Escherichia coli*, is by far the most common causative bacteria being responsible for about 80 percent of infections [11, 17, 18]. The remaining 20 percent is caused by other gram-negative enteric bacteria such as *Klebsiella* and *Proteus* species, and by gram positive cocci particularly *Enterococci* and *Staphylococcus saprophyticus* [11]. In all Gram-negative isolates formed a major constituent of bacterial uropathogens [11].

Antibiotics usually are the first line treatment for urinary tract infections. However, treatment is often started empirically, and therapy is based on information determined from the antimicrobial resistance pattern of the urinary pathogens [19]. The commonly prescribed ones include Levofloxacin, Ciprofloxacin, Cephalexin, Trimethoprim/sulfamethoxazole, Nitrofurantoin, Fosfomycin, Trimethoprim and Nalidixic acid [12, 15, 20]. Unfortunately, the extensive use of these antimicrobial agents has invariably resulted in the development of antibiotic resistance, which, has now become a major problem worldwide [21-23]. The increase in multidrug resistance in bacterial uropathogens is an important and evolving public challenge [24, 25]. The etiology of UTI and antibiotic resistance of uropathogens have been changing over past years, both in community and nosocomial infection [26-28]. Although several susceptibility testing studies have been undertaken, but not much seems to have been done on the resistance pattern of community acquired UTI pathogens. This study therefore evaluates the causative organisms present in urine specimen and their antibiotic susceptibility profile among patients suspected for UTI attending the General Outpatient department (GOPD) clinic of the University of Port Harcourt Teaching Hospital, Nigeria. The area-specific monitoring studies aimed at gaining knowledge about the type of pathogens responsible for urinary tract infections and their resistance patterns may help the clinician make inform decisions as previously reported [29].

2. Material and methods

2.1. Sample population, collection and processing

The study population was drawn from patients suspected for urinary tract infection and attending the General Outpatient Department (GOPD) clinic, University of Port Harcourt Teaching Hospital, Rivers State. A total of one hundred and fifty (150) early morning mid-stream urine specimens were collected, in sterile, dry, wide-mouth leak proof containers. The samples were labeled and transported immediately to pharmaceutical microbiology laboratory, University of Port Harcourt for examination.

2.2. Ethical considerations

This study was approved by the research and ethics committee of University of Port Harcourt Teaching Hospital, Rivers State. All samples were collected following voluntary informed consent of the participants and in some cases, their legal guardian. All patient's data and bacterial isolates gathered in this study were handled confidentially and in accordance with approved protocols of handling patient data.

2.3. Macroscopic examination of urine

The urine samples were physically examined by swirling or inverting the bottles for the presence or absence of cloudiness or turbidity. Turbidity would indicate the presence of bacteria, proteins, crystals, leucocytes, precipitation of urates (acids) or phosphates and carbonates (alkaline).

2.4. Urine culture

Using a standard wire loop, a loopful of each urine sample was inoculated on to Cystine Lactose Electrolyte Deficient (CLED) agar (a non-selective medium capable of supporting the growth of most urinary pathogens and giving good

colonial differentiation without spread of *Proteus* spp) (Lab M limited, U.K), and incubated at 37 °C for 24 hours [30]. Plates that showed significant growth were selected and the colonies aseptically isolated and sub-cultured onto Mannitol Salt Agar (Lab M limited, U.K), MacConkey Agar (Lab M limited, U.K), Sabouraud Dextrose Agar or Cetrimide agar and incubated at 37 °C for 24–48 hours.

2.5. Identification of isolates

After overnight incubation at 37 °C for 24–48 hours colonies were identified and characterized using colony characteristics, gram reaction of the organisms and biochemical test following standard procedure [31].

2.6. Antibiotic sensitivity testing

Sensitivity testing was performed following modified Kirby-Bauer disc diffusion method as recommended by National Committee for Clinical Laboratory Standards (NCCLS) [32]. Discs containing the following antibacterial agents were used: Amoxicillin/Clavulanate 30 µg, Ofloxacin 5 µg, Ciprofloxacin 5 µg, Gentamicin 10 µg, Cefuroxime 30 µg, Ceftazidime 30 µg, Ampicillin 10 µg, Nitrofurantoin 300 µg against gram negative bacteria and Amoxicillin/Clavulanate 30 µg, Ofloxacin 5 µg, Cloxacillin 5 µg, Erythromycin 5 µg, Ceftriaxone 30 µg, Gentamicin 10 µg, Cefuroxime 30 µg, Ceftazidime 30 µg against gram positive bacteria. Standard Ketoconazole solution (0.25%) was tested against *Candida albicans*. The plates were incubated at 37 °C for 24 hours after which the zones of inhibition were measured in millimeter. Using the interpretative chart derived from the zones of inhibition of Standard organisms according to the Clinical Laboratory Science Institute (CLSI), the zone size of each antimicrobial agent was interpreted [32].

2.7. Statistical analysis

Data collected were entered into Microsoft excel spreadsheet and analysed using T-test, Anova (Single factor) to calculate probabilities and determine significance. A p-value of less than or equal to 0.05 ($p \leq 0.05$) is considered to be statistically significant [22].

3. Results

3.1. Isolation and identification of isolates

Out of the 150 samples obtained from the patients suspected for UTI, 60 samples (40%) were found to contain heavy and appreciable bacterial growth (significant bacteriuria) while 90 (60%) had no appreciable bacterial growth (Figure 1). From the 60 samples investigated, 89 isolates were recovered with females having a greater percentage of occurrence 39 (26%) than males 21 (14%). The predominant isolate was *Staphylococcus aureus* constituting 42 (47.19%) of the total isolates followed by *Escherichia coli* 18 (20.22%), *Klebsiella pneumoniae* 14 (15.73%), *Pseudomonas aeruginosa* 5 (5.62%) and *Proteus mirabilis* 1 (1.12%). Fungal isolate as *Candida albicans* accounted for 9 (10.11%) and the result presented in Figure 2.

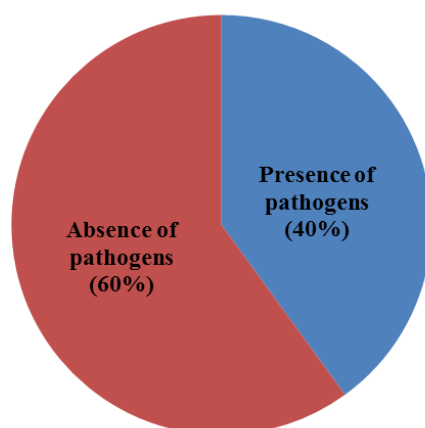


Figure 1 Percentage distribution for primary isolation of causative agents

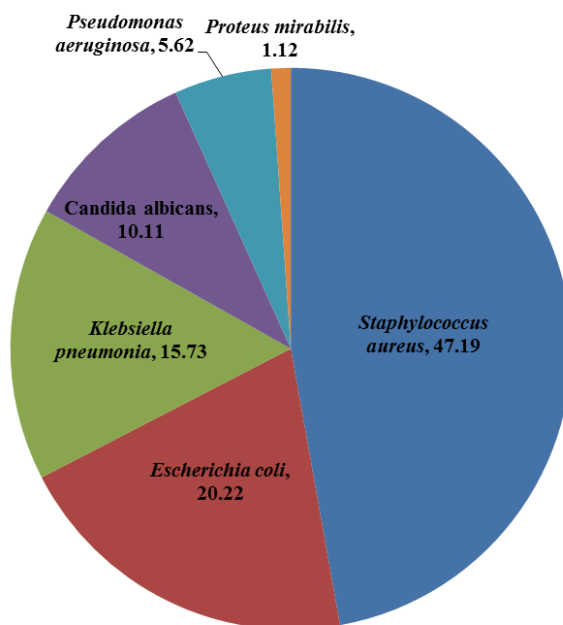


Figure 2 Percentage distribution of urinary isolates from samples

3.2. Antimicrobial susceptibility test

In-vitro sensitivity test for antimicrobial agents on Muller Hinton agar using the Kirby-Bauer disk diffusion method were carried out for the representative isolates, which include Gram-positive organism, Gram-negative organisms and fungi organism. The antibiotic susceptibility test results show that the Gram-positive organism (*Staphylococcus aureus*), are susceptible to Amoxicillin/Clavulanate (52.38%), Ofloxacin (52%) and Cloxacillin (45.24%) (Figure 3). The Gram-negative organisms were highly susceptible to Nitrofurantoin than other antibiotics used (Figures 4 - 7) while *Candida albicans* (77.78%) showed a good susceptibility to Standard Ketoconazole (Figure 8). Generally, there was a notable resistance to Amoxicillin/Clavulanate and Ampicillin.

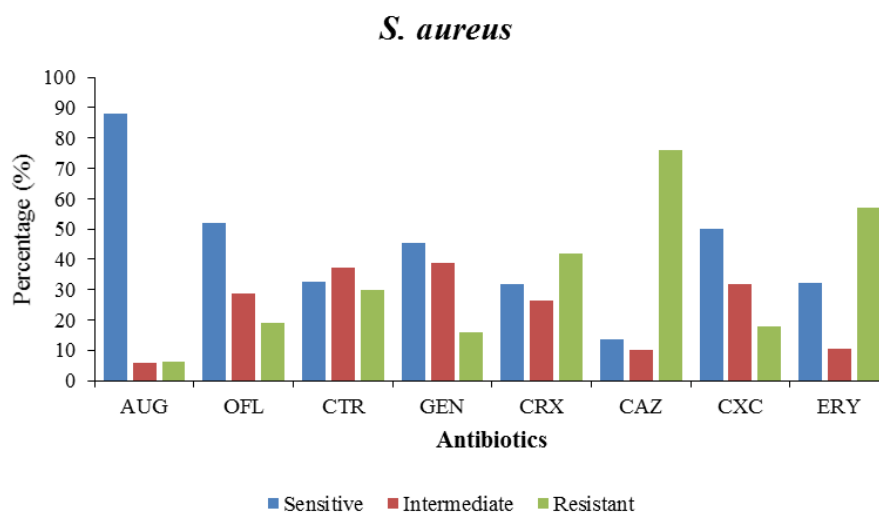


Figure 3 Antimicrobial susceptibility test for *Staphylococcus aureus*

AUG = Amoxicillin/Clavulanic acid, OFL= Ofloxacin, CTR= Ceftriaxone, GEN= Gentamicin, CRX= Cefuroxime, CAZ= Ceftazidime, CXC= Cloxacillin, ERY= Erythromycin

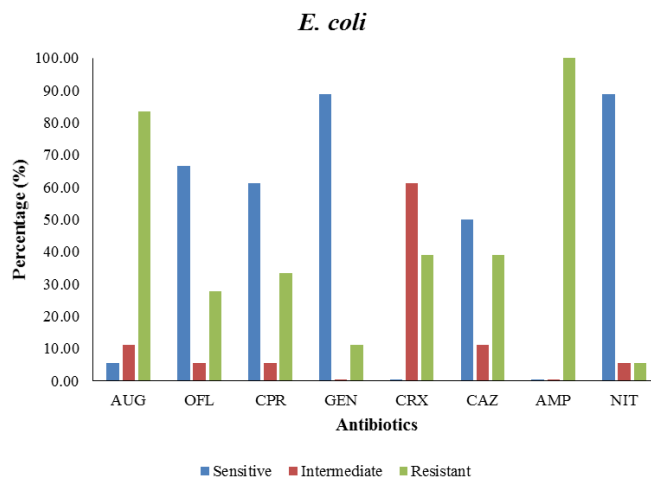


Figure 4 Antimicrobial susceptibility test for *Escherichia coli*

AUG = Amoxicillin/Clavulanic acid, OFL= Ofloxacin, CPR= Ciprofloxacin, GEN= Gentamicin, CRX= Cefuroxime, CAZ= Ceftazidime, AMP= Ampicillin, NIT= Nitrofurantoin

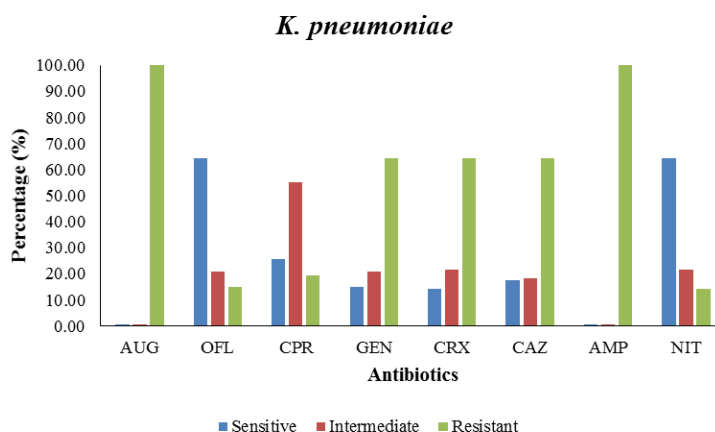


Figure 5 Antimicrobial susceptibility test for *Klebsiella pneumoniae*

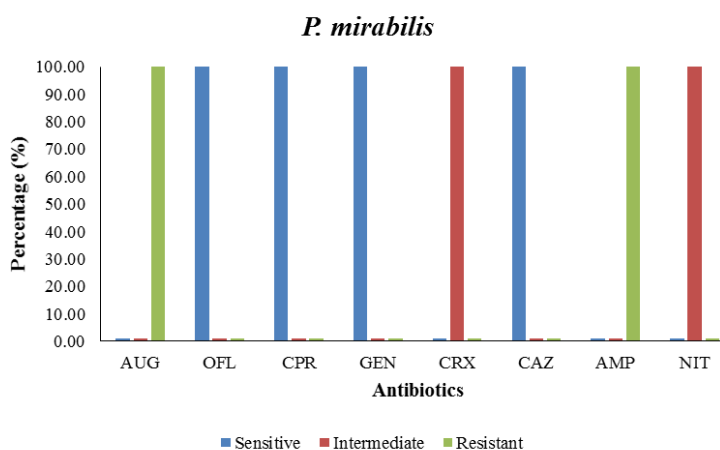


Figure 6 Antimicrobial susceptibility test for *Proteus mirabilis*

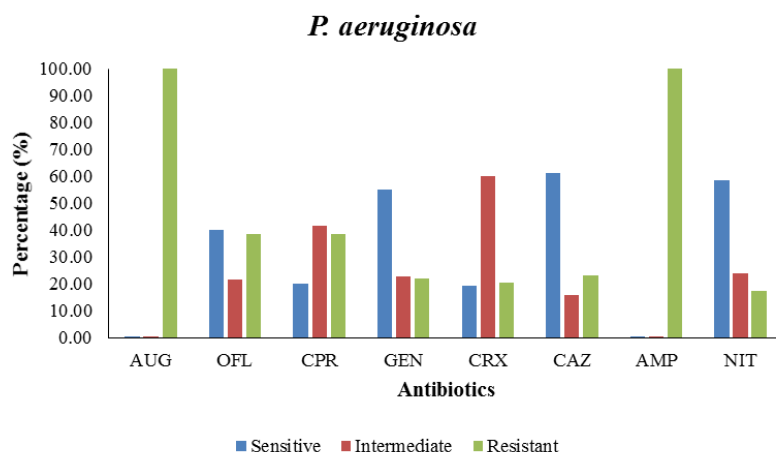


Figure 7 Antimicrobial susceptibility test for *Pseudomonas aeruginosa*

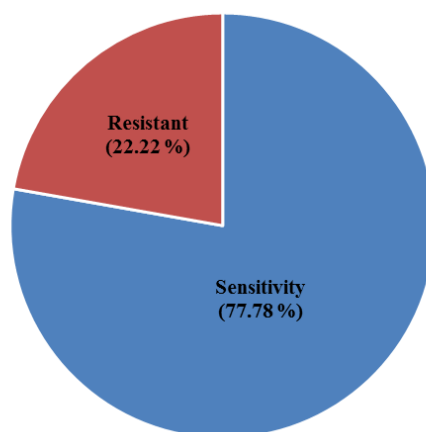


Figure 8 Susceptibility of *Candida albicans* to Ketoconazole (0.25 µg)

4. Discussion

4.1. Prevalence of commonly isolated urinary pathogens

This study presents the microbiological examination of urine samples of 150 outpatients that fall within the age group of 15 to above 66 years. The results of the study have shown the incidence of asymptomatic urinary tract infection in this population to be 40 %. The most predominant bacterial isolate from the urine samples is *Staphylococcus aureus* (47.19 %) followed by *Escherichia coli* (20.22 %), *Klebsiella pneumoniae* (15.73 %), *Pseudomonas aeruginosa* (5.62 %) and *Proteus mirabilis* (1.12 %). Fungal isolate as *Candida albicans* accounted for (10.11 %). This finding is similar to previous reports showing Gram-positive bacteria, particularly *S. aureus* as the most commonly implicated pathogen in patients with UTIs [33]. This could be as a result of its minimal growth requirements, ability to survive long in most unfavourable environments and to find a susceptible host. Akortha and Ibadin [33] previously noted the high incidence of *S. aureus* possibly due to the virulent nature of the organism, which gives it the ability to overcome body defense mechanisms and resistance to antibiotics. Earlier reports seem to suggest *S. aureus* as the most frequently isolated organism as well as the leading etiologic agent in urinary tract infection in our environment [34, 35]. It constituted as high as 28.09 % of cases in women suspected of UTI compared to men (19.1 %) and in agreement with the findings presented by Abdul and Onile [36] showing UTI caused by *S. aureus* as the most common among women in Ilorin, North-central, Nigeria. The high incidence of *S. aureus* in women could be due to the proximity between the genital tracts and the urethra/anus, which perhaps facilitate auto transmission as earlier reported [37].

Escherichia coli was recorded as the second prevalent isolate in this study, but fails to agree with previous report on uropathogens showing *E. coli* as the most frequently isolated organism in patients with UTIs [38, 39]. The high incidence of *E. coli* is attributed to the fact that it is a commensal of the bowel and infection is mostly through faecal contamination

occasioned by poor hygiene. Also, improper wiping after urination or defecation can result in transfer of organisms from the anus to the distal urethra. Previous studies on children hospitalized for non-infective urinary tract diseases shows *Klebsiella* species as the most frequently isolated organism, closely followed by *S. aureus* [40]. Samples obtained from students and commercial sex workers in Zaria, North-central, Nigeria showed prevalence of *P. aeruginosa* (53.4 %), *S. aureus* (43.3 %) and *E. coli* (40.7 %). As expected the commercial sex workers had the highest (30.6 %) prevalence of bacteria in their urine samples while the students had the least [41].

As shown in this study, *Candida albicans* was the fourth prevalent isolate in contrast to earlier report by Richards *et al.* [42] describing nosocomial infections in medical intensive care units. In their report, however, *C. albicans* was the most common single pathogen isolated from urine and these infections were associated with asymptomatic funguria rather than symptomatic urinary tract infections. Similar study on fungal urinary tract infection in patients at risk (such as severely ill patients, those with chronic renal failure, diabetes mellitus among others) Krcmery *et al.* [43] showed *C. albicans* as the most commonly isolated pathogen, followed by non-albicans *Candida* species (*Candida tropicalis*, *Candida krusei*) and then non-yeast *Candida* yeasts (*Blastoschizomyces capitatus*). The infection by this pathogen was reported to be associated with device use, fungal infections with urinary catheters.

4.2. Antimicrobial susceptibility

The knowledge of antibiotic susceptibility patterns of bacterial pathogens is essential to guide empirical and pathogen specific therapy [44]. As shown during the susceptibility testing, Amoxicillin/Clavulanate (88.1 %) was the most active against *S. aureus*, followed by the Fluoroquinolones in the following order Ofloxacin (52 %), Cloxacillin (50 %) and Gentamicin (45.2 %). The organism showed high resistance to Ceftazidime (76.2 %) and Erythromycin (57.14 %). Earlier studies in which similar antibiotics were used, indicates that *S. aureus* was sensitive to Amoxicillin/Clavulanate, Ofloxacin, Gentamicin, Cefuroxime, Erythromycin and resistant to Co-trimoxazole, Tetracycline and Nalidixic acid [33, 45]. Sensitivity patterns of *S. aureus* to antibiotics reported by Shittu and Mandere [46] showed slight contrast with a 100 % sensitivity to gentamicin and cephalosporin, but resistant to Amoxicillin/Clavulanate and Nitrofurantoin. These differences in sensitivity pattern of *S. aureus* could be attributed to environmental factors such as the misuse and abuse of antibiotics among the general population, which has favoured the emergence of resistance strains.

The susceptibility test of *E. coli* shows that Gentamicin and Nitrofurantoin (88.89 %) are active against the isolate, followed by Ofloxacin (66.67 %), Ciprofloxacin (61.11 %), Ceftazidime (50 %). The Nitrofurantoin demonstrated better activity against *E. coli* isolates probably because it is well absorbed orally and is rapidly excreted in the urine so that drug levels in urine are high while serum levels are minimal [47] *E. coli* was highly resistant to Ampicillin (100 %) and Amoxicillin/Clavulanate (83.33 %) and in agreement with previous studies [15, 48-51]. Besides resistance, the beta (β)-lactam antibiotics are known to have relatively poor action in treating symptomatic cystitis. The reason is that it is rapidly excreted and the duration of significant drug concentration in the urine is short. Also, they are relatively ineffective in clearing Gram-negative rods from the vaginal and colonic mucosa, thus possibly predisposing to recurrences when used to treat UTI. The susceptibility test of *K. pneumoniae* showed that Nitrofurantoin and Ofloxacin had same activity (64.29 %) against the isolates as opposed to Gentamicin, Cefuroxime and Ceftazidime. This is similar, to previous studies in which Nitrofurantoin showed high efficacy followed by Co-trimoxazole, Tetracycline, Norfloxacin, Cephalexin, Pefloxacin, Cefaclor, Nalidixic acid, Ciprofloxacin [52] but 100 % resistant to Amoxicillin and Ampicillin [29]. The low levels of resistance recorded on antibiotics such as ceftriaxone, nitrofurantoin and gentamycin seem to be as a result of accessibility and relatively high price compared to others. Thus, these drugs could be considered as alternative options in the empirical treatment of UTIs.

The susceptibility test of *P. aeruginosa* to the antibiotics used in this study showed that it was sensitive to Ceftazidime, Nitrofurantoin and Gentamicin. It however showed low susceptibility to the Fluoroquinolones, Ofloxacin (40 %) and Ciprofloxacin (20 %). Cefuroxime had an intermediate sensitivity of 60 % against *P. aeruginosa* while resistance was observed with Amoxicillin/Clavulanate and Ampicillin (100 %). This result seems to agree with previous studies by Inam *et al.* [16] in which isolates were found to be sensitive to Carbapenems, amikacin, ceftazidime and antipseudomonal penicillin such as piperacillin. The test involving *Proteus mirabilis* showed that the isolates were highly sensitive (100 %) to Ofloxacin, Ciprofloxacin, Gentamicin and Ceftazidime with intermediate sensitivity to Cefuroxime, Nitrofurantoin but resistant (100 %) to Amoxicillin/Clavulanate and Ampicillin. This is also in agreement with previous study in Sukkur city where Ofloxacin and Ciprofloxacin demonstrated effectiveness against *Escherichia* spp, *Klebsiella* spp and *Proteus* spp [18]. The high-level susceptibility to Ciprofloxacin might not be unconnected to its high cost. This high cost has probably restricted its procurement and misuse by the residents investigated, thereby reducing emergence of resistant bacterial strains [41]. Similarly, the reason for high resistance of bacteria isolates to some antibiotics such as Ampicillin, Amoxycillin would probably be an earlier exposure of the isolates to these drugs, which may have enhanced resistant development [53].

5. Conclusion

The study revealed *Staphylococcus aureus*, among other organisms isolated, as the leading cause of UTI in the study environment. Amoxicillin/Clavulanate, Ofloxacin or Cloxacillin can be the drug of choice in the treatment of UTI due to *Staphylococcus aureus* even though susceptibility to these antibiotics is more precise on individual basis. The isolates obtained from the urine samples in this study were resistant to Amoxicillin/Clavulanate, Ampicillin (Gram-negative organisms) and Ceftazidime and Erythromycin (Gram-positive organism). The fungal isolate present showed good susceptibility to Ketoconazole.

Compliance with ethical standards

Acknowledgments

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Disclosure of conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Statement of ethical approval

The study was approved by the research and ethics committee of University of Port Harcourt Teaching Hospital Rivers State, Nigeria.

Statement of informed consent

Informed consent was obtained from all individual participants included in the study.

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