

GSC Biological and Pharmaceutical Sciences

eISSN: 2581-3250 CODEN (USA): GBPSC2 Cross Ref DOI: 10.30574/gscbps

Journal homepage: https://gsconlinepress.com/journals/gscbps/



(RESEARCH ARTICLE)



Behavior of fluorescent *Pseudomonas* under different concentrations of heavy metals (Zn and Cu)

Oubeidillah Youssoufa Ali 1 , Adiouma Dangue $^{1, *}$, Demba Diaw 1 , Mame Arama Fall Ndiaye 1 and Tahir Abdoulaye Diop $^{1, 2}$

- ¹ Department of Plant Biology, Mushroom Biotechnology Laboratory, Faculty of Science and Technology, Cheikh Anta Diop University of Dakar, Dakar BP 5005, Senegal.
- ² Department Higher School of Agricultural and Food Sciences, Amadou Mahtar MBOW University of Dakar, Diamniadio, Senegal.

GSC Biological and Pharmaceutical Sciences, 2022, 18(01), 146-152

Publication history: Received on 26 December 2021; revised on 27 January 2022; accepted on 29 January 2022

Article DOI: https://doi.org/10.30574/gscbps.2022.18.1.0048

Abstract

The use of chemical fertilizers has resulted in the depletion of soils in mineral elements and telluric microorganisms. Thus, for an improvement in crop growth, the exploitation of telluric microorganisms such as *Pseudomonas* can be an alternative.

Laboratory work has made it possible to isolate strains of *Pseudomonas* from the rhizospheres of tomato, eggplant and onion. These strains were also tested in the presence of different concentrations of certain metals ($CuSO_4$ and $ZnSO_4$ (0, 1, 3, 4, 5 and 6 mmol/l).

The tests carried out in the laboratory in the presence of Cu and Zn revealed a very significant reduction in the number of colonies of the strains of *Pseudomonas* for the concentrations of 5 and 6 mmol/l.

The results showed better zinc tolerance for both tomato and onion rhizosphere strains. In the presence of Cu, the strain isolated from tomato rhizosphere shows more tolerance to heavy metals. These results made it possible to isolate different strains of *Pseudomonas* and to specify their tolerance thresholds for heavy metals.

Keywords: *Pseudomonas*; Heavy metals; Tomato; Onion; Eggplant

1. Introduction

Man, through strong industrialization and intensive agriculture using fertilizers and pesticides, contributes to the dissemination of metallic trace elements and their accumulation in the soil ([1]; [2]; [3]; [4]). Several of these metals such as iron, copper and zinc are essential for the proper growth of living organisms, especially plants and microorganisms [5]. But these metals can also be limiting factors for these organisms [6]. Copper and zinc are not dangerous for the organisms in themselves which is not the case for other metals such as lead or cadmium, but at an excessive level, they become toxic although the proper functioning of the metabolism of the organisms especially bacteria, is also dependent on these two metals ([7]; [8]; [9]). Although the effects of microorganisms on the availability of metal contaminants in soils are relatively well documented, the majority of these studies are based on a geochemical and not a biological approach [10]. It should be noted that in addition to fungi and lichens, bacteria play a major role in the phenomena of alteration of heavy metals. However, among the microorganisms present in the soil, there are bacteria

Department of Plant Biology, Mushroom Biotechnology Laboratory, Faculty of Science and Technology, Cheikh Anta Diop University of Dakar, Dakar BP 5005, Senegal.

^{*} Corresponding author: Adiouma Dangue

of agricultural interest ([11]; [12]) referred to by the acronym PGPR (Plant Growth Promoting Growth) in this case Pseudomonas fluorescents ([13]; [14]). They are strict aerobic Gram-negative bacilli [15]. These bacteria are motile thanks to one or more polar flagella. They get the second part of their name from the fact that they are fluorescent ([16]; [17]). This fluorescence is due to the production of fluorescein (or pyoverdine), which is a yellow-green pigment that fluoresces under ultraviolet light [18]. They are ubiquitous bacteria, particularly abundant in soils and waters [19]. Their ability to colonize roots and maintain a high population density there is therefore remarkable [20]. They have several intrinsic characteristics that make them interesting, in particular their usefulness as biological control agents thanks to the solubilization of phosphorus, phosphate and the sequestration of iron by siderophores as well as the production of antibiotics ([14]; [21]). Although siderophores are defined as specific iron chelators, they are also able to effectively complex metal cations other than iron [22]. This great competence comes from their higher growth rate at the rhizosphere level [23]. Added to this is their ability to use a very wide range of substrates, often derived from root exudates, as a source of nitrogen or carbon [10]. They are very easy to isolate and cultivate in the laboratory and easily lend themselves to genetic manipulation [24]. It is interesting to underline that although metallic trace elements are mobilized by various organisms, they cannot be degraded into less toxic products. They persist indefinitely in the environment and pose a major problem in ecology but also for public health due to their carcinogenic and mutagenic properties [25]. The toxicity and persistence of metallic trace elements in the environment require the development of different methods to reduce contamination [3]. The objective of this work is to first study the behavior of fluorescent Pseudomonas isolated from the rhizosphere of tomato (To), eggplant (Au) and onion (Oi) and then to assess their tolerance to different concentrations of heavy metals Cu and Zn.

2. Material and methods

2.1. Isolation of fluorescent Pseudomonas

Soil samples were taken at the Center for the Development of Horticulture (CDH Cambérène station, Niayes eco-geographical zone, Dakar Region, Senegal) during the off-season (February).

These are soils from the rhizosphere of tomato, onion and eggplant plants in the fruiting stage and on bare soil. For each soil sample, 500 g were taken from a depth of 0 to 25 cm. From these samples, isolations of fluorescent *Pseudomonas* spp were carried out by the suspension-dilution method. A sample of 1 g of each soil was suspended in 10 ml of sterile 0.1 M of MgSO₄,7H₂O. After stirring for 10 minutes using a vortex, decimal dilutions $(10^{-1} \text{ to } 10^{-8})$ were made from this suspension. 0.1 ml of each suspension was plated in Petri dishes each containing 10 ml of King B agar medium [26]. Petri dishes are incubated at 28°C in the dark.

After 48 hours of incubation, the bacterial colonies were observed under a UV lamp at 365 nm.

For each soil sample, three fluorescent pigment-producing isolates were selected and purified by streaking in three dishes containing King B's solid medium. Each isolate is designated by a code. These are the To, Oi and Au strains isolated respectively from the rhizosphere of tomato, onion and eggplant plants.

2.2. Effect of heavy metals on fluorescent Pseudomonas spp.

To study the effect of these heavy metals on fluorescent Pseudomonas, liquid KB media were prepared with different Zn and Cu concentrations: 0, 1, 3, 4, 5 and 6 mmol/l. These media were autoclaved at 120°C for 20 minutes. Then, aliquots of 100 μ l of bacterial suspension of each of the Pseudomonas strains were spread using a micropipette in Petri dishes each containing 10 ml of the medium. The Petri dishes were incubated in an oven at a temperature of 28°C for 72 h.

3. Results

3.1. Characteristics of fluorescent colonies on U.V

After 48 h of culture, the bacterial colonies were observed on a UV table at 365 nm. For each speculation (tomato, onion and eggplant as well as bare soil), the number of fluorescent colonies varies between 3 and 4. The fluorescence was clear and sharp (Figure 1). And it was the 10^{-2} and 10^{-3} dilutions that showed the best results regarding pigmentation intensity, size, elevation as well as color and shape. No fluorescent colonies were observed with bare soil. These colonies were then subcultured on KB medium for purification (Figure 2).

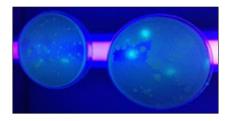


Figure 1 Colonies of fluorescent Pseudomonas under U.V.

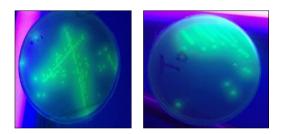


Figure 2 Colonies of fluorescent Pseudomonas after purification

3.2. Macroscopic aspects of fluorescent *Pseudomonas* strains

The strains showed slightly variable characteristics. (Table 1).

Table 1 Characteristics of fluorescent Pseudomonas strains observed under UV after 48 hours

Strains	Form	Size (mm)	Elevation	Color	Intensity of pigmentation
Au	Round	4.5	Domed	Yellow-green	+
То	Round	5	Domed	Green	+
Oi	Round	4	Flat	Green	+

3.3. Effect of Different ZnSO₄ Concentrations on Fluorescent Pseudomonas

Fluorescent *Pseudomonas* from the rhizosphere of tomato, eggplant and onion grown on KB medium for 72 hours showed variability in the number of colonies depending on ZnSO₄ concentration and speculation (Figure 3).

In the absence of ZnSO₄, the number of fluorescent *Pseudomonas* colonies is greater for the strains isolated from the eggplant rhizosphere (340 colonies). This value shows a significant difference in the number of colonies of onion rhizosphere compared to that of tomato rhizosphere (301 colonies).

For a ZnSO₄ concentration of 1 mmol/l, the number of colonies of strains isolated from the eggplant rhizosphere (214 colonies) decreased significantly compared to the number of colonies from the rhizospheres of tomato (324 colonies) and onion (276 settlements).

At a $ZnSO_4$ concentration of 3 mmol/l, the number of colonies of the strains isolated from the rhizosphere of eggplant and tomato are respectively 301 and 280 colonies. These values are twice as large compared to the number of colonies in the onion rhizosphere (124 colonies).

When the ZnSO₄ concentration is 4 mmol/l, the numbers of fluorescent *Pseudomonas* colonies in the onion and tomato rhizosphere are five times greater than the number of colonies isolated from the eggplant rhizosphere.

For a ZnSO₄ concentration of 5 mmol/l, it can be seen that the number of colonies of fluorescent *Pseudomonas* isolated from the eggplant rhizosphere (65 colonies) has significantly decreased compared to that of tomato (137 colonies).

For all the strains, the lowest values are obtained with a concentration of 6 mmol/l. The number of colonies of the different speculations did not vary significantly at this concentration (Figure 3).

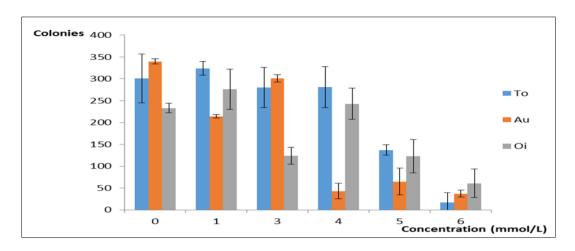


Figure 3 Effect of different ZnSO₄ concentrations on fluorescent *Pseudomonas* colonies

3.4. Effect of Different CuSO₄ Concentrations on Fluorescent *Pseudomonas*

Fluorescent *Pseudomonas* from the rhizosphere of tomato, eggplant and onion, cultured on KB medium for 72 hours showed a variability in the number of colonies depending on the concentration of CuSO₄ and speculation (Figure 4).

In the absence of CuSO₄ (control), there is no significant difference between the number of colonies from the tomato rhizosphere (301 colonies) and that of the eggplant rhizosphere (313 colonies). However, these values show a significant difference compared to the number of colonies obtained at the level of the rhizosphere of the onion.

For the CuSO₄ concentration of 1 mmo/l, the colonies of fluorescent *Pseudomonas* from eggplant (23) and onion (14) are very weak or even negligible compared to those from the tomato rhizosphere (335).

At 3 mmol/l of CuSO₄, the number of colonies of fluorescent *Pseudomonas* from the tomato rhizosphere (124) is significantly higher compared to those from the eggplant (68) and onion (49) rhizospheres.

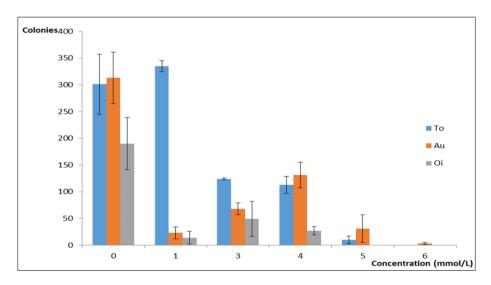


Figure 4 Effect of different concentrations of CuSO₄ on colonies of fluorescent *Pseudomonas*

For a CuSO₄ concentration equal to 4 mmol/l, colonies of fluorescent *Pseudomonas* from the rhizosphere of tomato (113) and eggplant (131) did not show any significant difference. However, we find a significant decrease in the number of colonies of fluorescent *Pseudomonas* originating from the onion rhizosphere (27).

At 5 mmol/l of CuSO₄, the numbers of colonies of fluorescent *Pseudomonas* originating from the rhizospheres of tomato (10) and eggplant (31) fell sharply and did not show any significant variation, on the other hand a total absence of colonies originating from the onion rhizosphere.

At 6 mmol/l of CuSO₄, only the strains isolated from the eggplant rhizosphere were recorded with a very low value (3 colonies) (Figure 4).

4. Discussion

Cultures in the presence of heavy metals may require the presence of microorganisms to ensure their growth. Thus, soil microorganisms provide a physical and chemical link between soil and plant roots. They participate in the recycling of plant nutrients and in maintaining soil stability as well as in the detoxification of toxic chemicals ([27]; [28]). On a larger scale, in the environment, essential or non-essential metals cause considerable modifications within the microbial community and their activities. Furthermore, *Pseudomonas* have developed mechanisms to deal with toxic metals [29]. With the strains of *Pseudomonas* isolated in the rhizosphere of tomato, eggplant and onion, we observe a very significant decrease in the number of colonies for concentrations of 5 and 6 mmol/l. These results are in agreement with those of Chen et *al.*, [29] who showed that from concentrations of 3 and 5 mmol/l, there is a significant decrease in the number of colonies of *Pseudomonas* respectively in the presence of Cu, and Zn. However, these microorganisms were related to *Pseudomonas* Putida based on 16S ribosomal DNA sequencing. For these levels of Cu and Zn, the numbers of colonies observed were higher for tomato and onion if it is Zn. In the presence of Cu, the strain of *Pseudomonas* from tomato rhizosphere was found to be more tolerant. The effect of heavy metals on the growth of colonies of *Pseudomonas* shows that the intensity of the effect of metallic trace elements varies according to the level and the nature of the metal and could also vary according to the cultivated speculation ([30]; [10]).

The results show better zinc tolerance for strains isolated from tomato and onion rhizosphere (Figure 3). In the presence of Cu, the strain isolated from the tomato rhizosphere shows better tolerance (Figure 4). Additionally, we found that the toxicity of copper was greater than that of zinc.

The effect of heavy metals on ecosystems and the mechanisms of their transfer from soil to living organisms are still poorly understood [31].

5. Conclusion

Tomato, onion and eggplant are three crops widely used by the populations of the Sahel where they play an important economic and socio-economic role. CDH soils showed a significant potential for rhizospheric microorganisms. However, this study showed that the different strains of *Pseudomonas* did not have the same level of tolerance to heavy metals. Our work has made it possible to define, under in vitro conditions, the tolerance thresholds for heavy metals (Zn and Cu) for the strains of fluorescent *Pseudomonas* isolated in the rhizosphere of the different speculations used.

Indeed, the results obtained showed that the colonies of *Pseudomonas* from the rhizosphere of tomato and onion were more tolerant to ZnSO₄ (5 mmol/l) compared to the strains isolated from the rhizosphere of eggplant which are more sensitive to ZnSO₄. The effect of ZnSO₄ from a concentration of 4 mmol/l. For Cu, colonies of *Pseudomonas* in the tomato rhizosphere are more tolerant to CuSO₄ (4 mmol/l) followed by those of eggplant and finally those of onion. Overall, it appears from this study that strains of *Pseudomonas* from the tomato rhizosphere are more tolerant to different abiotic constraints, namely the presence of heavy metals (Zn and Cu).

Compliance with ethical standards

Acknowledgments

The authors the Mushroom Biotechnology Laboratory.

Disclosure of conflict of interest

Authors declare that no conflict of interest exist.

References

[1] Baize D. Teneurs totales en « métaux lourds » dans les sols français: résultats généraux du programme ASPITET. Le Courrier de l'Environnement de l'INRA. 2000; (40): 39-54.

- [2] Adam S, Edorh PA, Totin H, Koumolou L, Amoussou E, Aklikokou K, Boko M. Pesticides et métaux lourds dans l'eau de boisson, les sols et les sédiments de la ceinture cotonnière de Gogounou, Kandi et Banikoara (Bénin). International Journal of Biological and Chemical Sciences. 2010; 4(4).
- [3] Nouri M, Haddioui A. Les techniques de dépollution des sols contaminés par les métaux lourds: une revue (The remediation techniques of heavy metals contaminated soils: a review). Maghrebian Journal of Pure and Applied Science. 2016; 2(2): 47-58.
- [4] Diaw D, Fall-Ndiaye MA, Ali OY, Sare IC, Diop TA. Effet de la salinité sur la densité des isolats de *Pseudomonas* spp fluorescents de rhizosphère de plants de tomate, d'aubergine et d'oignon au Sénégal. International Journal of Biological and Chemical Sciences. 2018; 12(4): 1914-1919.
- [5] Lurthy T. Interactions Pisum sativum–*Pseudomonas*: conséquences sur la nutrition en fer, la croissance et l'immunité de la plante hôte. Biodiversité et Ecologie. Université Bourgogne Franche- Comté, 2020. Français. NNT. 2020; 294.
- [6] Tou-dalill I. Bioremédiation du Cd, Zn et Cu, par deux bactéries rhizosphériques. Université des Sciences et de la Technologie Houari Boumediene. 2013; 72.
- [7] Wintz H, Fox T, Vulpe C. Reponses of plants to iron, zinc and copper deficiencies. Biochemical Society Transactions. 2002; 30(4): 766-768.
- [8] Cloutier-Hurteau B. Rôle des microorganismes sur la spéciation du Cu, Zn et Al dans la rhizosphère de sols forestiers. Université de Montréal. 2009; 234.
- [9] El Idrissi L. Cytotoxicité du cadmium, du plomb et du mercure et caractérisation du transport membranaire de cadmium dans les cellules alvéolaires (A549) et bronchiolaires (H441), Université du Québec à Montréal, Canada. 2009; 88.
- [10] Ferret C. Role of fluorescent *Pseudomonas* in the biodisponibility of metals contamining the soil minerals. Université de Strasbourg. 2012; 266.
- [11] Deshwal VK, Kumar P. Effect of salinity on growth and PGPR activity of *Pseudomonas*. Journal of Academia and Industrial Research. 2013; 2(6): 353-356.
- [12] Khan N, Bano A, Rahman MA, Guo J, Kang Z, Babar MA. Comparative physiological and metabolic analysis reveals a complex mechanism involved in drought tolerance in chickpea (*Cicer arietinum* L.) induced by PGPR and PGRs. Scientific reports. 2019; 9(1): 1-19.
- [13] Lemanceau P. Effets bénéfiques de rhizobactéries sur les plantes : exemple des *Pseudomonas* spp fluorescents. Agronomie. 1992; 12(6): 413-437.
- [14] David BV, Chandrasehar G, Selvam PN. *Pseudomonas fluorescens*: a plant-growth-promoting rhizobacterium (PGPR) with potential role in biocontrol of pests of crops. In Crop improvement through microbial biotechnology, Elsevier. 2018; 221-243.
- [15] Smits TH, Balada SB, Witholt B, Van Beilen JB. Functional analysis of alkane hydroxylases from gram-negative and gram-positive bacteria. Journal of bacteriology. 2002; 184(6): 1733-1742.
- [16] Haahtela K, Helander I, Nurmiaho-Lassila EL, Sundman V. Morphological and physiological characteristics and lipopolysaccharide composition of N2-fixing (C2H2-reducing) root-associated *Pseudomonas* sp. Canadian journal of microbiology. 1983; 29(8): 874-880.
- [17] Lakhdari MB, Bennecer MZE. Etude de la résistance aux antibiotiques chez *Pseudomonas aeruginosa*. Université Larbi Ben M'hidi Oum El Bouaghi, Algérie. 2021; 43.
- [18] Meyer JM. Pyoverdines: pigments, siderophores and potential taxonomic markers of fluorescent *Pseudomonas* species. Archives of microbiology. 2000; 174(3): 135-142.
- [19] Oulebsir-Mohandkaci H, Tihar-Benzina F, Belkacem CA, Belgrade AN. Recherche de molécules bioactives d'intérêt à partir d'une collection de souches bactériennes rhizosphèriques et étude de leur effet antifongique. Algerian Journal of Environmental Science and Technology. 2020; 6(3).
- [20] Shameer S, Prasad TNVKV. Plant growth promoting rhizobacteria for sustainable agricultural practices with special reference to biotic and abiotic stresses. Plant Growth Regulation. 2018; 84(3): 603-615.

- [21] Farida BT, Hakima OMK, Sonia H, Fatma SH. Détermination et caractérisation des sidérophores synthétisés par quelques souches de *Pseudomonas* spp. fluorescents phytobénéfiques. Revue Nature et Technologie. 2021; 12(3): 17-30.
- [22] David S. Altération de déchets amiantés par des bactéries et des sidérophores en vue du développement d'un procédé de bioremédiation. [Doctoral dissertation, Strasbourg], Université de Strasbourg. 2019.
- [23] Samia MM. Localisation des déterminants de la suppression de quelques souches de *Pseudomonas* isolées de la rhizosphère de la pomme de terre. Thèse de doctorat en sciences, Université Ferhat Abbas de Sétif 1, Algérie. 2012; 189.
- [24] Rabhi N. Isolement de *Pseudomonas* spp. fluorescents d'un sol salé: Effet d'osmoprotecteurs naturels, Université Ferhat Abbas Sétif, Algérie. 2011; 110.
- [25] Boutamine S, Boumelta R. Impact des décharges sauvages sur les sols et les méthodes de remédiation, Université Med-Seddik Benyahia-Jijel. 2019; 45.
- [26] King EO, Ward MK, Raney DE. Two simple media for the demonstration of pyocyanin and fluorescin. J Lab Clin Med. 1954; 44: 301-7.
- [27] Vivas A, Azcón R, Biró B, Barea JM, Ruiz-Lozano JM. Influence of bacterial strains isolated from lead-polluted soil and their interactions with arbuscular mycorrhizae on the growth of *Trifolium pratense* L. under lead toxicity. Canadian journal of microbiology. 2003; 49(10): 577-588.
- [28] Stepanauskas R, Glenn TC, Jagoe CH, Tuckfield RC, Lindell AH, King CJ, McArthur JV. Coselection for microbial resistance to metals and antibiotics in freshwater microcosms. Environmental Microbiology. 2006; 8(9): 1510-1514.
- [29] Chen C, Belanger RR, Benhamou N, Paulitz TC. Defense enzymes induced in cucumber roots by treatment with plant growth-promoting rhizobacteria (PGPR) and *Pythium aphanidermatum*. Physiological and Molecular Plant Pathology. 2006; 56(1): 13-23.
- [30] Vogel-Mikuš K, Pongrac P, Kump P, Nečemer M, Regvar M. Colonisation of a Zn, Cd and Pb hyperaccumulator *Thlaspi praecox* Wulfen with indigenous arbuscular mycorrhizal fungal mixture induces changes in heavy metal and nutrient uptake. Environmental Pollution. 2006; 139(2): 362-371.
- [31] Ndiaye M, Cavali E, Diop TA. Growth of mycorhized seedlings of a tropical gum tree in coper contaminated soil. 2012.